



## Review On Bovine Spongiform Encephalopathy (BSE)

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**Abstract:** Bovine spongiform encephalopathy (BSE), a member of the transmissible spongiform encephalopathies (TSE), primarily affects cattle. Transmission is via concentrate feed rations contaminated with infected meat and bone meal (MBM). In addition to cattle, other food animal species are susceptible to BSE and also pose a potential threat to human health as consumption of infected meat products is the cause of variant Creutzfeldt-Jakob disease in humans, which is invariably fatal. In the UK, farmed and free ranging deer were almost certainly exposed to BSE infected MBM in proprietary feeds prior to legislation banning its inclusion. Therefore, although BSE has never been diagnosed in any deer species, a possible risk to human health remains via ingestion of cervine products. Chronic wasting disease (CWD), also a TSE, naturally infects several cervid species in North America and is spreading rapidly in both captive and free-ranging populations.

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### 1. INTRODUCTION

Bovine spongiform encephalopathy (BSE) commonly known as mad cow disease, is a transmissible, neurodegenerative disease affecting cattle. The disease has a long incubation period ranging from 30 months to eight years, with the infectious agent thought to be a specific type of misfolded protein, called a prion. These malformed prions cause other native prion proteins in the brain to misfold and aggregate, leading to a spongy degeneration of the brain and spinal cord. Transmission between cattle occurs via the consumption of contaminated meat and bone meal in cattle feed, and BSE is fatal, with no known cure or treatment. It is now believed that BSE may be transmitted to humans who consume infected beef or come into contact with other products derived from the nervous tissues of infected cattle. In humans, the disease is known as vCJD [WHO, 2008].

Bovine spongiform encephalopathy is a chronic degenerative disease affecting the central nervous system of cattle. The disease was first diagnosed in 1986 in Great Britain. BSE has had a substantial impact on the livestock industry in the United Kingdom. The disease also has been confirmed in native-born cattle in Austria, Belgium, Czech Republic, Denmark, Finland, France, Germany, Italy, Ireland, Japan, Liechtenstein,

Luxembourg, the Netherlands, Northern Ireland, Portugal, Slovakia, Slovenia, Spain, and Switzerland.

### Objectives

The objective of this study is to

- ✓ review on bovine spongiform encephalopathy
- ✓ Evaluate the mode of transmission, clinical signs, risk factor and ways of prevention and control method.

### 2. LITERATURE REVIEW

#### 2.1. Epidemiology

Epidemiologic data suggest BSE in Great Britain is an extended common source epidemic involving animal feed containing contaminated meat and bone meal as a protein meat source. There are different scientific hypotheses concerning the origins of Bovine Spongiform Encephalopathy (BSE). BSE in Great Britain may have been caused by feeding cattle rendered protein produced from the carcasses of scrapie-infected sheep or cattle with a previously unidentified Transmissible spongiform encephalopathies (TSE). The practice of using products such as meat and bone meal as a source of protein in cattle rations has been common for several decades. Changes in rendering operations in the late 1970's and early 1980's may have played a part in the appearance of the disease. There is no evidence

that BSE spreads horizontally, i.e., by contact between unrelated adult cattle or contact between cattle and other species (Harman and Silva 2009). Limited research suggests that maternal or vertical transmission may occur at a very low level. This low level most likely would not perpetuate the epidemic under British farming conditions. BSE is classified as a transmissible spongiform encephalopathy (TSE). The agent responsible for BSE and other TSE's is smaller than the smallest known virus and has not been completely characterized (Harman and Silva 2009). There are three main theories on the nature of the agent: (1) the agent is a virus with unusual characteristics, (2) the agent is a prion—an exclusively host-coded protein that is modified to a partially protease-resistant form after infection, and (3) the agent is a virino a small, noncoding regulatory nucleic acid coated with a host-derived protective protein. The BSE agent is extremely resistant to heat and to normal sterilization processes. It also does not evoke any detectable immune response or inflammatory reaction in host animals (USDA FSIS 2005; St Rose *et al.*, 2006; Harman and Silva 2009). In cattle naturally infected with BSE, the BSE agent has been found only in brain tissue, in the spinal cord, and in the retina. The distal ileum, bone marrow, dorsal root ganglion, and trigeminal ganglion from experimentally infected cattle were also found to be infective. To date, there has been no evidence of infection detected in milk or muscle tissue. The presence of the BSE agent in tissues is determined by inoculating animals, usually mice, with material believed to be infected with BSE. Mouse inoculation studies take a long time (up to 700 days) to detect the agent, and failure to identify it in tissues may indicate either true absence of the agent or simply the limited sensitivity of current diagnostic methods prions (bovine spongiform encephalopathy. Bovine spongiform encephalopathy (BSE) is a fatal neurodegenerative disease of cattle. It is caused by proteinaceous infectious particles known as prions. BSE is the only Transmissible Spongiform Encephalopathy (TSE) of animals that is known to be infectious to humans through the consumption of contaminated meat. The human form of the disease is known as variant Creutzfeldt - Jakob disease (vCJD).

## 2.2. Incidence

### 2.2.1. In animals

More than 184,000 cases of classical BSE have been diagnosed in cattle, and at the peak of the epidemic 1,000 cases were being diagnosed each week in the UK (Imran and Mahmood 2011b; OIE 2013). The epidemic is believed to have been amplified from a single common source (Aguzzi and

Calella 2009). The infection was spread elsewhere in Europe and the world by exports of infected cattle and MBM from the UK (Seuberlich *et al.* 2010). The feeding of MBM to cattle was banned in the UK in 1988, and in 1996 it became illegal in the UK to prepare any feed containing any mammalian protein for any farm animal. However, because of the long incubation period of BSE cases continued to occur, peaking in 1992. The incidence of new cases has steadily declined since then, and the disease is now very rare (Hueston and Bryant 2005).

A number of zoo and domestic animals developed TSEs at the same time as the BSE epidemic in cattle. All the species affected belonged to either the *Bovidae* or *Felidae* family, with the exceptions of a small number of non-human primates (Imran and Mahmood 2011b). All cases in zoo animals were attributed to ingestion of infective material derived from bovine BSE cases, as were two cases in domestic goats (Spiropoulos *et al.* 2011). A number of domestic cats developed TSE concurrently with the bovine BSE epidemic, and these cases were attributed to consumption of infective material in beef or beef-derived pet food (Harman and Silva 2009; Imran and Mahmood 2011b). Epidemics of TSEs were not observed in other domestic species at the same time. Dogs and horses express PrPC with a very stable structure that is resistant to mis-folding, and these species are resistant to infection with PrPSc (Harman and Silva 2009; Zhang 2011).

Pigs are highly resistant to oral infection with the BSE prion, but may be infected by parenteral challenge (Harman and Silva 2009). The original source of the classical BSE epidemic has been the subject of much conjecture but remains unknown. Surveillance in some countries has shown that sporadic cases of BSE occur in cattle, although to date only two strains distinct from 'classical' BSE, the L- and Htype strains, have been observed. However, it is possible that classical BSE may also occur spontaneously and that the epidemic represented amplification of infective material originating from a single sporadic case. It is also possible that the BSE epidemic originated from material from another species also rendered to produce MBM. It has been suggested that the BSE epidemic arose from rendering of scrapie-infected sheep. However, although sheep scrapie samples can cause a TSE when inoculated intracerebrally into cattle, the disease does not resemble BSE, and experimental BSE in sheep does not resemble scrapie. In fact cattle are resistant to oral infection with scrapie or CWD (Harman and Silva, 2009).

### 2.2.2. In humans

The first 10 human patients with vCJD were reported in April 1996 in the UK (Mackay *et al.*, 2011; Imran and Mahmood 2011a). As of December 2012, 227 vCJD cases have been reported in total. Current data may be found on the UK National Creutzfeldt-Jakob Disease Research & Surveillance Unit website, <http://www.cjd.ed.ac.uk>. The majority of cases (176) occurred in the UK (Imran and Mahmood, 2011a). There is strong evidence to show that vCJD is caused by ingestion of BSE infective material. Classical BSE prions from affected cows and vCJD prions from the brains of infected humans produce the same lesions in mice. The biochemical properties of BSE prions from cattle and vCJD prions from humans are indistinguishable. Furthermore the great majority of vCJD cases have occurred in the UK, with a few cases in other countries including France, Ireland and Italy (Aguzzi and Calella 2009). A two-fold difference was seen between the prevalence of vCJD in the north versus the south of the UK. Contemporaneous National Dietary Surveys showed that consumption of mechanically recovered beef products was much higher in the north of the UK (Mackay *et al.* 2011). Mechanically recovered beef is more likely to be contaminated with infective material in the spinal cord, and recovery of beef by this method is no longer permitted for human foodstuffs across Europe.

### 2.3. Description of the infective agent

BSE and other TSEs are caused by a mis-folded isoform of the prion protein (PrP), a widely expressed glycoprotein. PrP is a normal constituent of cell membranes in vertebrates, and is encoded by the prion protein gene *PRNP*. The mis-folded pathogenic isoform protein is often referred to as a 'prion', a term made up from the contraction of the words 'proteinaceous' and 'infectious' (Prusiner 1982). By convention, the normal cellular isoform of PrP is represented as PrP<sup>C</sup>. The C superscript refers to the cellular form. The prion form has the same amino acid sequence as the normal form and is represented as PrP<sup>Sc</sup>. The Sc superscript is a reference to scrapie, a disease of sheep that is the prototypical animal prion disease. The prions replicate themselves by binding to the normal PrP<sup>C</sup> protein and acting as a template that coerces the PrP<sup>C</sup> molecule to refold into the abnormal PrP<sup>Sc</sup> form (Gains and LeBlanc 2007; Cobb and Surewicz 2009).

PrP<sup>C</sup> has been identified in mammals, birds, reptiles, amphibians, fish and yeasts. In mammals, the protein is expressed in a wide variety of tissues including spleen, lymph nodes, kidney, pancreas, salivary gland, adrenal gland, liver, thymus, and bone

marrow; and is highly expressed in the nervous system (Gains and LeBlanc 2007; Linden *et al.* 2008; Brown and Mastrianni 2010). However, the physiological function of PrP<sup>C</sup> remains obscure and a number of strains of mice bred not to express PrP<sup>C</sup> show only subtle, non-lethal differences in physiologic and locomotor activity when compared to wild-type mice (Cobb and Surewicz 2009; Chakrabarti *et al.*, 2009). Within some prion diseases, including BSE and scrapie, strains exist which exhibit distinct disease phenotypes. Differences between strains include patterns of protein deposition in the brain and lymphoid tissues, incubation times after experimental infection of animals, histopathology and clinical manifestation. For example, with scrapie some strains preferentially propagate in the central nervous system while others are characterized by substantial infectivity in lymphoid organs (Aguzzi and Calella 2009). There are three strains of BSE which have been identified. Only one strain, classical BSE, was responsible for the BSE epidemic which started in the United Kingdom (UK) and spread to other countries, and the associated epidemic of vCJD in humans (Harman and Silva 2009). The atypical strains, known as H (high) and L (low) type, are diagnosed rarely, typically in cattle of 8–20 years of age, and appear to be sporadic and arise spontaneously (Seuberlich *et al.*, 2010; Konold *et al.*, 2012).

### 2.4. Stability characteristics

Prions are notoriously resistant to inactivation with conventional sterilization procedures used for preparation of surgical instruments and materials. PrP<sup>Sc</sup> is resistant to UV irradiation at 254 nm, 70% alcohol treatment, gamma irradiation, and conventional autoclaving (121°C for 20 minutes). PrP<sup>Sc</sup> can be inactivated by a number of measures including severe autoclaving conditions (134°C for 8-18 minutes) in conjunction with detergents and hydrogen peroxide gas plasma sterilization (Aguzzi and Calella 2009; Sakudo *et al.*, 2011). A number of procedures that modify or hydrolyse proteins can reduce the infectivity of prions (Aguzzi and Calella 2009). However, while PrP<sup>C</sup> is protease-sensitive and soluble in nondenaturing detergents, PrP<sup>Sc</sup> is insoluble in detergents and contains a protease-resistant core (Gains and LeBlanc, 2007).

### 2.5. Transmission

#### 2.5.1. Infectivity

There is no robust evidence that BSE can be transmitted between cattle by routes other than consumption of feed contaminated with certain tissues from BSE-infected cattle. This is in marked

contrast to the horizontal infectivity of scrapie in sheep and chronic wasting disease (CWD) in deer. CWD prions are found in saliva, urine, faeces, placenta or decomposed carcasses (Gough and Maddison 2010; Haley *et al.*, 2011; Imran and Mahmood, 2011b). PrPSc from scrapie-infected sheep is found in faeces, milk, saliva, nasal secretions and placental tissues. Scrapie and CWD prions have also been shown to persist in the environment, bound to soil or other fomites, but there is no evidence that BSE has been transmitted between cattle by this route, or via exposure to excreta or secretions (Gough and Maddison, 2010). There is no evidence that vCJD has been transmitted between humans except due to medical intervention such as blood transfusion (Brown and Matrianni, 2010).

### 2.5.2. Infective Dose

It appears that ingestion of less than 1 mg of infected brain material may be sufficient to transmit

infection between cattle (Harman and Silva 2009). Transmission of BSE to macaques has been accomplished by oral administration of 5 g of infective brain homogenate, but the infective dose of bovine PrPSc to human beings is unknown (Mackay *et al.*, 2011). The BSE agent (designated PrPSc) is, according to the current prion theory, an aberrantly folded (isomer) of a normal cell-surface protein (designated PrPc) that is able to induce conformational changes in PrPc so that PrPSc is produced in increasing amounts in the central nervous system (CNS) of affected cattle. By the end of the long incubation period this process results in neurological disturbances characteristic of BSE (Griffiths, 1967; Reik *et al.*, 2011).

The pathogenesis is similar in all the so-called transmissible spongiform encephalopathies (TSEs) that affect humans and other mammals (Table 1).

**Table 1: Transmissible spongiform encephalopathies that affect humans and animals**

Disease	Natural host	Other hosts
Scrapie	Sheep and goats	
Transmissible mink encephalopathy	Mink	
Chronic wasting disease	Mule deer, elk & white tailed deer	
Bovine spongiform encephalopathy	Cattle & goats	Domestic cats, captive wild felids (lions, tigers, cheetahs) and bison & antelope
Variant Creutzfeldt-Jakob disease	Humans	
Kuru	Humans	
Creutzfeldt-Jakob disease	Humans	
Gerstmann-Sträussler-Scheinker syndrome	Humans	
Fatal familial insomnia	Humans	

There is compelling evidence that consumption by people of offal (particularly brain and spinal cord) from infected cattle causes a similar neurodegenerative disease in people known as (new) variant-Creutzfeldt-Jakob disease (vCJD), also after a protracted incubation period (Bradley & Verwoerd,

2004a). This makes vCJD a zoonosis, the only one known among the TSEs. Like BSE, vCJD is invariably fatal and affected people usually suffer months of debilitating neurological illness before they die.

### 2.5.3. Mode of transmission

The epidemic of BSE, first recognized in 1986 in the UK, was propagated by the rendering of dead cattle infected with BSE to produce MBM which was then included in feed for cattle (Harman and Silva 2009). Ingestion of infectious material in MBM made from BSE-infected animals was the only known route of transmission of the agent between cattle. Consumption of beef contaminated with infected bovine central nervous system tissue also led to an epidemic of vCJD in humans. Although the majority of vCJD cases have been attributed to consumption of such contaminated beef, four cases of person-to-person vCJD transmission by blood or plasma transfusion have been reported in the UK (PHE 2009; Imran and Mahmood 2011a). Rare cases of transmission of sCJD between humans have resulted from corneal grafts, dura mater grafts and growth hormone injections (Brown and Mastrianni 2010). Similar transmission of vCJD remains a concern because retrospective analysis of tonsil and appendix specimens led to the estimation that up to 1 in 4,000 persons exposed during the UK epidemic may be a sub-clinical carrier (Harman and Silva 2009; Collinge 2012). There is no evidence that sCJD is a TSE of animal origin because this disease develops even among lifelong vegetarians (Harman and Silva 2009).

### 2.6. Pathogenesis

The pathogenesis of BSE in cattle has been studied extensively although there are still a number of knowledge gaps. After oral exposure of calves to infective material, PrPSc is first observed in Peyer's patches of the ileum, and also detected in gut-associated lymphoid tissue (GALT) of the ileocaecal junction and the jejunum. The infectivity is located in macrophages and follicular dendritic cells (FDC). Later, infectivity can be identified in the enteric nervous system, although it is not clear how infectivity moves from the cells of the lymphoreticular system to those of the nervous system (Hoffmann *et al.*, 2011). It is possible that after crossing the mucosal barrier of the intestine, prions infect the nervous tissue when they come into contact with the fine nerve fibres directly under the intestinal mucosa (van Keulen *et al.*, 2008). Once the nervous system is infected, infectivity then ascends to the brain via both the sympathetic (e.g. splanchnic nerve) and parasympathetic (e.g. vagus nerve) nervous systems (Cobb and Surewicz 2009). Involvement of GALT is less extensive in BSE than in ovine scrapie (van Keulen *et al.*, 2008).

It has been proposed that orally acquired prion diseases can also reach the brain through the bloodstream (Caughey *et al.*, 2009; Gough and

Maddison 2010), but infectivity is not detectable in the blood of BSE affected cows (van Keulen *et al.*, 2008). This is in contrast to experimental BSE in sheep and human vCJD, in which GALT shows high levels of infectivity and the blood also contains prions (Gough and Maddison 2010). The role of replication of PrPSc in FDCs of the spleen in propagation of the agent is unclear, and may vary between species. Studies of scrapie have provided evidence that depletion of FDCs prevents or delays neuroinvasion, that increased nerve supply to the spleen promotes neuroinvasion and that denervation of the spleen delays or prevents neuroinvasion (Gains and LeBlanc 2007). Splenic PrPSc is found in BSE infection of mice expressing the ovine prion protein (Baron *et al.*, 2010). However, splenic PrPSc was detected in only one of three cattle terminated in the advanced clinical stage of BSE (Murayama *et al.*, 2010).

Once a cell is infected with PrPSc, spread of infection to adjacent cells may occur by transfer of PrPSc-containing membrane microparticles. Consistent with this hypothesis, it has been shown that PrPSc can be released from infected cells *in vitro* in association with exosomes. Exosomes are small membrane-bound vesicles that can be secreted by cells and can fuse with other cells. However, although exosome production by lymphoid cells has been demonstrated, exosomes have not been shown to be produced by neurons. Another possible route by which PrPSc could be transferred between adjacent cells is via tunneling nanotubes, thin membranous bridges that can form between cells and allow the transfer of organelles, plasma membrane components, cytoplasmic molecules and pathogens (Caughey *et al.*, 2009).

Other proposed pathways of propagation within the nervous system include axonal transport, sequential infection of Schwann cells (cells that support and insulate peripheral nerves) and via the flow of lymph in the vicinity of neurons (Kovacs and Budka 2008). The molecular pathways leading to cerebral damage are largely unknown, although various theories have been advanced. Depletion of PrPC does not appear to be a cause, as mice that have been genetically engineered to lack PrPC altogether, and those in which PrPC expression is turned off in adulthood, do not develop clinical signs of TSE. In fact depletion of PrPC in mice with established prion infection has been shown to reverse early spongiform degeneration and prevent progression to clinical disease. These findings suggest that the toxicity of PrPSc depends on some PrPC-dependent process (Aguzzi and Calella 2009). It has been suggested that PrPC is neuroprotective and its conversion to PrPSc interferes with this function and allows



neurodegeneration (Caughey *et al.*, 2009; Solomon *et al.*, 2009). Another possibility is that binding of PrPSc to PrPC triggers a signal transduction pathway leading to neuronal damage (Soto and Satani 2011). Other theories of PrPSc pathogenicity, based around *in vitro* observations, include impairment of breakdown of cellular waste by lysosomes, up-regulation of genes involved in endoplasmic reticulum function, and reduced degradation of proteins by the proteasome system (Kovacs and Budka 2008; Chakrabarti *et al.*, 2009).

## 2.7. Clinical signs and symptoms

### 2.7.1. Clinical signs in cattle

Field data suggest that susceptibility to BSE infection peaks around 12 months of age in cattle, although there have been BSE cases in cattle that were not fed meat-and-bone meal (MBM) until they were over 2 years of age. The incubation period in cattle is estimated to be from 30 months to 8 years (mean of 4.5–5.5 years), but the course of the clinical disease is short from the onset of clinical signs, with animals generally dying or requiring euthanasia within 6 months (USDA FSIS 2005; St Rose *et al.*, 2006; Harman and Silva 2009). Among 124,000 UK cattle for which the age of onset of clinical signs was known, 7% were 3 years old, 31% were 4 years old, 33% were 5 years old and 29% were 6 years old or older (Harman and Silva 2009). Clinical signs in cattle include changes in temperament, such as nervousness or aggression, abnormal posture, incoordination and difficulty in rising, decreased milk production, or loss of body weight despite continued appetite (USDA FSIS 2005; Seuberlich *et al.*, 2010).

### 2.7.2. Clinical signs in humans

Over 200 human cases of vCJD have been reported (Aguzzi and Calella 2009). Patients have ranged in age from 17–42 years (Brown and Mastrianni 2010). The great majority of patients were UK residents during the period 1985–1996 (Mackay *et al.*, 2011). Variant CJD is invariably fatal, and generally runs a course of approximately 18 months from the onset of symptoms. Symptoms and clinical signs include both cognitive and motor dysfunction. Early in the illness, patients usually experience psychiatric or sensory symptoms. Reported psychiatric symptoms include depression, apathy, agitation, insomnia, poor concentration, paranoid delusion, recklessness, aggression, withdrawal or anxiety. Approximately one third of the patients reported unusual persistent and painful sensory symptoms. Neurological signs develop as the illness progresses, including cerebellar ataxia, muscle spasms and involuntary movements. Late onset signs

include urinary incontinence, progressive immobility, and akinetic mutism. Death is often due to opportunistic infections (Imran and Mahmood 2011a). The great majority of cases of prion disease occurring in humans are spontaneous occurrences of sporadic Creutzfeldt-Jakob disease (sCJD). In comparison to vCJD, sCJD typically affects people between 55 and 70 years old (Mackay *et al.*, 2011). Cerebellar ataxia or progressive dementias predominate in the first few months of sCJD, which contrast with vCJD (Imran and Mahmood 2011a). Also, vCJD features distinctive histopathology (Mackay *et al.*, 2011). Besides classical BSE, the only other TSE known to be infectious to humans by the oral route is the now nearly extinct disease known as kuru, which occurred in a small group of communities of indigenous Papua New Guinean people who practised cannibalism (Aguzzi and Calella 2009).

### 2.7.3. Occurrence in food

The incidence of classical BSE in cattle has declined markedly since the 1990s through prevention measures based on knowledge of how the disease is transmitted between cattle. There are now very few cases of BSE reported in cattle worldwide (OIE 2013). A key component of prevention of both BSE in cattle and vCJD in humans is the prohibition on feeding ruminant derived protein to ruminants. This measure was enacted in 1994 in the UK when it became illegal to feed ruminants with mammalian proteins, with specific exceptions such as dairy proteins. The feed ban was further extended in 1996 by a ban on feeding any farmed livestock, including fish and horses, with mammalian MBM. Feeding of mammalian derived proteins was prohibited throughout the European Union (EU) in 1994 (EC No. 94/381).

This was further extended in EU Regulation (EC No. 999/2001) which introduced further EU-wide controls to combat the spread of BSE, including a ban on the feeding of processed animal proteins to any animals kept, fattened or bred for the production of food. With regards to infectivity in food, the risk of exposure of humans to BSE can essentially be removed by withholding the particular lymphoid and central nervous tissues known to harbour infectivity, termed specific risk materials (SRM), from the food supply. The list of SRM, and the ages of cattle from which they must be removed, have been modified over time with advancing knowledge, but currently include brain, eyes, tonsils, spinal cord and intestines, as well as the entire spinal column of older cattle. Bans on the use of SRM from bovine carcasses above specified ages have been implemented throughout Europe and other countries

to ensure the safety of beef and beef products. Slaughter procedures of cattle are designed to prevent contamination of the carcass with SRM, and SRM are rendered and incinerated to destroy infectivity in countries with BSE risk factors. In addition, mechanical recovery of meat from bones is prohibited in order to prevent inclusion of dorsal root ganglia, which may contain infectivity. Beef and beef products from countries with these and animal feed control systems are therefore considered to be safe for human consumption. Countries assessed as being of negligible risk of BSE in the cattle population are not required to practice these precautions.

## 2.8. Risk factors

### 2.8.1. Human host factors

A polymorphism at position 129 of the PrPC amino acid sequence has been identified in humans, with different genotypes exhibiting different susceptibilities to TSEs. Approximately 40% of Caucasians are homozygous for methionine (Met) at this position, 10% are homozygous for valine (Val) and 50% are Met/Val heterozygotes. To date, all confirmed clinical cases of vCJD have been homozygous for Met at codon 129 (Mackay *et al.* 2011). A presumptive clinical case in a Met/Val heterozygote was reported in 2009, but an autopsy was not done and the MRI findings were not typical of vCJD (Kaski *et al.* 2009; Mackay *et al.* 2011). A confirmed pre-clinical infection in a Met/Val heterozygote was identified in a patient who died of a ruptured aortic aneurysm five years after receiving a blood transfusion from a person who subsequently developed vCJD. The Met/Val heterozygote had PrPSc in the spleen but not in the central nervous system, and there was no evidence of central nervous pathology typical of vCJD (Gains and LeBlanc 2007; Mackay *et al.* 2011).

The three known clinical cases of vCJD infection by blood transfusion were all Met/Met homozygotes (Mackay *et al.* 2011). Two of three PrPSc-positive samples in an anonymous postsurgical study of appendices were from Val/Val homozygotes. This indicates that lymphoid tissue, at least, of all three genotypes may become infected (Harman and Silva 2009; Will 2010; Mackay *et al.* 2011). It is not yet clear whether the Met/Val and Val/Val genotypes are protective against neurological infection with vCJD, or whether onset is delayed rather than prevented (Mackay *et al.* 2011). Some authors have predicted a multiphasic vCJD epidemic with a late peak of cases affecting people heterozygous at codon 129 (Aguzzi and Calella 2009; Will 2010).

Besides vCJD, the only other orally acquired prion disease known in humans is kuru, a

historical disease of a small number of communities in Papua New Guinea who propagated the disease through the practice of funerary cannibalism. The mean incubation period of kuru is 12 years, but the incubation period has exceeded 50 years in some individuals (Imran and Mahmood 2011a).

Retrospective analysis of blood samples from kuru patients shows an age stratification of codon 129 genotype. The young kuru patients were mainly Met/Met or Val/Val homozygotes, whereas the elderly patients were mostly Met/Val heterozygotes. Eight of eleven of the more recent cases of kuru were Met/Val heterozygotes, which supports the hypothesis that the Met/Val genotype delays but does not prevent the onset of kuru in all individuals, because exposure of these individuals almost certainly ended more than 40 years ago when funerary cannibalism was outlawed (Mackay *et al.* 2011). The majority of vCJD cases in the UK affected people less than 40 years of age. Possible explanations include a higher rate of dietary exposure, increased susceptibility to infection or a reduced incubation period in this age group. Greater susceptibility could be conferred by the volume of GALT, which declines with age (Mackay *et al.* 2011). In addition, Peyer's patches that are thought to be involved in intestinal uptake of prions, decline during adulthood (St Rose *et al.* 2006).

## 2.9. Prevention and control

The control of BSE should be very simple; all that is needed is to prevent ruminant offal – brain and spinal cord particularly entering the animal feed chain being rendered and incorporated into feed for cattle. If that is done BSE cannot persist in a cattle population and vCJD will likewise not occur. It turns out this is easier said than done because although most countries in the world today (including southern African countries) have such legislation in place, enforcement is variable and sometimes perfunctory. In Africa the issue is, furthermore, contentious because indigenous cases of BSE have not so far been diagnosed in Africa and there is consequently a belief that it could not occur here. This belief belies the fact that cattle and possibly MBM were imported into some countries of the Region when the BSE epidemic in Europe was at its height. So the BSE agent could have been introduced and perpetuated because the controls in place in some southern African countries are inadequate (Thomson, 2010).

So making a sudden loud noise and watching the reaction of the animal concerned is a useful diagnostic aid. Inco-ordination is also a frequent early sign, especially of the hind limbs. Some animals may adopt a stance where the head is lowered and the neck extended with wide-based hind

legs. Hypermetria (over-shooting of intended limb position) is also a common feature of BSE. It should also be remembered that other disease and BSE may be concurrent (Konold *et al.*, 2006). More detailed analyses of the criteria for clinical suspicion are available (Bradley & Verwoerd, 2004b; Konold *et al.*, 2006). There are no macroscopically observable pathological lesions associated with BSE at any stage of the disease. In contrast to most other neurological diseases of cattle the course of BSE is often weeks or months long, i.e. before the animal becomes severely incapacitated or dies. This distinguishes BSE from infectious diseases such as rabies, heartwater, cerebral theileriosis and babesiosis, thrombotic meningio-encephalitis (*Haemophilus somnus*), sporadic bovine encephalomyelitis, cerebrocortical necrosis (thiamine deficiency), chemical and biological toxins (e.g. lead, botulism, urea, pesticides, mycotoxins, diploidosis & pesticides) and a range of poisonous plants. Non-infectious causes of BSE-like clinical signs include arthritis, lameness, joint trauma, recumbence (downer syndrome especially after falling on concrete), unsatisfactory milk yield and ear infection (Konold *et al.*, 2006).

### 3. Conclusions And Recommendations

The Bovine Spongiform Encephalopathy has got worldwide attention due to its transmissible nature to human beings. The risk from BSE or TSE infection had a major impact on trade in cattle; beef and bovine products in BSE infected countries. BSE is caused by a poorly understood type of infectious protein particle called 'Prion'. The 'Prion' particles are highly resistant to inactivation by physical as well as chemical agents. Prion is a modified form of a normal cellular protein known as PrPc. As there is no reliable treatment or vaccines of current concepts in the pathogenesis and bovine spongiform encephalopathy diseases so there is need to better understand the diagnosis of bovine spongiform encephalopathy. biology diseases, to develop strategies to manage prion disease outbreaks and minimize. Cause of mad cow disease: Suspected impacts and to apply learnings of prion diseases to the pathogens in bovine spongiform encephalopathy. There is also necessity for further research to clearly understand how bovine spongiform encephalopathy. The pathogenesis of prions causes no detectable immune or responses in the host need to be clarified and pathogenesis of scrapie and transmissible mink Conversion of a encephalopathy (TME) needs be explained. based on this the following recommendations are forwarded;

- Animals symptomatic for the disease are prohibited in animal food supplements, though this really goes without saying.
- Older animals, which will have experienced the most amplification be eliminated from animal food supplements.
- Animals with longer lifespans not be fed animal food supplements, especially in younger years.
- That effective separation procedure will be utilized when materials are fed both within and between species.
- Those quantities of animal food supplements must be reduced.
- further research to clearly understand how Bovine spongiform encephalopathy transmit must be done.

### 4. REFERENCES

1. Aguzzi A, Calella AM (2009). Prions Protein aggregation and infectious diseases and amplification of bovine spongiform encephalopathy PrPSc and enable ultrasensitive detection of bovine PrPSc. *PLoS ONE*; **5(10)**:13152.
2. Anderson RM, Donnelly CA, Ferguson NM, Woolhouse MEJ (1996). Transmission dynamics and epidemiology of BSE in British cattle. *Nature*; **382**:779-788.
3. Arnold ME, Hawkins SAC, Green R, Dexter I, Wells GAH (2009). Pathogenesis of experimental bovine spongiform encephalopathy (BSE): Estimation of tissue infectivity according to incubation period. *Veterinary Research*; **40**:08.
4. Baron T, Bencsik A, Morignat E (2010). Prions of ruminants show distinct splenotropisms in an ovine transgenic mouse model. *Biochemistry*; **48(12)**:2574-2585.
5. Bradley R & Verwoerd DW (2004a). Unclassified virus-like agents, transmissible spongiform encephalopathies and prion diseases. In: *Infectious Diseases of Livestock*, Vol.2. J.A.W. Coetzer & R.C. Tustin (eds.), pp 1388-1390. Oxford University Press: Cape Town, Oxford.
6. Bradley R & Verwoerd DW (2004b). Bovine spongiform encephalopathy. In: *Infectious Diseases of Livestock*, Vol.2. J.A.W. Coetzer & R.C. Tustin (eds.), pp 1408-1421. Oxford University Press: Cape Town, Oxford.
7. Bradley R & Verwoerd DW (2004). Transmissible spongiform encephalopathies related to bovine spongiform encephalopathy in other domestic and captive wildlife species. In: *Infectious Diseases of Livestock*, Vol.2.



- J.A.W. Coetzer & R.C. Tustin (eds.), pp 1422-1424. Oxford University Press: Cape Town, Oxford.
8. Brown K, Mastrianni JA (2010). The prion diseases *Journal of Geriatric Psychiatry Canadian Journal of Neurological Sciences*; **34**:126–145.
  9. Caughey B, Baron GS, Chesebro B, Jeffrey M (2009). Getting a grip on prions: Oligomers, amyloids and pathological membrane interactions. *Annual Review of Biochemistry*; **78**:177–204.
  10. Chakrabarti O, Ashok A, Hegde RS (2009). Prion protein biosynthesis and its emerging role in neurodegeneration. *Trends in Biochemical Sciences*; **34**(6):287–295.
  11. Colby DW & Prusiner SB (2011). Prions Cold Spring Harbor perspectives in biology, 3, a006833.
  12. Collinge J (2012). The risk of prion zoonoses *biomedical Science*; **335**:411–413.
  13. Ducrot C, Arnold M, de Koeijer A, Heim D, & Calves D (2008). Review of the epidemiology and dynamics of BSE epidemics. *Veterinary Research*; **39**:15.
  14. encephalopathies in ruminants: A challenge for disease surveillance and control. *Journal of Veterinary Diagnostic Investigation*; **22**:823–842.
  15. feeding them with extraneural tissues of affected cattle. *Vet. Rec.* 1993 *132*: 545-547.
  16. Fukuda S, Onoe S, Yokoyama T, Mohri S (2010) Sulfated dextrans enhance *in vitro*
  17. Gains MJ, LeBlanc AC (2007). Prion protein and prion disease: The good and the bad.
  18. Goni F, Mathiason C K, Yim L, Wong K, Hayes-Klug J, Nalls A, Wisniewski T (2015). Mucosal immunization with attenuated *Salmonella* vaccine partially protects white-tailed deer from chronic wasting disease. *Vaccine*; **33**(5); 726–733.
  19. Gough KC, Maddison BC (2010). Prion transmission: Prion excretion and occurrence in the environment. *Prion*; **4**(4):275–282.
  20. Griffiths SJ (1967). Nature of the scrapie agent: Self-replication and scrapie. *Nature*; **215**: 1043-1044.
  21. Haley NJ, Mathiason CK, Carver S, Zabel M, Telling GC, Hoover EA (2011). Detection of chronic wasting disease prions in salivary, urinary, and intestinal tissues of deer: Potential mechanisms of prion shedding and transmission. *Journal of Virology*; **85**(13):6309–6318.
  22. Harman JL, Silva CJ (2009). Bovine spongiform encephalopathy. *Journal of the American*
  23. Hoffmann C, Eiden M, Kaatz M, Keller M, Ziegler U, Rogers R, Hills B, Balkema-Buschmann A, van Keulen L, Jacobs JG, Groschup MH (2011). BSE infectivity in
  24. Hoffmann C, Ziegler U, Buschmann, A, Weber A, Kupfer L, Oelschlegel A (2007). Prions spread via the autonomic nervous system from the gut to the central nervous system in cattle incubating bovine spongiform encephalopathy. *Journal of General Virology*; **88**: 1048-1055.
  25. [http://www.fsis.usda.gov/Factsheets/Bovine\\_Spongiform\\_Encephalopathy\\_Mad\\_Cow\\_Disease/index.asp#10](http://www.fsis.usda.gov/Factsheets/Bovine_Spongiform_Encephalopathy_Mad_Cow_Disease/index.asp#10). Accessed 9 August 2012
  26. [http://www.hpa.org.uk/webw/HPAweb&HPAwebStandard/HPAweb\\_C/1234859690542?p=1231252394302](http://www.hpa.org.uk/webw/HPAweb&HPAwebStandard/HPAweb_C/1234859690542?p=1231252394302). Accessed 9 May 2013
  27. Hueston W, Bryant CM (2005). Transmissible spongiform encephalopathies. *Journal of Food Science*; **70**(5):77–87
  28. Imran M, Mahmood S (2011a). An overview of human prion diseases. *Virology Journal*; **8**:559
  29. Imran M, Mahmood S (2011b). An overview of animal prion diseases. *Virology Journal* 8:493. Isolation of prion with BSE properties from farmed goat. *Emerging Infectious Diseases*, 17. [wwwnc.cdc.gov/eid/article/17/12/11-0333\\_article.htm](http://wwwnc.cdc.gov/eid/article/17/12/11-0333_article.htm). jejunum, ileum and ileocaecal junction of incubating cattle. *Veterinary Research*; **42**:21
  30. Kaski D, Mead S, Hyare H, Cooper S, Jampana R, Overell J, Knight R, Collinge J, Rudge P (2009). Variant CJD in an individual heterozygous for *PRNP* codon 129. *The Lancet*; **374**:2128.
  31. Konold T, Bone GE, Clifford D, Chaplin MJ, Cawthraw S, Stack MJ, Simmons MM (2012). Experimental H-type and L-type bovine spongiform encephalopathy in cattle: Observation of two clinical syndromes and diagnostic challenges. *BMC Veterinary Research*; **8**:22.
  32. Konold T, Sivan S.K, Ryan J, Gubbins S, Laven R & Howe JH (2006). Analysis of clinical signs associated with bovine spongiform encephalopathy in casualty slaughter cattle. *The Veterinary Journal*; **171**: 438-444.
  33. Kovacs GG, Budka H (2008). Prion diseases: From protein to cell pathology. *The American Journal of Pathology*; **172**(3):555–565.

34. Lasmezas CI, Deslys JP, Robain O, Jaegly A, Beringue V, Peyrin JM, Dormont D (1997). Transmission of the BSE agent to mice in the absence of detectable abnormal prion protein. *Science*; **275**:402–405.
35. Lewis PA, Tattum MH, Jones S, Bhalt D (2006). Codon 129 polymorphism of the human prion protein influences the kinetics of amyloid formation. *Journal of General Virology*; **87**:2443-2449.
36. Linden R, Martins VR, Prado MAM, Cammarota M, Izquierdo I, Brentani RR (2008)
37. Mackay GA, Knight RSG, Ironside JW (2011). The molecular epidemiology of variant CJD. *International Journal of Molecular Epidemiology and Genetics*; **2(3)**:217–227.
38. Masujin K, Mathews D, Wells GAH, Mohri S & Yokoyama T (2007). Prions in the peripheral nerves of bovine spongiform encephalopathy-affected cattle. *Journal of General Virology*; **88**: 1850-1858.
39. Middleton DJ & Barlow RM: Failure to transmit bovine spongiform encephalopathy to mice by
40. mortem. Public Health England, London.
41. Murayama Y, Yoshioka M, Masujin K, Okada H, Iwamaru Y, Imamura M, Matsuura Y,
42. OIE (2013). BSE situation in the world and annual incidence rates. World Organisation for Animal Health, Paris. <http://www.oie.int/en/animal-health-in-the-world/bse-specific-data/>. Accessed 24 May 2013.
43. PHE (2009). vCJD abnormal prion protein found in a patient with haemophilia at post *Physiological Reviews*; **89**:1105–1152. Physiology of the prion protein. *Physiological Reviews*; **88**:673–728. prion diseases (dog, rabbit and horses). Ch 2 In: Verdier JM (ed) Prions and prion diseases: New developments. NOVA Science Publishers, New York p. 41-48
44. Prusiner SB (1982) Novel proteinaceous infectious particles cause scrapie. *Science*
45. Prusiner SB (1994). Biology and genetics of prion diseases. *Annual Reviews in Microbiology*; **48**: 655–686.
46. Reik, R., Hornemann, S., Wider, G., Billeter, M., Clockshuber, R., Wüthrich, K,( 1996).
47. Sajjani G, Silva CJ, Ramos A, Pastrana M A, Onisko BC, Erickson ML, Requena JR (2012). PK-sensitive PrP is infectious and shares basic structural features with PK-resistant PrP. *PLoS Pathogens*, **8**, e1002547.
48. Sakudo A, Ano Y, Onodera T, Nitta K, Shintani H, Ikuta K, Tanaka Y (2011). Fundamentals of prions and their inactivation. *International Journal of Molecular Medicine*; **27**:483–489.
49. Seuberlich T, Heim D, Zurbriggen A (2010). Atypical transmissible spongiform
50. Solomon IH, Schepker JA, Harris DA (2009). Prion neurotoxicity: Insights from prion protein mutants. *Current Issues in Molecular Biology*; **12**: 51–62.
51. Soto C, Satani N (2011). The intricate mechanisms of neurodegeneration in prion diseases. *Trends in Molecular Medicine* ;**17(1)**:14–24.
52. Spiropoulos J, Lockey R, Sallis RE, Terry LA, Thorne L, Holder TM, Beck KE, Simmons MM (2011). Isolation of prion with BSE properties from farmed goat. *Emerging Infectious Diseases* ;**17(12)**:2253–2261.
53. St Rose SG, Hunter N, Matthews L, Foster JD, Chase-Topping ME, Kruuk LEB, Shaw DJ, Rhind SM, Will RG, Woolhouse MEJ (2006). Comparative evidence for a link between Peyer's patch development and susceptibility to transmissible spongiform encephalopathies. *BioMedCentral Infectious Diseases*; **6(5)**: 1471-2334.
54. Taylor K (1995): Origin of BSE. *Vet. Rec*; **137(26)**: 674-675.
55. Thomson GR (2010). Qualitative risk assessment of the presence and persistence of the BSE agent in the cattle population of one or more countries of the SADC Region. *GOPA Worldwide Consultants, Rural Development & Environment, Bad Homburg, Germany.*
56. USDA FSIS (2005). Bovine spongiform encephalopathy"Mad cow disease". *USDA Food Safety Inspection Scheme, Washington D.C.*
57. van Keulen LJM, Bossers A, van Zijderveld F (2008). TSE pathogenesis in cattle and sheep. *Veterinary Research* **39**:24. *Veterinary Medical Association*; **234(1)**:59–72.
58. Wells GAH, Dawson M, Hawkins SAC, Green RB, Dexter I, Francis ME, Simmons MM, Austin AR, Horigan MW (1994). Infectivity in the ileum of cattle challenged orally with bovine spongiform encephalopathy. *Vet. Rec*; **135**: 40-41.
59. Wells GAH, Spiropoulos J, Hawkins SAC, & Ryder, SJ (2005). Pathogenesis of experimental bovine spongiform encephalopathy: Preclinical infectivity in tonsil and observations on the distribution of

- 
- lingual tonsils in slaughtered cattle. *Veterinary Record*; **156**: 401-407.
60. [WHO, 2008]. World health organization, bovine spongiform encephalopathy,” <http://www.who.int/zoonoses/diseases/bse/en/> (Accessed 03/06/08); **216**:136–144. 23(4):277–298.
61. Will B (2010). Variant CJD: Where has it gone, or has it? *Practical Neurology*; **10**:250–251.
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