

Effect of Prebiotics and Probiotics on Immune Status in Poultry: A Review

Eman R. Hassan¹, Nagwa S. Rabie¹ and Mona S. Zaki²

¹Poultry Diseases Department, National Research Centre, Cairo, Egypt

²Hydrobiology Department, National Research Centre, Cairo, Egypt

nagwasrabie@hotmail.com

Abstract: The use of antibiotics becomes important to treat and protect broiler from microbial diseases. There is a relationship between hazard of pathogenic diseases and antibiotic used in poultry production. The Prebiotics and probiotic are known as antibiotic alternatives in poultry. Prebiotics, are non-digestible feed ingredients that have the ability to enrich or support the growth of single or multiple species of beneficial organisms in the gastrointestinal tract. Prebiotics include essential oils, plant extracts, spices, and organic acids. Probiotics are means as live microorganisms confer a health benefit on the host (broiler, turkey, and layers). Lactic acid bacteria, bacillus spores, and yeasts are the main groups of probiotic. The combination of probiotics and prebiotics is called synbiotics. Synbiotics are used for the synergistic improvement in reducing pathogen colonization in poultry by enhancing pre-harvest microbiological safety. Enhancement of the epithelial barrier, increase adhesion to intestinal mucosa, competitive exclusion of pathogenic microorganisms, and inhibition of pathogen adhesion production of antimicrobial substances are the major mechanisms of action of probiotics. Probiotics and prebiotics could modulate the systemic antibody response to antigens in chickens. Moreover, the administration of probiotics results in the secretion of cytokines and changes in lymphoid cells in the chicken gut, which may lead to enhanced immunity to pathogenic diseases.

[Eman R. Hassan, Nagwa S. Rabie and Mona S. Zaki. **Effect of Prebiotics and Probiotics on Immune Status in Poultry: A Review.** *Rep Opinion* 2018;10(2):24-28]. ISSN 1553-9873 (print); ISSN 2375-7205 (online). <http://www.sciencepub.net/report>. 3. doi: [10.7537/marsroj100218.03](https://doi.org/10.7537/marsroj100218.03).

Keywords: Poultry, Coccidiosis, Probiotics

Introduction

Poultry protein is the main favourite protein in most countries is the. Random use of antibiotics leads to increase microbial resistance (**Abd El- Hakim et al., 2009**). Antibiotics are important for the elimination of diseases. However, they also eliminate a large incidence of beneficial bacteria in the digestive system after treatment (**Dhama and Singh, 2010**).

Over use of antibiotics in poultry industry and its residue in poultry meat is harmful and has a toxic effect on consumers (**Kabir et al., 2004**). Elimination of antibiotics from poultry industry becomes an exhaustive search for alternatives to treat and protect broiler flocks from microbial diseases. The idea was produced when **Metchnikoff (1907)** studied the effect of "lactic acid bacteria" in fermented milk products. Intestinal bacteria played a role in the maintenance of health. Prebiotic supplementation resulted in improved body weight, feed conversion efficiency and carcass weight in broilers and subsequently improving productivity and increasing health benefits. (**El-Nagmy et al., 2000, and Hajati and Rezaei, 2010**).

The economic losses associated with enteric bacterial diseases can be summarized in; high mortality, loss of weight, poor feed conversion, the downgrading of carcasses, lowering fertility and hatchability as well as severe drop in egg production. *Colibacillosis, salmonellosis* and clostridial infections

considered as the major bacterial poultry enteric problems that adversely affect gut integrity leading to severe economic losses in the poultry industry (**Abd El-Samee, 2001**).

Prebiotics can be found in a variety of foods, including onions, and garlic, supplementary diets. They improve poultry resistance to enteric disease and promotes growth by several different means (**Mokhtari et. al., 2015**). Prebiotics affect- the immune system in poultry by stimulation of both cell-mediated and humeral immunity for better production of natural interferon/cytokines, increased macrophage, lymphocyte and natural killer (NK) cell activity, and increased immunoglobulin (IgG, IgM and IgA) (**Haghighi et al., 2005**).

Probiotics or direct feed microbial (DFM) are live microbial cultures, if administered in adequate quantity, result in desired health benefits in the host (**FAO, 2001**).

Probiotic are used in the poultry industry as alternative to antibiotics, it to have the ability to raise immunity during the management of microbial populations in the digestive system, to protect poultry from pathogenic bacteria and reduce the economic losses resulting from these diseases and modulate the immune system (**Amer et al., 2016**).

The mode of action of probiotics includes: competitive exclusion and antagonism, altering metabolism, improving digestion and feed intake

related to increasing digestive enzyme and decreasing bacterial enzyme and ammonia production (**Ghasemi et al., 2010**).

Competitive exclusion protect newly hatched chicks, turkey poults, quails and pheasants from infection (**Yang et al., 2008**).

Synbiotic is a combination, of probiotics and prebiotics which act synergistically to result in the selective enrichment/survival of both exogenous bacteria (probiotic) and endogenous bacteria; This combination could positively modulate beneficial microbiota in gastrointestinal tract and reduce pathogens including *Salmonella*, *Campylobacter*, and *Coccidia* (*Eimeria acervulina* and *Eimeria tenella*) (**Baffoni et al., 2012**). It was found that probiotic not only improves IgM antibody titer in broiler chickens but also improve antibody against specific against Newcastle disease virus (**Rowghanei et al., 2007**) Also it was found that *Bacillus subtilis* probiotic improve bursal body weight ratio indicating immune stimulation (**Teo and Tan, 2007**).

In addition *Bacillus subtilis* improves both bursal and spleen body weight ration indicating strong immune stimulation for both cellular and humoral immune response (**Willis et al., 2008**).

It was found that probiotic not only improves IgM antibody titer in broiler chickens but also improve antibody specific against Newcastle disease virus (**Rowghanei et al., 2007**).

Prebiotics

Prebiotics are dietary substances which increase the growth and/or activity of some beneficial intestinal bacteria. Prebiotics cannot be digested and not absorbed in the upper part of the digestive system. The site of action of prebiotics is the lower intestine, especially the colon, where short chain fatty acids (SCFAs) are produced after the prebiotics are fermented (**Patterson and Burkholder, 2003**).

Types of Prebiotics:

Prebiotics are small fragments of carbohydrates and are commercially available as oligosaccharides of galactose, fructose or mannose (**Biggs et al., 2007**). The most commonly used prebiotic preparations are fructo oligosaccharide (FOS), transgalactooligosaccharide (TOS), inulin, gluco oligosaccharide, xylo oligosaccharide, isomalto oligosaccharide, soybean oligosaccharide, polydextrose, lacto sucrose (**Vulevic et al., 2004**) Natural sources of prebiotics for vertebrates include chicory, onion, garlic, tomato and honey (**Chen and Chen, 2004**). Prebiotics were found to be effective against pathogenic bacteria like *Clostridium perfringens* (*C. perfringens*) and *Escherichia Coli*. Dietary supplementation of chicory oligofructose and inulin improved feed conversion ratio, egg production and increasing egg weight per bird in white leghorn hens. (**Yang et al., 2008**).

The principal mechanisms of prebiotics:

The principal mechanisms of prebiotics are immuno-modulation, include that

High fermentation activity and concentration of the SCFA are correlated with a lower pH, change in mucin production contribute to lower incidence of bacteria moving across the gut barrier and help in increasing solubility of certain nutrients and suppression of pathogens. (**Teo and Tan, 2007 and Lee and Salminen, 2009**). This phenomenon may inhibit some pathogenic bacteria and reduce colonization of some species like *Salmonella*, *Campylobacter* and *E. coli*, (**Chambers and Gong, 2011**).

Immune response Titer antibodies for Newcastle virus diseases Effect of diets supplemented with garlic powder was highly significant on titer of antibodies for Newcastle virus diseases (NVD) at different periods of 7, 14 and 21 days post vaccination (**Vetvicka and Sima, 2004**). Titer antibodies for avian influenza virus diseases (AIV) Results in show a trend similar to the effect of garlic powder on antibody titer against NVD. (**Kelly-Quaglian et al., 2003**). Inulin is the most common type of FOS. β -1 Fructans, which include inulin (IN) and fructo oligosaccharides (FOS), fulfil the criteria for prebiotics. (**Gibson et al., 2004**). IN is found naturally in a variety of plant foods such as barley, chicory, garlic, artichoke, leeks, onions and wheat. (**Roberfroid et al., 2005**).

Probiotics:

Probiotics are, nontoxic and nonpathogenic organisms in nature, which when administered through the digestive route, are favorable to the host's health (**Beynen, 2006**). Probiotics Stimulate the immune system and colonization of pathogenic strains of *E-coli* were reduced by native gastrointestinal microflora in the chicken and turkey (**Patterson and Burkholder., 2003**).

Types of probiotic and Micro-organisms used:

There are several types of probiotic micro-organism available in market, they are varied according to ability of colonization inside intestinal tract, (**Mountzouries et al., 2007**). Probiotic available commercially contains bacterial strains either identical to same micro-organisms that normal inhabitant in poultry intestinal tract such as *Lactobacillus casei*, *Lactobacillus lactis* and *Lactobacillus acidophilus*, while other contains strains differ from that normal inhabitant inside intestinal tract such as *Lactobacillus bulgaricus* and *Streptococcus thermophiles* (**Kabir, et al. 2004**).

Lactic acid bacteria:

Lactobacilli are a heterogeneous, non-spore, rod shaped, catalase negative gram-positive bacteria and several species viz. *Lactobacillus casei*, *L. zeeae*, *L. paracasei*, and *L. rhamnosus* have been commonly

used as probiotic (Jayaraman, et al 2015)

Aspergillus oryzae (A. oryzae):

The efficacy of commercially available fermentation product from *Aspergillus oryzae* and *A. niger* as a nutritional aid in alleviating protein insufficiency (Fuller, 1992). *A. oryzae* in broiler chickens has been shown to enhance the growth performance, significantly lower ammonia gas production, reduces serum cholesterol concentration in broilers and influenced microflora population in a beneficial manner (Yoon, et al., 2004).

Yeasts:

The genus *Saccharomyces* has 4 different species. *Saccharomyces cerevisiae* has thousands of strains. Only a few *S. cerevisiae*'s strains are used for poultry nutrition. Some of these strains have intestinal action result by: neutralization of certain bacterial toxins adherence of flagellate bacteria, due to the presence of mannose receptors (Tollba, et al., 2004).

Bacillus Species:

Several types of *Bacillus* (B) have antagonistic activity against *C. perfringens* due to the production of bacteriocins. For example, *B. thuringiensis* produces thuricin which is active against *C. perfringens*, (Nisbet, 2002).

Mechanisms of Action of Probiotics:

Major Probiotic mechanisms of action include competitive exclusion of pathogenic microorganisms, enhancement of the epithelial barrier, increased adhesion to intestinal mucosa, and inhibition of pathogen adhesion, production of anti-microorganism substances and modulation of the immune system (Ghareeb and Böhm, 2009).

Competitive exclusion:

The exact mechanism of action of probiotics associated with "competitive exclusion" or "bacterial antagonism" which refers to the physical blocking of pathogen colonization within the intestinal tract which are favourite sites of enteric pathogens by the probiotic bacteria (Duggan et al., 2002). Antibacterial substances produced like bacteriocins, lactocin, lactocidin, acidolin, acidophilin, nisin, organic acids (lactic and acetic acid), lactoferrin, hydrogen peroxide and lacto peroxidase inhibit pathogenic microbes (Dhama et al., 2007).

Enhancement of the Epithelial Barrier:

Defenses of the intestinal barrier consist of the mucous layer, antimicrobial peptides, secretory IgA and the epithelial junction adhesion complex (Ohland et al., 2010). Once this barrier function is disrupted, bacterial and food antigens can reach the submucosa and can induce inflammatory responses, which may result in intestinal disorders, such as inflammatory bowel disease (Farnell et al., 2006).

Increased Adhesion to Intestinal Mucosa:

Adhesion of probiotics to the intestinal mucosa is

also important for modulation of the immune system and antagonism against pathogens adhesion has been one of the main selection criteria for new probiotic strains and has been related to certain beneficial effects of probiotics (Juntunen, et. al. 2001).

Immune effect of prebiotics and probiotics in Poultry:

The bursa of Fabricius and the thymus regulate the development of the humoral and cellular compartments of the immune system, respectively. (Amit, et al. 2004).

Peripheral organs: spleen, gut-, bronchus-and head-associated lymphoid tissue (GALT, BALT and HALT, respectively) (Christensen et al., 2002).

The positive effect of feeding diet containing probiotic on the immune response indicates the enhancement of the formulating bacteria on an acquired immune response exerted by T and B lymphocytes. *The production of the mucosal IgA response is dependent on other cytokines, such as transforming growth factor* (Nisbet et al., 1993).

Prebiotics affect on the immune system in poultry via. Stimulation of both cell-mediated immunity and humeral immunity. with better production of natural interferon/cytokines, increased macrophage, lymphocyte and natural killer (NK) cell activity, and increased immunoglobulin (IgG, IgM and IgA) (Haghighi et al., 2005).

Particularly through balanced control of anti-inflammatory and pro-inflammatory cytokines. Probiotics increase the number of lymphocytes and lymphoid cells in lamina propria and intraepithelial lymphocytes (IEL) in the small intestine and are found to inhibit the growth of infectious organisms (Mason, et al., 2005).

Conclusion

Prebiotics, probiotics and synbiotics plays an important role in improving the productive and economic efficiency of poultry farms.

It is of great value to use probiotic or prebiotic as immune stimulant as it was found that it stimulate both cellular or humoral immune response including local immune response.

Some probiotics are responsible of production of proteins peptides which cause reduction in pathogenic bacteria of gut and producing antibodies thus giving immunity against the diseases attack.

References

1. Abd El-Hakim, A. S., Cherian, G., Ali M. N. (2009). Use of organic acid, herbs and their combination to improve the utilization of commercial low protein broiler diets. *Int. J. Poul. Sci*; 8 (1): 1420.
2. Abd El-Samee, M. O. (2001). Broiler

- performance as affected by crude protein, lysine and a probiotic. *Egypt. Poult. Sci.*, 21 (4): 943-962.
3. Amer M. M., Elbayoumi Kh. M., Zeinab M. S. Amin Girh, Eman R. Hassan, M. A. Bosila and Hoda M. Mekky (2016). *International Journal of Pharm Tech Research* Vol.9, No. 12, pp 58- 69.
 4. Amit-Romach, E., D. Sklan, and Z. Uni. (2004). Microflora ecology of the chicken intestine using 16S ribosomal DNA primers. *Poult. Sci.* 83:1093–1098.
 5. Baffoni, L., Gaggia, F., Di Gioia, D., Santini, C., Mogna, L. and Biavati, B. (2012). AB ifidobacterium-based synbiotic product to reduce the transmission of *C. jejuni* along the poultry food chain. *International J. Food Microbiol*; 157(2): 156-161.
 6. Beynen, A C; (2006). Mortality and growth performance of broilers given drinking water supplemented with chicken-specific probiotics. *Poult. Sci.*, 85(8): 1383-1388.
 7. Biggs, P., Parsons, C. M., and Fahey, G. C. (2007). The effects of several oligosaccharides on growth performance, nutrient digestibilities, and cecal microbial populations in young chicks. *Poult. Sci*; 86: 2327-2336.
 8. Chambers; J. R. and Gong, J. (2011). The intestinal microbiota and its modulation for *Salmonella* control in chickens. *Food Res. Internatl*;44. (10): 3149-3159.
 9. Chen, Y. C. and Chen, T. C. (2004). Mineral utilization in layers as influenced by dietary oligofructose and inulin. *International J. Poultry Sci.* 3(7), 442-445.
 10. Chen, Y. C., Nakthong, C. and Chen, T. C., (2005). Improvement of laying hen performance by dietary prebiotic chicory oligofructose and inulin. *Int. J. Poult. Sci*; (4): 103-108.
 11. Christensen, H. R., H. Frokiaer, and J. J. Pestka (2002). *Lactobacilli* differentially modulate expression of cytokines and maturation surface markers in murine dendritic cells. *Journal of immunology.* 186:171-17.
 12. Dhama K and Singh S D. (2010). Probiotics improving poultry health and production: an overview. *Poultry Punch*, 26(3): 41.
 13. Dhama K, Mahendran M and Tomar, S. (2007). Probiotics and prebiotics: A safer way towards improving health and productivity in poultry. *Poultry World.*2 (7): 28-32.
 14. Duggan C., Gannon J. and Walker W. A. (2002). Protective nutrients and functional foods for the gastrointestinal tract. *Am J. Clin. Nutr.*,75:789-808.
 15. El-Nagmy K. Y. Ali, A. M., and Abd El-Samee M. O. (2000). The effect of dietary protein and yeast culture levels on performance of growing Japanese Quail. *Egypt. Poult. Sci.*, 20:777-787.
 16. FAO/WHO. (2001). Health and nutritional properties of probiotics in food including powder milk with live lactic acid bacteria. Food and Agriculture Organization of the United Nations.
 17. Ferket, P. R. (2003). Controlling gut health without the use of antibiotics. In *Proceedings of the 30th Annual Carolina Poultry Nutr. Conf.* (pp. 57-68).
 18. Fernandez, F., Hinton, M., and Gils, B. V. (2002). Dietary mannan- oligosaccharides and their effect on chicken caecal microflora in relation to *Salmonella Enteritidis* colonization. *Avian Pathol.* 31(1), 49-58.
 19. Fuller R. (1992). History and development of probiotics. In Fuller R. (Ed), *probiotic, The Scientific Basis.* Chapman and Hall, London, pp: 1-8.
 20. Ghareeb, K. and Böhm, J. (2009). Stress indicators to pre-slaughter transportation of broiler chickens fed diets supplemented with a synbiotic. *Internatl. J. Poultry Sci.* 8(7), 621-625.
 21. Ghasemi, H. A., Shivazad, M., Esmaeilnia, K., Kohram, H., and Karimi, M. A. (2010). The effects of a synbiotic containing *Enterococcus faecium* and inulin on growth performance and resistance to coccidiosis in broiler chickens. *J. Poultry Sc.* 47(2), 149-155.
 22. Gibson G. R., Probert H. M., Van Loo J., Rastall R. A. and Roberfroid M. B. (2004). Dietary modulation of the human colonic microbiota: updating the concept of prebiotics. *Nutr Res Rev*17, 259 – 275.
 23. Haghighi, H. R. J. Gong, C. L. Gyles, M. A. Hayes, B. Sanei, P. Parvizi, H. Gisavi, J. R. Chambers and S. Sharif (2005). Modulation of Antibody-Mediated Immune Response by Probiotics in Chickens. *Clinical and Vaccine Immunology.* 12:1387-1392.
 24. Hajati H and Rezaei M. (2010). The application of probiotics in poultry production. *Inter. J. Poult. Sci.*, 9:298- 304.
 25. Jayaraman, S., Thangavel, G., Kurian, H., Mani, R., Mukkalil, R. and Chirakkal, H.; Doron, S. and Snyderman, D. R. (2015). Risk and Safety of Probiotics. *Clinical Infectious Diseases*,60(suppl. 2).
 26. Juntunen M., Kirjavainen P. V., Ouwehand A. C., Salminen S. J., Isolauri E. (2001). Adherence of probiotic bacteria to human intestinal mucus in healthy infants and during rotavirus infection. *Clin Diag Lab Immunol*; 8: 293–296. 42.
 27. Kabir, S. M. L.; Rahman, M. M.; Rahman, M. B.; Ahmed, S. U. (2004). The dynamics of probiotics on growth performance and immune response

- inbroilers. *Int. J. Poult. Sci.* 3: 361 -364.
28. Kelly-Quagliana K., Nelson P. and Buddington R. (2003). Dietary oligofructose and inulin modulate immune functions in mice. *Nutr Res* 23, 257 – 267.
 29. Mason C. K., Collins M. A. and Thompson K. (2005). Modified electroporation protocol for lactobacilli isolated from the chicken crop facilitates transformation and the use of genetic tool. *J. Microbiol. Methods*, 60:353-363.
 30. Mokhtari, R.; Yazdani, A. and Kashfi, H. (2015). The effects of different growth promoters on performance and carcass characteristics of broiler chickens *Journal of Veterinary Medicine and Animal Health, academic Journals*. 7(8):271 -277.
 31. Mountzouris K. C., Tsirtsikos P., Kalamara E., Nitsch S., Schatzmayr G. and Fegeros K. (2007). Evaluation of the efficacy of a probiotic containing *Lactobacillus*, *Bifidobacterium*, *Enterococcus* and *Pediococcus* strains in promoting broiler performance and modulating cecal microflora composition and metabolic activities. *Poult Sci.*, 86(2): 309-317.
 32. Nisbet D., (2002). Defined competitive exclusion cultures in the prevention of enteropathogen colonization in poultry and swine. *Antoine van Leeuwenhoek*, 81:481-486.
 33. Ohland C. L., Macnaughton W. K. (2010). Probiotic bacteria and intestinal epithelial barrier function. *Am J Physiol Gastrointest Liver Physiol* 298: G807–G819.
 34. Patterson J. A. and Burkholder K. M. (2003). Application of prebiotics and probiotics in poultry production. *Poult. Sci.*, 82: 627-631.
 35. Roberfroid, M. B., Van Loo, J. A., and Gibson, G. R. (2005). The bifidogenic nature of chicory inulin and its hydrolysis products. *J. Nutr.* 128(1), 11-19.
 36. Rowghani E., Arab M. and Akbarian A. (2007). Effects of probiotic and other feed additives on performance and immune response of broiler chicks. *Int. J. Poult. Sci.*, 6: 261-265.
 37. Teo A. Y. and Tan H. M. (2007): Evaluation of the Performance and Intestinal Gut Microflora of Broilers Fed on Corn-Soy Diets Supplemented With *Bacillus subtilis* PB6 (Clo STAT). *Journal of applied poultry*. 16:296-303.
 38. Teo, A. Y. L. and Tan, H. M. (2005). Inhibition of *Clostridium perfringens* by a novel strain of *Bacillus subtilis* isolated from the gastrointestinal tracts of healthy chickens. *Applied Environmental Microbiology*, 71(8): 4185-4190.
 39. Tollba A. A. H., Sabery M. M. and Medani G. G. (2004). Effect of microbial probiotics on performance of broiler chicks under normal or heat stress conditions. 2-Bacterial concentration or yeast culture. *Egypt. Poult. Sci. J.*, 24: 333-349.
 40. Vetricka V. and P. Sima, (2004). β -glucan in invertebrates. *ISJ*, 1:60-65.
 41. Vicente J. L., Wolfenden A., Torres-Rodriguez A., Higgins S., Tellez G. and Hargis B. M. (2007). Effect of probiotic culture candidates on *Salmonella* prevalence in commercial turkey houses. *J. Appl. Poult. Res.*, 16: 471-476.
 42. Vulevic J., Rastall R. A. and G. R. Gibson, (2004). Developing a quantitative approach for determining the in vitro prebiotics potential of dietary oligosaccharides. *FEMS Microbiol. Lett.*, 236:153-159.
 43. Willis W. L. and Reid L. (2008). Investigating the effects of dietary probiotic feeding regimens on broiler chicken production and *Campylobacter jejuni* presence. *Poult Sci.*, 87(4): 606-11.
 44. Yang, Y., Iji, P. A., Kocher, A., Thomson, E., Mikkelsen, L. L., and Choct, M. (2008). Effects of mannanoligosaccharide in broiler chicken diets on growth performance, energy utilisation, nutrient digestibility and intestinal microflora. *British Poultry Sci.* 49(2), 186-194.
 45. Yoon, C., Na, C. S., Park, J. H., Han, S. K., Nam, Y. M., and Kwon, J. T. (2004). Effect of feeding multiple probiotics on performance and fecal noxious gas emission in broiler chicks. *Korean J. Poultry Sci.* 31(4), 229-235.