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#### Prevalence of Bovine Trypanomosis and Associated Risk Factors in Pawi special District of Benishangul Gumuz region, North Western Ethiopia.

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Abstract: A cross sectional study was carried out in pawe special district of Benishangul Gumuz Regional State, North West Ethiopia from January to March 2019 to determine the prevalence of trypanosomosis, prevailing species of trypanosomes, associated risks and its vector density. Blood samples collected from 384 randomly sampled cattle (Bos indicus) was examined using parasitological (buffy coat technique) and haematological (Measurement of packed cell volume) procedures. An overall, 10.94% (42/384) prevalence of trypanosomosis was recorded. The infection was caused mainly by Trypanosoma congolense 31/42 (73.8%), Trypanosoma vivax 11/42 (26.8%), and the infection rate was statistically significant among different trypanosome species (P < 0.05). Mean packed cell volume (PCV) value of infected animals was lower  $(19.4 \pm 1.3)$  than non-infected animals  $(22.03 \pm 1.73)$  and the variation was found statistically significant (P<0.05). Similarly, higher prevalence (17.8%) of trypanosomosis infection was registered in animals with poor body condition when compared to animals with medium (13.46%) and good (4.76%) body condition and the difference was statistically significant (P<0.05). In contrast, prevalence of trypanosomosis was not statistically significant among study sites, age categories and sex groups of study animals (P > 0.05). Glossina morsitans submorsitans, and Glossina tachinoedes were the tsetse fly species caught and their mean apparent density measured as flies/trap/day was 0.78. In addition, other mechanical vectors such as stomoxy and tabanids were captured with flies/trap/day 0.12 and 0.8, respectively. To wrap up, the result of the current finding reveals moderately high prevalence of trypanosomosis in the study district signaling the need for strategic and participatory approach to control the vector and to minimize the impact of the disease in the study district. [Mohammed Abdulkadir, Mubarik Kedir, Haile Worku and Kedir Yemer. Prevalence of Bovine Trypanomosis and Associated Risk Factors in Pawi special District of Benishangul Gumuz region, North Western Ethiopia. Researcher 2021;13(1):34-40]. ISSN 1553-9865 (print); ISSN 2163-8950 (online). http://www.sciencepub.net/researcher. 6. doi:10.7537/marsrsj130121.06.

Key words: Pawi, Trypanosomosis, Tsetse fly, prevalence, Risk factors.

#### 1. Introduction

Trypanosomes are extracellular protozoan parasites that cause debilitating diseases called trypanosomosis in animals and sleeping sickness in humans and have great socio-economic impact adversely affecting food production and economic growth in many parts of Africa, particularly in Sub-Saharan Africa (SSA), (Taylor, 2015). The disease in animal is transmitted cyclically by the genus Glossina, but it can also be transmitted mechanically by other biting flies among which Tabanus and Stomoxys are presumed to be the most important, as exemplified by their presence in South and Central America as well as in some areas of Africa free or cleared of Glossina such as Ethiopia, Chad, Senegal, Sudan etc (Truc et al., 2013).

The distribution of the disease coincides with the habitat of the tsetse fly vector and is called the tsetse

fly "belt" or it is sometimes referred to as "green desert" because  $\sim 10$  million km<sup>2</sup> of potential fertile land is rendered to be unsuitable for cultivation (Shaw *et al.*, 2014). Within this area, the majority of the *Glossina* infested countries are underdeveloped, poor, heavily indebted and food-deficit due to lack of productive animals as far as meat/milk production and draft power are concerned, resulting in an annual economic loss of about 5 billion US\$ (Giordani *et al.*, 2016).

In Ethiopia, the most important trypanosome spp affecting livestock include *T. congolense*, *T. vivax* and *T. brucei* in cattle, sheep and goats; *T. evansi* in camels and *T. equiperdium* in horses (Abebe, 2005). Western and southern river basins of Ethiopia are the most severely affected areas by trypanosomosis in the country. In area specifically in the western part, a wide diversity of *Glossina* and trypanosome spp and strains co-exist (Abebe, 2005). Therefore, the current study was conduct to assess the prevalence of trypanomosis; to determine distribution and apparent density of tsetse fly in the study area and to recommend possible control and prevention measures.

#### 2. Materials and Methods

### 2.1 Study area

The study was conducted from January to March, 2019 in Pawe special district of Metekel zone, Benishangul Gumuz Regional State. The study was conducted in five villages namely: Almu, Mender-14, Mender-17, Mender-5 and Mender-7. The district has 20 kebeles covering an area of 64,300 hectare with human population of 42,000. The study area is located at latitude of 11° and 15° 24.7"N and, longitude of 360 and 23'10"E. It has an altitude of 1064m above sea level. The mean annual average temperature of the district is 32<sup>°c</sup> and the mean annual rainfall range from 900-1400 mm (NMSA, 2007). The livelihood of the society largely depends on mixed livestock and crop production having livestock population of 58,810 Cattle, 5440 Goat, 5523 Sheep, 843 Equines, 29378 Poultry (CSA, 2015). The district lies at Dabus and Beles rivers water shed system which is one of the tributary of Nile River.

## 2.2. Study Design and Study Animals

Cross sectional study design was used. A local zebu cattle (Bos indicus), which are usually kept under an extensive husbandry system grazing the communally owned pasture land throughout the year were randomly sampled. They grazed together during the day time and returned to their individual owner's farmstead each evening. The body condition of each of the study cattle was scored as good, medium and poor (Nicholson and Butterworth, 1986). Concurrently, their age was categorized in years ((< 2, 2-5 and > 5) based on De-Lahunta and Habel (1986) principles.

# 2.3. Sampling Techniques and Sample Size Determination

The study sites were selected purposively as convenient. The animals were sampled randomly involving both sexes, all age groups, and all types of body conditions. The desired sampling size was calculated according to the formula given by Thrusfield, (2007). The sample size was determined based on the expected prevalence of 50%; confidence level of 95%, and 5% desired absolute precision. As result a total of 384 cattle were calculated and cattle were sampled at their communal grazing area using simple random sampling method.

## 2.4. Study Methodology

#### 2.4.1. Packed cell volume (PCV) determination

Blood samples were obtained by puncturing the marginal ear vein with a lancet and collected directly into a pair of heparinized capillary tubes. The tubes were then sealed at one end with crystal seal. PCV was measured in a micro-haematocrit centrifuge (Hermmle Labortechnik, type Z, Germany). The capillary tubes were placed in microhaematocrit centrifuge with sealed end outermost. Then the tube was loaded symmetrically to ensure good balance. After screwing the rotary cover and closing the centrifuge lid, the specimens were allowed to centrifuge at 12,000 rpm for 5 minutes. After centrifugation, the capillary tubes were placed in a haematocrit reader. The length of the packed red blood cells column is expressed as a percentage of the total volume of blood. PCV was read and recorded for each sample Animals with PCV less than 24% were considered to be anemic (OIE, 2008)

### 2.4.2. Buffy coat technique

Heparinized microhaematocrit capillary tubes containing blood samples were centrifuged for 5 minutes at 12,000 rpm. After the centrifugation, trypanosomes were usually found in or just above the buffy coat layer. The capillary tube was cut using a diamond tipped pen 1 mm below the buffy coat to include the upper most layers of the red blood cells and 3 mm above to include the plasma. The content of the capillary tube was expressed onto a glass slide, and covered with cover slip. The slide was examined under x40 objective and x10 eye piece for movement of parasite. Trypanosome species were identified according to their morphological descriptions as well as movement in wet film preparations (OIE, 2008).

## 2.4.3. Entomological Survey

To assess the apparent densities, distributions and species of tsetse flies and other biting flies the entomological data were collected in late rain season (May to Jun 2019) along the suitable tsetse habitats. These include livestock grazing areas, watering points, wild game reserve areas, rivers, savanna grass land and area of dense river side forests in the Districts. Monopyramidal traps baited with acetone, octanol and cow urine Taylor., *et al.*, (2007), were deployed at an interval of about 100 meters along watershed and vegetation to assess the fly density. The coordinates of each trap position were recorded with a Global Positioning System (GPS).

In all study sites a total of 50 Monoconical traps were deployed early in the morning and maintained in position for 48 hrs at five different peasant associations (PAs) in 5 trapping sites. The underneath of each trap pole was smeared with grease in order to prevent the ants climbing up the pole towards the collecting cage that could damage the tsetse flies and the cages from these traps were emptied to caught tsetse flies and other biting flies were counted, recorded and identified and sexed for the tsetse fly, other biting flies according to their morphological characteristics such as size, color, wing venation structure and proboscis at the genus level Taylor., *et al.*, (2007). Sexing was done for tsetse fly just by observing the posterior end of the ventral aspect of abdomen by hand lens, as a result male flies easily identified by enlarged hypophageum.

#### 2.5. Data Analysis

All the collected raw data and the results of parasitological and hematological examination data were entered into a Microsoft excel spread sheets program and then was transferred to Intercool STATA version 10.0 for analysis. The prevalence of trypanosome infection was calculated as the number of positive animals as examined by buffy coat method divided by the total number of animals examined at the particular time. Data collected on PCV values were analyzed by ANOVA to compare the mean PCV values of infected animals against that of uninfected animals. Pearson's chi-square ( $\chi 2$ ) was used to evaluate the association of different variables with the

prevalence of trypanosome infection. In all of the statistical analysis, a confidence level of 95% is used and P-value of less than 0.05 (at 5% level of significance) was considered as statistically significant.

#### 3. Results

## **3.1.** Prevalence of Trypanosome species in Pawi District

Out of total animals examined, 10.94% (42/384), were infected with trypanosomes. The prevalence in terms of trypanosome species was 73.8% T. congolense and 26.8 % T. vivax. The proportion of trypanosome species was 31/42 (73.8%) T. congolense and 11/42 (26.2%) T. vivax. The maximum and minimum Trypanosome infection rate in examined peasant association of pawe district was 17 % and 5 %recorded at mender 17 and mender 5 peasant associations (Table 1). The infection rate difference between trypanosomes was statistically significant (P<0.0001).

|          | Table 1. Trevalence of Trypanosonic by peasant association |                 |             |               |                     |         |  |  |
|----------|--|-----------------|-------------|---------------|---------------------|---------|--|--|
| No       | PA's   | No-Sample taken | No-positive | Prevalence %  | Trypanasome species |         |  |  |
| INU      |  |                 | No-positive | Trevalence /0 | T.congo             | T.vivax |  |  |
| 1        | Mender 5   | 80              | 4           | 5 %           | 3                   | 1       |  |  |
| 2        | Mender 14  | 100             | 17          | 17 %          | 11                  | 6       |  |  |
| 3        | Mender 17  | 80              | 12          | 15 %          | 10                  | 2       |  |  |
| 4        | Almu   | 44              | 3           | 6.8 %         | 2                   | 1       |  |  |
| <u>5</u> | Mender 7   | 80              | 6           | 7.5 %         | 5                   | 1       |  |  |
|          | Total  | 384             | 42          | 10.94%        | 31                  | 11      |  |  |

Table 1: Prevalence of Trypanosome by peasant association

#### Table 2: Prevalence of Trypanosomosis by species of the parasites.

| Trypanosome     No positive     Prevalence % |    |       |  |  |  |  |
|--|----|-------|--|--|--|--|
| T.congolense                                 | 31 | 73.8% |  |  |  |  |
| T.vivax                                      | 11 | 26.2% |  |  |  |  |

## 3.2. Cattle PCV Distribution and Anemia in Studied Area

The mean PCV value for whole examined animals was  $25.5 \pm 1.73$  SD. However, the mean PCV value for uninfected animals was  $22.03 \pm 1.73$  SD and mean PCV value of the infected animals was  $19.4 \pm$ 1.3 SD. The mean PCV values of cattle were significantly (= 0.0011) influenced by trypanosome infection as 22.79% and 25.81% PCV values in trypanosome positive and trypanosome negative animals were registered, respectively. The overall anemia prevalence in the studied district was 49.5% (190/384). The anemia prevalence was significantly higher in trypanosome infected cattle (56.8%) than in non-infected cattle (43.2%) (<0.05). Out of 49.5% anemia prevalence, 10.94% (42/384) was trypanosome infected animals. However, large number of animals 38.5% (148/384) had anemia (PCV < 24) without having trypanosome infection. Some animals 3.6% (14/384) were infected by trypanosome but their PCV was found normal.

| Table 3: Prevalence of Trypanosomosis a | according to different risk factors |
|---|-------------------------------------|
|---|-------------------------------------|

| Status       | frequency | Mean Pcv | SE   | Overall pcv | $X^2$    | p-value |
|--------------|-----------|----------|------|-------------|----------|---------|
| Infected     | 42        | 19.4     | 1.3  | 824         | 194.2868 | < 0.001 |
| Non infected | 348       | 22.03    | 1.52 | 8462        |          |         |
| Total        | 384       | 25.5     | 1.73 | 9784        |          |         |

The highest trypanosomosis prevalence 218 (12.8%) was recorded in >5 years old animals whilst the lowest prevalence 48 (4.34%) was in animals < 2 years old. Slightly higher prevalence was registered in females 24 (11.32%) than in males 18 (10.5%). Trypanosomosis was recorded across the study sites with the highest (17%) prevalence in Mender-14 and the lowest (5% in Mender-5). Trypanosomosis

prevalence was statistically non-significant between age categories, sex groups and across study sites. The highest prevalence (17.8%) was found in poor body condition animals while the least (4.76%) in good body conditions. This difference was statistically significant. The effect of age, sex, sites and body condition on trypanosomosis prevalence is summarized in table 4 and 5.

| Table 4: Prevalence of Trypanosomosis with age category in pawi spec   | cial district. |
|--|----------------|
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| Variable | No-of examined | No-positive | Prevalence %  |  |
|----------|----------------|-------------|---------------|--|
| Age      | No-of examined | No-positive | Trevalence 78 |  |
| <2       | 46             | 2           | 4.34%         |  |
| 2-5      | 130            | 12          | 9.23%         |  |
| >5       | 218            | 28          | 12.8%         |  |

| Table 5. Drevalence of Turmanagements | according to Dody | andition of no. | district     |
|---------------------------------------|-------------------|-----------------|--------------|
| Table 5: Prevalence of Trypanosomosis | according to bouy | condition of pa | wi uistrict. |

| Variable       | No-of examined | No-positive | Prevalence %  |
|----------------|----------------|-------------|---------------|
| Body condition | No-of examined | no-positive | Flevalence 76 |
| Good           | 42             | 2           | 4.76%         |
| Medium         | 104            | 14          | 13.46%        |
| Poor           | 146            | 26          | 17.8%         |
| Total          | 384            | 42          | 10.93%        |

| Variable | No-of examined | No-positive | Prevalence %  |  |
|----------|----------------|-------------|---------------|--|
| Sex      | No-or examined | No-positive | Trevarence 70 |  |
| Male     | 172            | 18          | 10.5%         |  |
| Female   | 212            | 24          | 11.32%        |  |
| Total    | 384            | 42          | 10.9%         |  |

#### 3.3. Entomological survey result

The entomological survey showed that the apparent densities of different flies across the rainy seasons in study area with total mean catches per trap per day were 0.78 for tsetse flies, also numeral number of *Stomoxys* and *Tabanus* were recorded. In Districts tsetse flies have highly infested and identified in selected Peasant association. The fly species were identified *G.m.submorsitans, and G.tachninoides* and it was coughed in mender 5, 14 and 17 PA around Beles river. In this study the distribution of *G. tachinoides*, was found to be along the large rivers of Beles and its tributaries, which cross most of zone the

region and has covered a vast area with even distribution in all PAs from low to high apparent densities in the District. A total of 78 tsetse fly were cached during the study period from different site. Out of the total, 60 (76.8%) were belonging to tsetse of the species Glossina tachinoides, and 18 (23%) were belonging to tsetse of the species Glossina morsitans submorsitans. In addition to tsetse fly, other biting fly such as 12(0.12) stomoxys and 8(0.8) tabanus were identified. The highest fly density 27 (1.35 F/T/D) were observed in Mender 14 and the lowest 9 (0.45 F/T/D) recorded in Almu and detail of the each District summarized in table below.

| No       | Kebele    | No of trap deployed | No of Catched fly | FTD  | Sex |    |
|----------|-----------|---------------------|-------------------|------|-----|----|
| INO      | Kebele    | No of trap deployed | No of Catched Hy  | I'ID | Μ   | F  |
| 1        | Mender 5  | 10                  | 9                 | 0.45 | 3   | 6  |
| 2        | Mender 14 | 10                  | 27                | 1.35 | 10  | 17 |
| 3        | Mender 17 | 10                  | 20                | 1    | 6   | 14 |
| <u>4</u> | Almu      | 10                  | 10                | 0.5  | 4   | 6  |
| <u>5</u> | Mender 7  | 10                  | 12                | 0.6  | 3   | 9  |
|          | Total     | 50                  | 78                | 0.78 | 25  | 53 |

 Table 7: Tsetse fly distribution of pawi district

| No       | Kebele    | No of Catched fly | Species of tsetse fly |                  | Biting fly |         |
|----------|-----------|-------------------|-----------------------|------------------|------------|---------|
|          |           |                   | G.tachnoides          | G.m.submorsitans | Stomoxy    | tabanas |
| 1        | Mender 5  | 9                 | 7                     | 2                | 3          | 1       |
| 2        | Mender 14 | 27                | 21                    | 6                | 1          | 2       |
| 3        | Mender 17 | 20                | 16                    | 4                | 2          | 2       |
| 4        | Almu      | 10                | 8                     | 2                | 2          | 1       |
| <u>5</u> | Mender 7  | 12                | 8                     | 4                | 4          | 2       |
|          | Total     | 78                | 60                    | 18               | 12         | 8       |

 Table 8: Vectors of trypanosomosis identified from the study sites

## 4. Discussions

The present study revealed an overall prevalence of 42/384 (10.94%) in the study area. This finding was in agreement with earlier works of Bayisa and Getachew who reported 11.7% prevalence in the neighboring Mandura district; Asmamaw, (2017) worked on the prevalence of bovine trypanosomosis in mandura district and reported prevalence of 13.10%, Bayisa and Getachew who reported 11.7% prevalence in the neighboring Dangur district; however present study was lower than works of Bayisa and Getachew who reported 22.38% prevalence in the neighboring Asossa district; Asmamaw and Getachew (2017), worked on the prevalence of bovine trypanosomosis in Asossa district and reported prevalence of 28.10%, Dawud and Molalegne (2011), worked on the prevalence of bovine trypanosomosis in Asossa district and reported prevalence of 24.7%, Mulaw et al. worked on the prevalence of bovine trypanosomosis in Asossa district and reported prevalence of 25.8%.

The difference in the prevalence of trypanosomosis in the previous and the current findings might be due to the difference in agro ecology and climatic conditions of the areas and partly it might be the difference in seasons in the study period and Assosa District from the regionis heavily infested with tsetse fly and have high prevalence of trypanomosis. T. congolense proportional prevalence of 61.4%; Biyazen, et al. (2014) reported T. congolense proportional prevalence 63.64% during his work on trypanosomosis and anemia in cattle population of Dale Wabera district of Kellem Wollega Zone, Western Ethiopia; Bayisa et al. (2015) demonstrated T. congolense proportional prevalence of 85% during his research on cattle trypanosomosis prevalence in Asossa district, Benishangul Gumuz Regional State, Western Ethiopia. Mulaw et al reported T. congolense proportional prevalence 66.7% during his work on trypanosomosis and anemia in cattle population of Dale Wabera district of Kellem Wollega Zone, Western Ethiopia Abraham and zerihun, (2012) reported T. congolense proportional prevalence 61.4% during his work on bovine trypanosomosis and anemia in selected sites in Arba minch district, southern

Ethiopia.

The high proportion infection rate of T. congolense in cattle might be attributable to the high number of serodems of T. congolense relative to T. vivax. It could also be due to the possible development of better immune response to T. vivax by the infected animals as demonstrated by Leak, et al. (1993). Further, it might be attributed to the efficient transmission of T.congolense by cyclical vectors than T.vivax in tsetse-infested areas. Previous reports indicated that T. congolense and T.vivax are the most prevalent trypanosomes that infect cattle in tsetse infested and tsetse free areas of Ethiopia respectively (Leak, 1999). Different studies (Leak, et al., 1993; Rowlands, et al., 1995) have indicated that T. vivax is highly susceptible to treatment while the problems of drug resistance are higher in T. congolense, and T. congolense is mainly confirmed in the blood, while T. vivax and T. brucei also invade the tissues.

The prevalence of bovine trypanosomosis was studied between sex categories, age groups and body conditions, though; significant association was not observed ( > 0.05). This might be because of an equal chance of exposure to the parasite. This result is in agreement with previous reports (Mihreteab and Mubarek, 2011; Lelisa, et al., 2015). The overall anemia prevalence in the studied district was 35.93% (138/384). The anemia prevalence was significantly higher in trypanosome infected cattle (58.6%) than in non-infected cattle (43.%) ( <0.05). This is in concordance with previous results from different researchers (Mihret and Mamo, 2007; Biyazen, et al., 2014).

Out of 49.5% anemia prevalence, 10.94% (42/384) was trypanosome infected animals. Nonetheless, 38.5% (148/384) of non-infected animals were found to be anemic (PCV < 24). This indicates the fact that other factors such as gastrointestinal parasitism, nutritional deficiencies, fasciolosis and vector-borne diseases could affect the PCV value of cattle (Rowland, et al., 1995).

This study revealed that 3.6% (14/384) of the cattle were infected by trypanosome; however, their PCV was laid in the normal range. This might be attributed to the capability of infected cattle to

maintain their PCV within the normal range for a certain period of time. It could also be possibly due to inadequacy of the detection method used (Murray, et al., 1997), other anemia causing diseases (Rowland, et al., 1995), or delayed recovery of the anemic situation after current treatment with trypanocidal drugs. Furthermore, the occurrence of positive animals with PCV of greater than 24% might be thought of as recent infections of the animals (Rowland, et al., 1995). The overall mean PCV value for examined animals was  $25.64 \pm 5.61$  SD. The mean PCV value of the infected animals was significantly lower ( $22.79 \pm 4.51$  SD) than that of uninfected animals ( $25.81 \pm 5.53$  SD). This result is in alignment with previous works (Mulaw, et al., 2011; Bayisa, et al., 2015).

Glossina tachinoides Glosina m.submorsitans was tsetse fly caughted and their mean apparent density measured as f/t/d was 0.78. In addition, other mechanical transmitters of trypanosomosis such as stomoxys 12 (0.12%), tabanus 8 (0.8%) were recorded. The current findings were in consistent with previous works of Solomon and Fitta, (2010) at Metekel Awi zones of Northwest Ethiopia, who reported 6.49 f/t/d and 0.65 f/t/d for tsetse and biting flies, respectively. It was also in agreement with findings of NTTICC, (2004) at Bure Iluababor zone of Western Ethiopia which was reported to be 7.23 f/t/d, 3.13 f/t/d and 0.06 f/t/d for tsetse, Stomoxys and Tabanus, respectively.

#### 5. Conclusions

The most common trypanosomes species identified were T.congolense followed by T.vivax. The animal parameters such as sex, age and body condition were not found to be a risk factor. The mean PCV value of infected animals was significantly lower than that of uninfected animals indicating the adverse effect of trypanosomosis on the PCV profile of cattle. Trypanosomes were not detected in some anemic cattle indicating the occurrence of other causes of anemia in the area. G.tachinoides and G.m.submorsitans was the tsetse fly species discovered in this study. Other mechanical transmitters of trypanosomosis such as stomoxys, and tabanus were recorded in the area. In wrapping up, trypanosomosis is an economically important disease threatening the health and productivity of cattle in Pawi district. Therefore, proper strategies have to be designed and implemented to minimize the impact of the disease on livestock production in the studied district.

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#### References

- 1. Abebe,G., (2005): Current situation of Trypanosomosis,a review article on Trypanosomosis in Ethiopia. Ethiop. J. Biol.Sci., 4: 75-121.
- Abraham, Z.A., and Zeryehun, T., (2012): Prevalence of Bovine Trypanosomosis in Selected District of Arba Minch, SNNP, Southern Ethiopia, Global Veterinaria, 8(2):168-173,2012,DOI:10.5829/ idosi.gv.2012.8.2.61312.
- 3. Asmamaw, A., and Mengistu, G., (2016): Across sectional Study on Bovine Trypanosomosis and Apparent Vector density in Bambasi District of Benishangul Gumuz Regional State, Western Ethiopia, prevalence and Vector density.
- Assosa Regional Veterinary Diagnostic, Surveillance, Monitoring and Study Laboratory (2015): laboratory annual report, Assosa, Ethiopia.
- Bayisa, K., and. Getachew, D., (2016): Trypanosomosis and its Associated Risks in Cattle Population of Dangur District of Benishangul Gumuz Regional State, Western Ethiopia. European Journal of Applied Sciences 7 (6): 291-296, 2015 ISSN 2079-2077 © IDOSI Publications,2015DOI:10.5829/idosi.ejas.2015.7. 6.101185.
- Biyazen, H., R., Duguma, and Asaye, M., (2014): Trypanosomosis, Its Risk Factors, and Anaemia in Cattle Population of Dale Wabera District of Kellem Wollega Zone, Western Ethiopia, Journal of Veterinary Medicine.
- Cecchi, G., Mattioli R.C., Slingenbergh, J., and De la Rocque, S., (2008): Land cover and tsetse fly distribution in sub-Saharan Africa. Med. Vet. Entom, 22: 364-373.
- 8. Central Statistical agency, (2014): Federal Democratic Republic of Ethiopia, Agricultural Sample Survey volume 2. 573 Statistical bulletin, pp: 39-49, 71.
- 9. Central Statistical agency, (2014 2017): Federal Democratic Republic of Ethiopia,

Population Projection of Ethiopia for all regions at woreda Level.

- Daud, A., and Molalegne, B., (2011): Epidemiological study of Bovine Trypanosmosis in Mao-komo Special District, BenishangulGumuz Regional State, Western Ethiopia. G lobal Veterinaria, 6: 402-408.
- De-Lahunta, A., and Habel, R.E., (1986): Applied veterinary Anatomy. USA. W. B. Sounders. Company, pp: 4-16.
- Getachew, D., and Asmamaw, A., (2016): Epidemiology of Cattle Trypanosomosis and Associated Anaemia in Mandura District. Nat Sci 2016, 14(5):85-90]. ISSN 1545-0740 (print); ISSN 2375-7167 (online). <u>http://www.sciencepub.net/nature.12</u>. doi:10.7537/marsnsj14051612.
- Leak, S.G.A., Mulatu W, E., Authie, D'Ieteren, G.D.M., and Peregrine, A.S., (1993): Epidemiology of bovine trypanosomosis in the Gibe valley, Southern Ethiopia. Tsetse challenge and its relationship to trypanosome prevalence in cattle. Acta Tropica, 53: 1221-1234.
- Leak, S.G.A., (1999): Tsetse biology and ecology: The role in the epidemiology and control of trypanosomosis. CAB International. Wallingford, UK, CABI Publishing and ILRI, pp: 152-210.
- Lelisa, K., Damena, D., Kedir, M., and Feyera, T., (2015): Prevalence of Bovine Trypanosomosis and Apparent Density of Tsetse and Other Biting Flies in Mandura District, Northwest Ethiopia. J Veterinar Sci Technol, 6: 229. doi:10.4172/2157-7579.1000229.
- Mihreteab, B., and Mubarek, N., (2011): Prevalence and host related risk factors of bovine trypanosomosis in Hawagelan district, West Wellega zone, Western Ethiopia. *African Journal* of Agricultural Research Vol. 6(22), Pp. 5055-5060.
- Mulaw, S., Addis, M., and Fromsa, A., (2011): Study on the Prevalence of Major Trypanosomes Affecting Bovine in Tsetse Infested Asossa District of Benishangul Gumuz Regional State, Western Ethiopia. Global Veterinaria, 7(4): 330-

336.

- Murray, M., and Dexter, T.M, (1988): Anemia in Bovine in African Animal Trypanosomosis. Acta. Top, 45: 389-432.
- National Meteorological Services Agency, (2015): Monthly report on temperature and Rainfall distribution for Mao-komo special district, Regional Metrological Office, Assosa, Ethiopia, pp: 17-19.
- 20. National Tsetse and Trypanosomosis Investigation and control center (1996): Annual Report. Ministry of Agriculture, Ethiopia.
- Rowlands, G.J., Mulatu, W.S., Nagda, M., Dolan, R.B., and D'Ietere, n G.D.M., (1995): Genetic variation in packed red cell volume and frequency of parasitaemia in East African Zebu cattle exposed to drug-resistant trypanosomes," Livestock Production Science, vol. 43, no. 1: pp. 75–84.
- 22. Shaw A. P. M., Cecchi G., Wint G. R. W., Mattioli R. C. and Robinson T. P., (2014): Mapping the economic benefits of livestock keepers from intervening against bovine trypanosomosis in Eastern Africa. Prev. Vet. Med. 113 (2):197-210.
- 23. Solomon, M., and Fitta, G., (2010): Survey on Bovine Trypanosomosis and its vector in Metekal and Awi Zones of Northwest Ethiopia, Acta Tropica, 117: 146-151.
- 24. Standardized techniques for the diagnosis of tsetse transmitted trypanosomosis, (2008): OIE Terrestrial Manual,; pp: 49, Rome, Italy.
- Taylor, M.A., Coop, R.L., and Wall, R.L., (2007): Veterinary Parasitology 3<sup>rd</sup> ed. Black Well publishing Ltd, oxford. Uk, Pp 96-102, 212-214.
- 26. Thrusfield, M., (2005): Veterinary Epidemiology. 3<sup>rd</sup> ed., UK, Blackwell Science Ltd., pp: 233-250.
- Truc P., Buscher P., Cuny G., Gonzatti M. I., Jannin J., Joshi P., Juyal P., Lun Z. R., mattioli R., Pays E., Simarro P. P., Teixeira M. M., Touratier L., Vincendeau P. and Desquesnes M., (2013): A typical human infections by animal trypanosomes. PLoS Negl. Trop. Dis. 7: 2256.

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