



Performance, Haematology And Serum Biochemical Parameters Of Growing Grass Cutters (*Thyronoymys Swinderianus*) Fed *Phyllanthus Amarus* And *Pilogstigma Thonngii* Leaf Meal Mixture As Partial Replacement For Soya Bean Meal

^{1*}Alagbe, J.O., ²Agubosi, O.C.P and ³Liu, S.D

^{1,3} Sumitra Research Institute, Gujarat, India

² Department of Animal Science, University of Abuja, Nigeria

*Orcid number: 0000-0003-0853-6144; demsonfarms@yahoo.com

*Corresponding Author

Abstract: The objective of the present study was to evaluate the performance, haematology and serum biochemical parameters of growing grasscutters (*Thrynomys swinderianus*) fed *Phyllanthus amarus* (PAM) and *Pilogstigma thonngii* leaf meal (PTM) mixture as partial replacement for soya bean meal. A total of 35 weaned grasscutters of mixed sex between 5-6 weeks with an average weight of 436.1 and 437.0 g were divided into five groups of 7 animals each and randomly assigned to 5 experimental diets each animal served as a replicate in a completely randomized design. The dietary treatments include a control diet with no PATML, T2 (10.0% PATML), T3 (20.0 % PATML), T4 (30.0 % PATML) and T5 (40.0 % PATML) respectively. Feed and water were offered *ad libitum* throughout the experiment which lasted for 12 weeks. The data obtained was used to evaluate the growth, haematological parameters (PCV, RBC, Hb, MCV, MCH, MCHC, WBC and its differentials), serum biochemical indices (Albumin, globulin, total protein, creatinine, cholesterol, calcium, phosphorus, ALP, AST and ALT) and fatty acid composition. Average weight gain, average daily feed intake and feed conversion ratio were influenced by the dietary treatments ($P < 0.05$). Haematological and serum parameters were significantly ($P < 0.05$) different among the treatments and were within the normal physiological range for grasscutters. It could be concluded that replacement of soya meal with 40 % PATML does not have any deleterious effect on the performance and blood profile of the animal.

[Alagbe, J.O., Agubosi, O.C.P and Liu, S.D. **Performance, Haematology And Serum Biochemical Parameters Of Growing Grass Cutters (*Thyronoymys Swinderianus*) Fed *Phyllanthus Amarus* And *Pilogstigma Thonngii* Leaf Meal Mixture As Partial Replacement For Soya Bean Meal.** *Biomedicine and Nursing* 2021;7(3):21-31]. ISSN 2379-8211 (print); ISSN 2379-8203 (online). <http://www.nbmedicine.org> 4. doi:[10.7537/marsbnj070321.04](https://doi.org/10.7537/marsbnj070321.04).

Keywords: Weaner grasscutters; *Phyllanthus amarus*; *Pilogstigma thonngii*; haematology

INTRODUCTION

Success in livestock production is impossible without giving enough attention to nutrition and improving wholesome feed in adequate quantity daily. In fact, nutrition is one of the important cardinals of management, but scarcity of feed resources has been a hindrance in the production of livestock products to meet the animal protein requirements of human and other industrial needs (Unigwe *et al.*, 2016). Onu *et al.* (2004) reported that protein intake in developing countries is low due to high cost of feeding with the accompanying high cost of production. To achieve breakthrough in production and maximize profit there is a need to employ the use of agricultural by products and plants that are cheaper, safe and available to feed animals (Alagbe, 2016). Some plants have been reported to perform multipurpose roles of supplying proteins, vitamins, minerals and as a phytobiotics (Alagbe and Omokore, 2019). Among the potential

underexplored plants are *Phyllanthus amarus* and *Pilogstigma thonngii*.

Grass cutter (*Thrynomys swinderianus*) is a wild herbivores rodent, which lives mainly on grass and other succulent forages (Wogar and Agwunobi, 2011). It is an explored in most areas of Africa as a source of animal protein and it significantly contributes to both local and foreign earnings of most African countries (Asibey, 1974). Feeds containing 2-20 % crude protein have been reported to suitable for grass cutters (Meduna, 2002); they are prolific and contain low cholesterol in their meat which makes them safe for consumption.

Phyllanthus amarus belongs to the family of Euphobiaceae. It is commonly found in forest areas, arid land, savannah areas, leached and exhausted soil in many countries including India, China, Cuba and Philippines among others (Zubair *et al.*, 2016; Burkill, 1994). Omokore and Alagbe (2019) reported that

Phyllanthus amarus leaves contained dry matter (91.2 %), crude fibre (15.11 %), crude protein (10.22 %), ether extract (5.88 %), ash (9.11 %) and nitrogen free extract (50.80%). Reports on the plant have revealed that the various parts are loaded with bioactive chemicals or secondary metabolites (Zubair *et al.*, 2016) which confers it the ability to perform several biological activities such as anti-inflammatory functions (Adeolu and Sunday, 2013), antimicrobial (Evi *et al.*, 2011), anti-bacterial (Dada *et al.*, 2014), antioxidant (Chandan *et al.*, 2012), antidiabetic (Khartoon *et al.*, 2004; Oluwafemi and Debiri, 2008), antiviral (Prasad *et al.*, 2013), anticonvulsant (Manikkoth *et al.*, 2011) and anticancer (Rajashkumar *et al.*, 2002).

Piliostigma thonningii belongs to the family Caesalpiniaceae. It has large two-lobed simple leaves and without thorns or spines and perennial in nature (Jimoh and Oladiji, 2005). Phytochemical screening of the leaves revealed the presence of phenol, alkaloids, saponin, flavonoids and steroids as secondary metabolites (Jimoh and Oladiji, 2005). Alagbe *et al.* (2019) reported that the leaves of *Piliostigma thonningii* contain dry matter, ether extract, crude protein, crude fibre and ash at 91.20 %, 0.31%, 11.21 %, 14.22% and 7.22 % respectively. Ighodaro *et al.* (2012) also evaluated the leaves to have 10.09 % crude protein, 2.81 % ether extract, 6.10 % ash, 5.23 % crude fibre and 72.17 % carbohydrates. The plant parts (seed and leaves) play a vital role as an antimicrobial (Lina, 2017), antiviral (Nakayama *et al.*, 1993), antifungal (Oyagade, 1999), antibacterial (Salazar *et al.*, 2011), anti-inflammatory (Sofowora, 1993), antihelminthic (Tasheen and Mishra, 2013), anticancer (Isidorov *et al.*, 2008) and antidiuretic (Zhang, 2014).

Previous studies have shown that administration of *Phyllanthus amarus* at 8 % in the diet of rabbit significantly improved the growth of rabbits (Omokore and Alagbe, 2019). Similarly, administration of *Piliostigma thonningii* leaf extract at 0.4g/kg significantly ($P>0.05$) reduce blood cholesterol level in wister rats (Akinpelu and Obutor, 2000). Nutrients are known to influence the responses of livestock to a disease challenge (Gary and Richard, 2002) and blood plays an important role in the transportation of nutrients, metabolic waste product and gases around the body (Tavares *et al.*, 2004). Both *Piliostigma thonningii* and *Phyllanthus amarus* leaves are loaded with various bioactive chemicals when could synergistically work together to improve performance and blood parameters of animals.

Therefore, this study was designed to evaluate the performance, haematology and serum biochemical parameters of growing grasscutters (*Thryonomys swinderianus*) fed *Phyllanthus amarus* and *Pilogstigma*

thonngii leaf meal mixture as partial replacement for soya bean meal

MATERIALS AND METHODS

Site of the experiment

The experiment was carried out at Division of Animal Nutrition, Sumitra Research Institute, Gujarat, India during the month of December to February, 2019.

Source and processing of test materials

Fresh and mature leaves of *Piliostigma thonningii* and *Phyllanthus amarus* leaves were purchased from a local market in Gujarat and authenticated at biological science department of the research farm. It was air dried under shade to reduce moisture content until they were crispy to torch. The leaves thereafter were crushed separately with hammer mill to form *Piliostigma thonningii* leaf meal (PTM) and *Phyllanthus amarus* leaf meal (PAM) respectively. The samples were later mixed in ratio of 1:1 to form PATML. The samples were kept in an air tight container for further analysis.

Pre-experimental operations

Prior to the commencement of the experiment the hutches were disinfected two weeks before the arrival of the animals. Cement feeders, drinkers, foot deep at the entrance of the pens were properly cleaned and kept in the store. A separate cage was also prepared to accommodate sick or culled animals.

Experimental animals and management

Thirty five (35) weaned grasscutters of mixed sex between 5-6 weeks with an average weight of 436.1 and 437.0 grams were used for the experiment. The animals were randomly divided into five groups of seven grasscutter per replicate and each animal served as a replicate in a completely randomized design. Grass cutters were allowed two weeks adjustment period during which they were fed with basal diet (morning and evening), housed in an all wired cage measuring 45×53×40 cm and given prophylactic treatment with Oxytetracycline administered intramuscularly and Ivermectin given subcutaneously adhering strictly to the package insert. Animals were fed twice daily between 7:30 am and 3:30 pm (110 – 130 g each). Fresh feed and water were provided ad libitum and all other management practices were strictly observed.

Diet formulation and chemical analysis

The basal diet was formulated to meet the nutrient requirements of weaner grasscutters according to Adeniji (2009).

Treatment 1 (Control): Basal diet + 0 % PATML

Treatment 2: Basal diet + 10.0 % PATML

Treatment 3: Basal diet + 20.0 % PATML

Treatment 4: Basal diet + 30.0 % PATML

Treatment 5: Basal diet + 40.0 % PATML

Proximate analysis of experimental diet was determined using methods described by AOAC (2000). Phytochemical screening of PTM and PAM were analyzed according to methods outlined by Harbone (1973); Trease and Evans (1983). Mineral analyses were carried out using Atomic Absorption Spectrophotometer (AAS) model 12-0TA.

Measurements

✓ Performance parameters

Feed intake (g) was determined by subtracting feed left over from feed served, it was estimated for each of the replicate daily.

Weight gain (g) was calculated by finding the difference between initial weight and final weight at the end of the experiment.

Feed: gain ratio was determined from the average feed consumed by the average weight gained in each treatment.

Mortality rate was recorded as it occurs daily.

✓ Blood collection and analysis

At the end of the 12th week of the experiment, blood samples were collected by cardiac puncture from four randomly selected grass cutters per treatment. The operation is done very early in the morning to reduce

stress on the animals, 6ml each was collected from each animal. 3 ml of blood sample was transferred into a sterile EDTA bottle for hematological assay. The parameters (Pack cell volume, haemoglobin concentration, red blood cell, mean corpuscular volume, mean corpuscular haemoglobin concentration, white blood cell and its differentials) were analyzed using Diasis Diagnostic Systems ASI-671N, India (an automated digital analyzer).

The remaining 3 ml was put into another bottle without anticoagulant (EDTA) for serum biochemical analysis. Albumin, globulin, cholesterol and total glucose, creatinine, calcium, sodium and chlorine ions were analyzed using Randox ® commercial kits, USA (Model 2R-TR4). Activities of Alanine transaminase (ALT), Aspartate transaminase (AST) and Alkaline phosphatase (ALP) were recorded according to (Doumas and Briggs, 1972).

Statistical analysis

All data were subjected to one -way analysis of variance (ANOVA) using SPSS (23.0) and significant means were separated using Duncan multiple range tests (Duncan, 1955). Significant was declared if $P \leq 0.05$.

Table 1. Chemical composition of experimental diets

Ingredients	T1	T2	T3	T4	T5
Maize	40.00	40.00	40.00	40.00	40.00
Wheat offal	25.00	25.00	25.00	25.00	25.00
PKC	11.10	11.10	11.10	11.10	11.10
Soya meal	20.00	18.00	16.00	14.00	12.00
PATML	0.00	2.00	4.00	6.00	8.00
Bone meal	2.00	2.00	2.00	2.00	2.00
Limestone	1.00	1.00	1.00	1.00	1.00
Lysine	0.20	0.20	0.20	0.20	0.20
Methionine	0.20	0.20	0.20	0.20	0.20
Premix	0.25	0.25	0.25	0.25	0.25
Salt	0.25	0.25	0.25	0.25	0.25
Total	100.00	100.00	100.00	100.00	100.00
Cal analysis (%)					
Dry matter	86.87	87.10	88.61	88.92	88.95
Crude protein	19.02	18.51	18.38	18.24	18.10
Crude fibre	7.62	7.78	7.94	8.10	8.26
Ether extract	3.89	3.78	3.67	3.56	3.46
Calcium	0.99	1.09	1.19	1.29	1.39
Phosphorus	0.41	0.47	0.47	0.58	0.63
Energy (Kcal/kg)	2754.0	2750.2	2755.0	2759.0	2761.0

* Premix supplied per kg diet :- Vit A, 10,000 I.U; Vit E, 5mg; Vit D3, 3000I.U, Vit K, 3mg; Vit B2, 5.5mg; Niacin, 25mg ; Vit B12, 16mg ; Choline chloride, 120mg ; Mn, 5.2mg ; Zn, 25mg ; Cu, 2.6g ; Folic acid, 2mg ; Fe, 5g ; Pantothenic acid, 10mg ; Biotin, 30.5g ; Antioxidant, 56mg

Table 2 Proximate composition of PATML

Parameters (%)	PATML
Dry matter	89.73
Crude protein	37.21
Crude fibre	16.22
Ether extract	1.31
Ash	10.22
Energy	2510.5

PATML: *Piliostigma thonningii* and *Phyllanthus amarus* leaf meal mixture

Table 3 Mineral composition of PATML

Parameters (mg/100g)	PATML
Calcium	100.04
Phosphorus	78.02
Potassium	30.56
Sodium	11.31
Magnesium	4.07
Zinc	5.02
Copper	2.93
Iron	1.81
Selenium	2.00
Manganese	1.88

Table 4 Phytochemical analysis of PATML

Parameters	PATML	*Permissible range (%)
Flavonoids	29.67	36.11
Phenol	15.08	20.01
Alkaloids	3.05	3.50
Tannins	3.38	11.50
Saponin	4.88	7.02
Glycosides	1.33	-
Steroids	1.02	1.30

*Alagbe, J.O (2019)

Table 5 Performance characteristics of weaner grasscutters fed different levels of PATML

Parameters	T1	T2	T3	T4	T5	SEM
Initial weight (g)	437.0	436.1	436.9	436.5	436.1	0.29
Final weight (g)	1000.4 ^c	1193.0 ^c	1200.4 ^b	1291.5 ^b	1300.1 ^a	3.11
Weight gain (g)	563.0 ^c	756.9 ^b	763.5 ^b	855.0 ^a	864.0 ^a	2.93
Feed intake (g)	3500.2 ^a	3300.6 ^b	3300.5 ^b	3300.1 ^b	3300.0 ^b	2.04
A.F.I (g)	50.15 ^a	47.15 ^b	47.10 ^b	47.05 ^b	47.02 ^b	0.06
Feed: gain	5.86 ^a	4.36 ^b	4.32 ^b	3.86 ^c	3.82 ^c	0.01
Mortality	1	-	-	-	-	-

Means in the same row with different superscripts are significantly different ($P < 0.05$)

A.F.I = Average feed intake ; SEM= standard error of mean

Table 6 Haematological parameters of weaner grasscutters fed different levels of PATML

Parameters	T1	T2	T3	T4	T5	SEM
PCV (%)	36.71 ^b	41.04 ^a	43.12 ^a	43.57 ^a	44.10 ^a	0.42
Hb (g/dl)	9.88 ^c	11.44 ^b	12.33 ^b	14.08 ^a	14.53 ^a	1.10
RBC ×10 ⁶ /μl	4.94 ^c	5.03 ^b	5.18 ^b	5.63 ^b	6.72 ^a	0.10
MCV (fl)	71.22 ^b	90.21 ^a	92.56 ^a	92.67 ^a	93.41 ^a	2.33
MCH (pg)	24.51 ^b	25.43 ^b	26.04 ^b	27.11 ^a	27.83 ^a	1.26
MCHC (g/dl)	28.34 ^b	28.67 ^b	29.60 ^b	30.18 ^a	31.60 ^a	0.51
WBC ×10 ⁶ /μl	2.41 ^b	2.62 ^b	2.74 ^b	2.88 ^b	3.04 ^a	0.31
Lymphocytes (%)	29.45 ^c	31.44 ^b	33.60 ^b	33.71 ^b	34.04 ^a	5.32
Monocytes (%)	0.06 ^b	1.01 ^a	1.44 ^a	1.56 ^a	1.82 ^a	0.03
Neutrophils (%)	41.23 ^b	52.06 ^a	53.44 ^a	55.89 ^a	56.02 ^a	4.41
Eosinophils (%)	0.74 ^b	1.22 ^a	1.45 ^a	1.61 ^a	1.87 ^a	0.01

Means in the same row with different superscript are significantly different ($P < 0.05$)

RBC, red blood cell; WBC, white blood cell; MCV, mean corpuscular volume; MCHC, mean corpuscular haemoglobin concentration; MCH, mean corpuscular haemoglobin

Table 7 Serum biochemical parameters of weaner grasscutters fed different levels of PATML

Parameters	T1	T2	T3	T4	T5	SEM
Albumin(g/dl)	1.74 ^b	2.09 ^a	2.11 ^a	2.17 ^a	2.30 ^a	0.17
Globulin (g/dl)	3.11 ^b	3.78 ^b	3.96 ^b	4.00 ^a	4.02 ^a	0.06
T.PRO (g/dl)	4.85 ^c	5.87 ^b	6.07 ^a	6.17 ^a	6.32 ^a	1.01
CHOL. (mg/dl)	109.2 ^a	100.2 ^a	99.61 ^b	90.88 ^b	88.45 ^c	6.71
GLU (mg/dl)	63.10 ^c	73.21 ^b	81.44 ^a	87.80 ^a	89.34 ^a	4.02
Creatinine (mg/dl)	0.09 ^b	1.82 ^a	1.31 ^a	1.24 ^a	1.05 ^a	0.07
Sodium (mmol/l)	109.4 ^c	121.5 ^b	128.9 ^b	134.8 ^a	135.1 ^a	10.02
P (mmol/l)	2.01 ^b	3.03 ^a	3.18 ^a	3.03 ^a	3.00 ^a	1.14
Chlorine (mmol/l)	89.81 ^b	93.10 ^a	97.83 ^a	98.44 ^a	98.09 ^a	5.41
ALP (u/l)	73.44 ^a	69.44 ^b	65.32 ^b	60.15 ^b	56.30 ^c	3.56
AST (u/l)	63.91 ^a	57.10 ^b	51.50 ^b	44.72 ^c	41.29 ^c	2.19
ALT (u/l)	40.34 ^a	40.01 ^a	33.40 ^b	29.67 ^c	28.12 ^c	1.23

Means in the same row with different superscripts differ significantly ($P < 0.05$)

ALP, alanine phosphatase; AST, alanine serum transaminase; ALT, alanine transaminase; CHOL, cholesterol; P, phosphorus; T.PRO, total protein; GLU, glucose.

RESULTS AND DISCUSSION

Table 2 shows the chemical composition of PATML. The chemical components of *Piliostigma thonningii* and *Phyllanthus amarus* leaf meal mixture (PATML) used for this study were 90.73 %, 37.21 %, 16.22 %, 1.31 %, 10.22 % and 2510.5 (Kcal/kg) for dry matter (DM), CP, CF, EE, ash and energy respectively. This crude protein and crude fibre value obtained in this study is higher than those of *Telfaria occidentalis* 31.49 % and 9.80 % respectively (Olanipekun et al., 2016). PATML is high in protein above 20%; it can therefore be used as a protein supplement in grass cutters (NRC, 1994). The ether extract and ash content is similar to those of *Momordica charantia* and *Enantia chlorantha* leaves reported by Olanipekun et al. (2016). Higher fibre in PATML can increase digestion in animals, lower the serum cholesterol level and risk of coronary heart disease (Alinnor et al., 2011). The

energy level of PATML is lower than the value reported by Obikaonu et al. (2011) for neem leaves.

The mineral composition of PATML is presented in Table 3. The leaf meal contained calcium, potassium, phosphorus, sodium, zinc, iron, magnesium, manganese, selenium and copper at 100.04, 30.56, 78.02, 11.31, 5.02, 1.81, 4.07, 2.88, 2.00 and 2.93 (mg/100 g). Ca > P > K > Mg > Zn > Cu > Fe > Na > Se > Mn. The leaves are abundant in calcium which plays a key role in constitution of biological systems; it also provides strong bones to the animal (Ibrahim et al., 2011). Phosphorus provides energy for the breakdown of carbohydrates, protein and fats; also needed for growth, maintenance and repair of tissues and cells and for the production of DNA and RNA (Akpanyung, 2005). Alagbe (2020) reported a higher calcium level of 101.17 mg/100g and potassium level (71.62 mg/100g) in dried *Lagenaria brevilifera* fruit.

According to Ojewuyi *et al.* (2015) potassium is responsible for the regulation of water and electrolyte in the body. Ajibade and Fagbohun (2010) reported that magnesium is necessary for major biological processes, including the production of cellular energy and the synthesis of nucleic acids and proteins. Iron combines with myoglobin to transport nutrients within the body and enhance the normal functioning of the central nervous system (Moyo *et al.*, 2011). Sodium is needed for proper fluid balance, nerve transmission, and muscle contraction; zinc is a component of many enzymes; needed for making protein and genetic material (Ross, 2005).

The phytochemical constituent of PATML is presented in Table 4. The leaf meal contained the following phytochemical components 3.05 mg/100 g alkaloids, 29.67 mg/100g flavonoids, 15.08 mg/100 g phenols, 3.38 mg/100 g tannins, 4.88mg/100 g saponin, 1.02 mg/100 g steroids and 1.33 mg /100 g glycosides. Flavonoids had the highest value while steroids had the least value. Phytochemicals are secondary metabolites or a bioactive chemical which performs multiple biological functions such as antimicrobial, antiviral, anti-inflammatory, anti-helminthic, antidiuretic and antioxidant effect (Liu *et al.*, 2012). The concentration of phytochemicals in plants are not the same for all species, it depends on the extraction method, storage condition, environmental factors, storage conditions and geographical origin (Lillehoj and Lee, 2012; Alagbe and Oluwafemi, 2019). Flavonoids are used as adjuvants in vaccine production and have the ability to scavenge free radicals (Allan and Miller, 1996). Saponin inhibits the growth of gram +ve and gram -ve bacteria (Min *et al.*, 2005) and also performs antiprotozoal role (Wallace *et al.*, 1994). A phenol act as antioxidant and prevents the risk of diseases (Hollman, 2001). Tannins have been shown to possess antimicrobial, antiviral and antibacterial activity (Redondo *et al.*, 2014). Alkaloid exhibits cytotoxic effect and growth inhibition against a variety of all making them have anti-inflammatory and anti-plasmodic properties (Faizi *et al.*, 2008). However, all the phytochemical components were within the safety level recommended by Alagbe and Oluwafemi (2019).

The performance characteristics of weaner grasscutters fed PATML as partial replacement for soyameal is shown in Table 5. Final live weight (LW) and weight gain (WG) range between 1000.4 – 1300.1 g and 563.0 – 864.0 g respectively. They were highest in T4 and T5, intermediate in T2 and T3 and lowest in T1 ($P<0.05$). The result obtained is in agreement with the findings of Omer *et al.* (2012) who observed a positive effect on the growth of rabbits fed diet mixed with 0.5 % oregano leaves but contrary to the reports of Alagbe (2013) when weaner grasscutters were fed wild sunflower (*Tithonia diversifolia*) at 20 %. Total feed

intake (TFI), average feed intake (AFI) and feed: gain range between 3300 – 3500.2 g, 47.02 – 50.15 g and 3.82 – 5.86 respectively. There is a significant ($P<0.05$) difference among the treatments in terms of the feed intake and feed:gain. According to Frankič *et al.* (2009) plants have the ability to regulate feed intake and stimulate digestive secretions due to the presence of phytochemicals in PATML. This agrees with the report of Srinivasan (2005) who concluded in his experiment that supplementation of spices such as black pepper, turmeric and clove at 0.5% in feed of rabbit is capable of stimulating the secretion of pancreatic enzymes and also increase the digestive enzymes of gastric mucosa. One (1) mortality was recorded in T1 and none was recorded in other treatments, this could be attributed to the presence of bioactive chemicals or phytochemicals in PATML. According to Kim *et al.* (2015), presence of phytochemicals in diets stabilizes the intestinal microbiota and reduces microbial toxic metabolites in the gut preventing intestinal challenge and immune stress.

Hematological parameters of weaner grasscutters fed PATML is presented in Table 6. PCV values ranged between (36.71 – 44.10 %), Hb (9.88 – 14.53 g/dl), RBC 4.94 – 6.72 ($10^6/\mu\text{l}$), MCV (71.22 – 93.41 fl), MCH (24.51-27.83 pg), MCHC (28.34– 31.60 %), WBC 2.41 – 3.04 ($10^6/\mu\text{l}$), lymphocytes (29.45 -34.04 %), monocytes (0.06 – 1.82 %), neutrophils (41.23 – 56.02 %) and neutrophils and eosinophils (0.74 – 1.87 %) respectively. The treatments were significantly ($P<0.05$) influenced by the dietary inclusion of PATML, the values follow similar pattern as they increased from treatment 1 to 5. However, all the results are within the physiologic range established for healthy grasscutters by Opara *et al.* (2006); Ogunsanmi *et al.* (2002); Byanet *et al.* (2008). Similar observation was made by Gboshe *et al.* (2020) who reported a PCV, Hb, RBC and WBC range of (41.70 – 41.90 %), (12.07 – 12.55), 4.95 – 5.35 ($10^6/\mu\text{l}$) and 2.50 – 2.55 ($10^6/\mu\text{l}$) respectively. The normal range in the values could be an indication that the test material (PATML) is safe and that the animals were well nourished (Alagbe, 2018; Gboshe *et al.*, 2020).

According to Ovuru and Ekweozor (2004) Haematological studies are of ecological and physiological interest in helping to understand the relationship of blood characteristics to the environment. Fluctuations in haematological parameters could be attributed to age and sex (Azeez *et al.*, 2009), nutritional deficiencies and feed replacement (Adeyemo *et al.*, 2010), breed (Elagib and Ahmed, 2011; Oloyede *et al.*, 2010). The increase in the RBC level from treatment 1 to 5 is an indication of enough oxygen in the blood, which also has the ability to carry nutrients round the body and keep the animal

healthy. PCV, Hb and RBC are indices used to ascertain anaemia (Isaac *et al.*, 2013). Reductions in red and white blood cell indicate haemolytic anaemia and expose the animals to high risk of infection (Akporhualo, 2011).

White blood cells play a vital role in the prevention of disease or infection, thus animals with low WBC level stand a risk of disease infections. Thus, animals in T₂, T₃, T₄ and T₅ with high WBC are capable of generating antibodies and have a high degree of resistance to diseases (Isaac *et al.*, 2013). Leucocyte counts have also been reported to increase during stress, breed, age and unfavorable conditions (Gotoh *et al.*, 2001). Butterworth (1999) described basophils and eosinophils as important effector cells in allergy and host defense responses particularly against parasitic infections.

The serum biochemical indices of the experimental animals are presented in Table 7. Whereas total protein (4.85 – 6.32 g/dl), globulin (3.11 – 4.02 g/dl), albumin (1.74 – 2.30 g/dl), glucose (63.10 – 89.34 mg/dl), creatinine (0.09 – 1.82 mg/dl), sodium (109.4 – 135.1 mmol/l), phosphorus (2.01 – 3.18 mmol/l) and chlorine (89.81 – 98.09 mmol/l) were lowest ($P<0.05$) for T1. Cholesterol (88.45 – 109.2 mg/dl) were highest ($P<0.05$) for T1 relative to other treatments. ALP (56.30 – 98.09 u/l), AST (41.29 – 63.91 u/l) and ALT (28.12 – 40.34 u/l) were lowest ($P<0.05$) for T5 relative to other treatments.

Serum biochemical analysis is basically used to ascertain the chemical constituents in the blood (Hrubec *et al.*, 2002). The serum total protein, albumin and globulin increases with increasing inclusion of PATLM ($P<0.05$). According to Alikwe *et al.* (2010), serum protein may be used as an indirect measurement of dietary protein quality. Globulin play a significant role in fighting infections, hormone carrier as well as blood clotting process because of the presence of antibodies and enzymes in them (Vivian *et al.*, 2015). However, all within the recommended range according to Opara *et al.* (2006). Cholesterol level decrease with increase in PATML, this clearly removes the risk of cardiovascular disease and ensures that the meat of the animal is safe for consumption. This result is in agreement with the reports of Obikaonu *et al.* (2011) but contrary to the findings of Shittu *et al.* (2019) when rabbits were fed different levels of Ipomoea batatas leaf meal. According to Jenkins (2008), glucose levels in animal's can be elevated due to environmental stress, nutritional stress and improper handling during blood collection. The serum glucose levels determined in this experiment were higher than in some other studies. For instance Gboshe *et al.* (2020) reported a range of 69.35 – 87.73 (mg/dl). Sodium, creatinine, phosphorus and chloride levels were significantly ($P<0.05$) influenced by the inclusion of PATML, the values obtained in this

study is in consonance with the reports of Byanet *et al.* (2008). The creatinine and sodium level is an indication that the integrity of the kidney is maintained. ALT, AST, ALP levels were depressed as the level of PATML increased, indicating that PATML is non-toxic and thus, the risk of liver damage is prevented. This result is consistent with the report of Iyayi (1994) on the supplemental feeding of high and low cyanide cassava to growing pigs.

Conclusion

It can be concluded that PATML contains it contains several secondary metabolites or bioactive chemicals such as tannins, saponins, alkaloids, flavonoids, oxalate etc. which offers wide range of activities including animal performance and increasing nutrient availability which makes them useful as digestive stimulants, antioxidants as well as treatment and prevention of diseases. The inclusion of PATML at 40% resulted in a significant weight gain and does not have any deleterious effect on the general performance of the animal.

Funding:

This research received no external funding.

Conflicts of Interest:

The authors declare no conflict of interest.

REFERENCES

- [1]. Adeniji, A.A. (2009). Protein and energy requirements of weaner grasscutters. *Animal Nutrition and Feed Technology*, 9(1):73-79.
- [2]. Allan, L and Miller, N.D. (1996). Antioxidant, flavonoids structure, function and clinical usage. *Alternative Medicine and Research*, 1:320-329.
- [3]. Alagbe, J.O. (2020). Performance, haematology and serum biochemical parameters of weaner rabbits fed different levels of fermented lagenaria brevilifora whole fruit extract. *Advances in Research and Reviews*, 1(5):1-12.
- [4]. Alagbe, J.O., Sharma, D.O and Xing, Liu. (2019). Effect of aqueous *Piliostigma thonningii* extracts on the haematological and serum biochemical indices of broiler chicken. *Noble International Journal of Agriculture and Food Technology*, 1(2):62-69.
- [5]. Ajibade, V.A and Fagbohun, E.D.(2010). Physicochemical, Proximate Analysis and Antimicrobial Activities of Methanolic Crude Extract of *Tylophora glauca* (Bullock.) *Journal of Pharmaceutical and Biomedical Sciences*, 1(1):7-12.

- [6]. Byanet, O., Adamu, S., Salami, S.O and Obadiah, H.I. (2008). Haematological and plasma biochemical parameters of the young grasscutter reared in the northern Nigeria. *Journal of Cell and Animal Biology*, 2(10): 177-181.
- [7]. Ogunsanmi, A.O., Ozegbe, P.C., Ogunjobi, O., Taiwo, V.O and Adu, J.O. (2002). Haematological, plasma biochemistry and whole blood minerals of the captive adult African grasscutter. *Tropical Veterinarian*, 20(1):97-100
- [8]. Opara, M.N., Ike, K.A and Okoli, I.C. (2006). Haematology and plasma biochemistry of the wild adult African grasscutter. *Journal of Animal Science*, 2(2): 17-22.
- [9]. Ighodaro, I., Agunbiade, S.O., Omale, J.O and Kuti, O.A. (2012). Evaluation of the chemical nutritional, antimicrobial and antioxidant vitamin profiles of *Piliostigma thonningii* leaves. *Research Journal of medicinal plants* 6: 537-543.
- [10]. Alagbe, J.O. (2016). Nutritional evaluation of sweet orange (*Citrus sinensis*) fruit peel as replacement for maize in the diet of weaner grass cutters. *Scholarly Journal of Agricultural Science*, 6(8): 277-282.
- [11]. Alagbe, J.O and Omokore, E.A. (2019). Effect of replacing soya bean meal with *Indigofera zollingeriana* leaf meal on the performance and carcass characteristics of growing rabbits. *International Journal of Multidisciplinary Research and Development*, 6 (5): 74-77.
- [12]. Asibey, E.O.A. (1974). Wildlife as a source of protein in Africa South of the Sahara. *Biological Conservation* 6:32-39.
- [13]. Omokore, E.O and Alagbe, J.O. (2019). Efficacy of dried *Phyllanthus amarus* leaf meal as an herbal feed additive on the growth performance, haematological and serum biochemistry of growing rabbits. *International Journal of Academic Research and Development*, 4(3):97-104.
- [14]. Adeolu, A.A and Sunday, O.O (2013). Anti-inflammatory and analgesics activities of soft drink extract of *Phyllanthus amarus* in some laboratory animals. *British Biotechnology Journal*, 3: 191-204.
- [15]. Meduna, A. J. (2002). Preliminary observations on cane rat feeding and breeding of the Federal College of Forestry, Ibadan. *Proceeding of the 27th Annual Conference on Nigerian Society of Animal Production (FUTA)*, pp 304-305.
- [16]. Wogar, G.S.I and Agwunobi, L.N. (2011). Performance and energy requirements for gestating grasscutters fed Agro-industrial by products. *Journal of Agricultural Sciences*, 4(3): 275-280.
- [17]. Alagbe, J.O and Oluwafemi, R. A. (2019). Performance and haematological parameters of broiler chicks given different level of dried lemon grass and garlic extract. *Research in: Agricultural and Veterinary Sciences*, 3(2): 102-111.
- [18]. Akinpelu, D. A and Obuotor, E.M. (2000). Antibacterial activity of *Piliostigma thonningii* stem bark. *Fitoterapia* 71: 442-443.
- [19]. Butterworth, A.E (1999). Cell mediated damage in helminthes. *Advanced Journal of Parasitology* 23:143.
- [20]. Frankič, T., Volje, M., Salobir, I and Rezar, V. (2009). Use of herbs and spices and their extracts in animal nutrition. *Acta Agric. Slov.* 94:95-102.
- [21]. Gotoh, S., Takennako, O., Vatanabe, K., Kawamoto, R and Watanabe, T. (2001). Hematological values and parasitic fauna in free ranging *Macaca hecki* and the *Macaca tonkeana* *hecki* hybrid group of Sulawesi Island. *Indonesia Primates* 6:91-100.
- [22]. Alagbe, J.O. (2013). Performance and blood profile of grasscutters (*Thryonomys swinderianus*) fed wild sunflower leaf meal. *International Journal of Science and Research*, 5(6): 124-128.
- [23]. A.O.A.C.(2000). Association of Official Analytical Chemists. *Official Methods of Analysis 19th Edition* Washington, D.C Pages 69-77.
- [24]. Duncan, D.B. (1955). Multiple range and multiple F-test. *Biometrics* 11(1):1-42.
- [25]. Alagbe, J.O. (2019). Performance and Haemato-Biochemical Parameters of Weaner Rabbits Fed Diets Supplemented with Dried Water Melon Peel (Rind) Meal. *Journal of Dairy and Veterinary Sciences* 8(4):001-007.
- [26]. Tavares, F.L., Sousa, M.C.C., Santaro, M.L., Barbaro, K.C., Rebecchi, I.M.M and Santos-Martins, I.S. (2004). Changes in haematological, hemostatic and biochemical parameters induced experimentally in rabbits by *Loxosceles gaucho* spider venom. *Journal of Human Experimental Toxicology*, 23:477-486.
- [27]. Burkill, H.M. (1994). *The useful of West Tropical Africa*. 2, 2nd Ed. Royal botanic.
- [28]. Chandan, S., Umehsa, S and Balamurugan, V. (2012). Anti leptospiral antioxidant and DNA

- damaging properties of *Eclipta alba* and *Phyllanthus amarus*. *Open Access Scientific Reports*, 1(4):1-8.
- [29]. Evi, P.L and Degbeku, K. (2011). Antidiabetic activity of *Phyllanthus amarus* and *Thonn* on Alloxan induced diabetes in male wister rats. *Journal of Applied Sciences* 11(6): 2968-2973.
- [30]. Isidorov, V.A., Lech, P., Zolciak, A and Szczepaniak, L. (2008). Gas chromatographic mass spectrometric investigation of metabolites from the needles and roots of pine seedlings at early stages of pathogenic fungi *Armillaria ostoyae* attack. *Trees*, 22: 531-542.
- [31]. Manikkoth, S., Dcepa, B., Joy, A.E and Rao, S. (2011). Anticonvulsant activity of *Phyllanthus amarus* in experimental animal models, 4:144-149.
- [32]. Oluwafemi, F and Debiri, F. (2008). Antimicrobial effect of *Phyllanthus amarus* and *Parquetina nigrescens* on *Salmonella typhi*, *African Journal of Biomedical Research*, 11:215-219.
- [33]. Omer, H.A.A., El-Nomeary, Y.A.A., El-kady, R.i., Badr, A.M.M., Ali, F.A.F., Ahmed, S and Ibrahim, S.A.M. (2012). Growth performance of rabbits fed diets containing different levels of energy and mixture of some medicinal plants. *Journal of Agricultural Science*, 4:201-212.
- [34]. Khartoon, S., Rai, V and Rawat, A. (2004). Comparative pharmacognostic studies of 3 *Phyllanthus* spp. *Journal of Ethnopharmacology*, 104:79-86.
- [35]. Rajeshkumar, N.V., Joy, K.L., Kuttan, G., Ramesewak, R.S., Nair, M.G and Kuttan, R. (2002). Antitumor and anticarcinogenic activity of *Phyllanthus amarus* extract. *Journal of Ethnopharmacology*, 81(1):17-22.
- [36]. Prasad, P.D., Kavimani, S., Suba, V., Nudu, T., Sanatorium, T and Nadu, T. (2013). Antimicrobial activity of the root extracts of *Phyllanthus amarus*. *Journal of Ethnopharmacology*, 4(1):1039-1043.
- [37]. Kim, J.E., Lillehoj, H.S., Hong, Y.H., Kim, G.B., Lee, S.H., Lillehoj, E.P and Bravo, D.M. (2015) Dietary Capsicum and *Curcuma longa* oleoresins increase intestinal microbiome and necrotic enteritis in three commercial broiler breeds. *Res Vet Sci* 102:150–158
- [38]. Sofowora, A. (1993). Medicinal plants and traditional medicine. Spectrum Books Ltd, Ibadan, Nigeria, 224-227.
- [39]. Tasheen, M and Mishra, G. (2013). Ethnobotany and diuretic activity of some selected Indian medicinal plants. *The Pharma Innovation*, 2:112.
- [40]. Zhang, Z. (2014). Bufalin attenuates the stage and metastatic potential of hepatocellular carcinoma in nude mice. *Journal of Traditional Medicine* 12: 57-62.
- [41]. Zubair, M.F., Atolani, O., Ibrahim, S.O., Adebisi, O.O., Hamid, A.A and Sowunmi, R.A. (2016). Chemical constituents and antimicrobial properties of *Phyllanthus amarus*. *Bayero Journal of Pure and Applied Sciences*, 10(1): 238-246.
- [42]. Alagbe, J.O. (2018). Effect of different levels of dried *Delonix regia* seed meal on the performance, haematology and serum biochemistry of growing grasscutters. *Agricultural Research and Technology Open Access Journal*, 18(4): 12-18
- [43]. Jimoh, F.O and Oladiji, A.T. (2005). Preliminary studies on *Piliostigma thonningii* seeds: Proximate analysis, mineral composition and phytochemical screening. *African Journal of Biotechnology*, 4(12): 1439-1442.
- [44]. Oyagade, J.O., Awotoye, O.O., Adewumi, T.J and Thorpe, H.T. (1999). Antibacterial activity of some Nigerian medicinal plants screening. *Journal of Biomedical Research* 11(3):193-197.
- [45]. Nakayama, N.G., Lindsey, M.L and Michael, L.H. (1993). Inhibition of the infectivity of influenza virus by tea polyphenoids. *Antiviral Research* 21:289-299.
- [46]. Srinivasan, K. (2005). Spices as influencers of body metabolism: an overview of three decades of research. *Journal of Food Research*, 38:77-86.
- [47]. Liu, Y., Song, M., Che, T.M., Bravo, D., Pettigrew, J.E. (2012) Anti-inflammatory effects of several plant extracts on porcine alveolar macrophages in vitro. *Journal of Animal Science*, 90:2774–2783
- [48]. Onu, M.M., Oluwafemi, F and Debiri, F (2004). Effect of graded levels of dietary penicillin on the growth rate and feed conversion of broiler chicks. *Journal of Agriculture and Social Research*, 4(2): 81-89.
- [49]. Olatunji, A.K., Alagbe, J.O and Hamed, M.A. (2015). Effect of varying levels of *Moringa olifera* leaf meal on performance and blood profile of weaner rabbits. *International Journal of Science and Research*, 5(6): 803-806.
- [50]. Olanipekun, M.K., Adewuyi, D and Adedeji, D.E. (2016). Ethnobotanical importance and

- phytochemical analysis of some selected medicinal plants used in Ado-Ekiti Local Government Area, Ekiti State. *Journal of Herbal Medicine Research*, 1(3): 0007-0016.
- [51]. Obikaonu, H.O., Okoli, I.C., Opara, M.N., Okoro, V.M.O., Ogbuewu, I.P., Etuk, E.B and Udedbie, A.B.I. (2011). Haematological and serum biochemical indices of starter broilers fed neem leaf meal. *Online Journal of Animal and Feed Research*, 1(4):150-154.
- [52]. National Research Council (1994). Nutrient requirement of poultry 9th Rev Edn, Washington D.C. National Academy Press.
- [53]. Alinnor, J and Oze, R.(2011). Chemical Evaluation of the Nutritive value of *Pentaclethra macrophylla* benth (African Oil Bean) Seeds. *Pakistan Journal of Nutrition*, 10 (4):355 – 359.
- [54]. Akpanyung, E.O. (2005). Proximate and mineral composition of bouillon cubes produced in Nigeria. *Pakistan Journal of Nutrition*, 4(5): 327-329.
- [55]. Ibrahim, N.D.G., Abdulrahman, E.M and Ibrahim, G. (2001). Elemental analysis of the leaves of *Vernonia amydalina* and its biological evaluation in rats. *Nigerian Journal of Natural Products and Medicine*, 5:13-17.
- [56]. Ross, I.A. (2005). Medicinal plants of the world : chemical constituents, traditional and modern medicinal uses. Humma Press Inc. New Jersey 1: 15-31.
- [57]. Lillehoj, H.S and Lee, K.W. (2012) Immune modulation of innate immunity as alternatives to antibiotics strategies to mitigate the use of drugs in poultry production. *Journal of Poultry Science*, 91:1286–1291
- [58]. Min, B.R., Hart, S.P., Miller, D., Tomita, G.M., Loetz, E and Sahlu, T. (2005) The effect of grazing forage containing condensed tannins on gastro-intestinal parasite infection and milk composition in Angora does. *Journal of Veterinary Parasitology*, 130:105–113.
- [59]. Wallace, R.J, Arthaud, L., Newbold, C.J. (1994) Influence of *Yucca shidigera* extract on ruminal ammonia concentrations and ruminal microorganisms. *Applied Environmental Microbiology* 60: 1762–1767.
- [60]. Redondo, L.M., Chacana, P.A., Dominguez, J.E and Miyakawa, M.E.F. (2014) Perspectives in the use of tannins as alternative to antimicrobial growth promoter factors in poultry. *Front Microbiology*, 5:118.
- [61]. Hollman, P.C. (2001). Evidence for health benefits of plant phenols: Local or systemic effects. *Journal of Food Science and Agriculture*, 81: 842-852.
- [62]. Faizi, S., Khan, R.A., Mughal, N.R., Malik, M.S., Sajjadi, K.E and Ahmad, A. (2008). Antimicrobial activity of various effect of *Polyalthia longifolia* : isolation of active principles from the leaves and barriers. *Journal of Phytochemistry*, 22: 907-912.
- [63]. Ovuru, S.S and Ekweozor, I. K. E. (2004). Haematological changes associated with crude oil ingestion in experimental rabbits. *African Journal of Biotechnology*, 3(6):346-348.
- [64]. Ojewuyi, O.B., Ajiboye, T.O., Adebajo, E.O., Balogun, A and Mohammed, A.O. (2014). Proximate composition, phytochemical and mineral contents of young and mature *Polyalthia longifolia* Sonn. Leaves. *Fountain Journal of Natural and Applied Sciences*, 3(1):10-19.
- [65]. A.O.A.C.(2000). Association of Official Analytical Chemists. Official Methods of Analysis 19th Edition Washington, D.C Pages 69-77.
- [66]. Duncan, D.B. (1955). Multiple range and multiple F-test. *Biometrics* 11(1):1-42.
- [67]. Iyayi.E.A. (1994). Supplemental effect of low and high cyanide cassava on the performance, nutrient digestibility and serum metabolites of growing pigs. *Journal of Agricultural Tropics and Sub-tropics*. 95:199-205.
- [68]. Isaac, L.J., Abah, G., Akpan, B and Ekaette, I.U. (2013). Hematological properties of different breeds and sexes of rabbits (p.24-27). *Proceedings of the 18th Annual Conference of Animal Science Association of Nigeria*.
- [69]. Shittu, M.D., Adesina, G.O and Eseigbe, S. (2019). Productive performance and blood profile of weaner rabbits fed different inclusion levels of *Ipeomoea asarifolia* leaf meal in replacement of soyabean meal. *Journal of Biotechnology Research*, 5(11):107-112.
- [70]. Adeyemo, G.O., Ologbodo, A.D and Adebisi, O.A. (2010). The effect of graded levels of dietary methionine on the hematology and serum biochemistry of broilers. *International Journal of Poultry Science*, 9(2):158-161.
- [71]. Akporhwarho, P.O. (2011). Effect of crude oil polluted water on the hematology of cockerel reared under intensive system. *International Journal of Poultry Science*, 10(4):287-289.
- [72]. Alikwe,P.C.N., Faremi, A.Y and Egwaikhide, P.A. (2010). Biochemical evaluation of serum metabolites enzymes and hematological indices of broiler chicks fed with varying

- levels of rumen epithelial scraps in place of fish meal proteins. *Research Journal of Poultry Science*,6(12):855-857.
- [73]. Azeez, O.I., Ayagbemi, A.A and Oyewale, J.O.(2009). Diurnal fluctuation in haematological parameters of the domestic fowl in the hot humid tropics. *International Journal of Poultry Science*, 8(3):247-251.
- [74]. Elagib, H.A.A and Ahmed, A.D.A. (2011). Comparative study on haematological values of blood of indigenous chickens in Sudan. *Asian Journal of Poultry Science*, 5:41-45.
- [75]. Oloyede, O.B., Minari, J.B and Mohammad, N.O. (2010). Evaluation of growth characteristics and haematological indices of broiler chicks fed raw and processed Bambara nut seed as a component of poultry feed, 9(7):652-655.

6/2/2021