Using Carboxylic Enriched With Carboxylic Calcium, Algae Extract, and Glycine to Promote Yield and Fruit Quality of Williams Banana Plants

Hamdy I. M. Ibrahim¹, Ahmed Y. Mohamed² and Hassan E. M. Ibrahim¹

¹Hort. Dept. Fac. of Agric. Minia Univ., Egypt. ²Tropical Fruits Res. Dept. Hort. Res. Instit. ARC. Giza faissalfadel@yahoo.com

Abstract: During 2016/2017 and 2017/2018 seasons, Williams banana plants were treated with carboxylic -Ca and glycine each at 0.1 % as well as Algae extract at 0.05 to 0.2% either singly or in all possible combinations. Plants received three sprays at the first week of May and at one month intervals. The objective was examining the impact of these materials on vegetative growth aspect, plant nutritional status, bunch weight and fruit characteristic of Williams banana plants. Subjecting Williams banana plants thrice with carboxylic -Ca and glycine each at 0.1% and Algae extracts at 0.05 to 0.2% singly or in combinations had considerable promotive effect on all growth aspects, N, P, K, Mg, bunch weight as well as all quality parameters. Using glycine was materially superior than using carboxylic -Ca in this respects. Using all materials together gave the best results. For promoting yield and fruit quality of Williams banana plants grown under Minia region conditions, it is suggested to use a mixture of Algae extract, carboxylic -Ca and glycine each at 01% thrice at are the first week of May and at one month intervals. [Hamdy I. M. Ibrahim, Ahmed Y. Mohamed and Hassan E. M. Ibrahim. Using Carboxylic Enriched With Carboxylic Calcium, Algae Extract, and Glycine to Promote Yield and Fruit Quality of Williams Banana **Plants.** N Y Sci J 2019;12(1):17-29]. ISSN 1554-0200 (print); ISSN 2375-723X (online).

http://www.sciencepub.net/newyork. 3. doi:10.7537/marsnys120119.03.

Keywords: Algae extract, carboxylic- Ca, glycine, Williams banana, growth, plant nutritional status, bunch weight, fruit quality

1. Introduction

Many attempts were accomplised for promoting yield and fruit quality of Williams banana by using natural substances namely Algae extracts, carboxylic -Ca and glycine for avoiding our environment from pollution.

Amino acids as organic nitrogenous compounds are the building blocks in the synthesis of proteins, which are formed by as process in which ribosomes catalyze the polymerization of amino acids (**Davies**, **1982**). Several hypothesis have been proposed the explain for the role of amino acids in plant. Available evidence suggests several alternative routes of IAA and ethylene synthesis in plants, starting from amino acids. In this respect, (**Taiz and Zeiger 2002**) suggested that the regulatory effect of certain amino acids like phenylalanine and ornithine in plant development appeared through their influence on the biosynthesis of gibberellins, tryptophan and methionine in building of I.A.A and ethylene respectively.

Calcium has been known as an essential element for trees for a long time and it has received much attention from the fertility standpoint and also very important for facilitating marketing of the fruits. Calcium is required for cell division and chromosome stability. As Ca-pectate, it is a constituent of the middle lamella of cell walls (Dodd *et el*, 2010). Calcium indirectly influences many

enzyme systems such as amylase and ATPase. It also seems to have a regulating role in respiration and prolonging shelf life of fruits (Marschner, 2010).

One approach to increase crop yield is the developing of non polluting organic biostimulants. The application of Algae extract which contains most nutrients, organic compound, enzyme, vitamins, antioxidants, amino acids and natural hormones is fast becoming an accepted practice. It increases yield quantitatively and qualitatively in various fruit crops (Soliman *et al.*, 2008 and Spinelli *et al.*, 2010).

Using biofertilizer extracts nowadays for fruit crops has called the attention of research workers as an alternative to synthetic auxins and mineral N fertilization. They are very safe for human, animal and environment. Clean cultivation is greatly achieved by using biofertilizers or their exudates (Kullk, 1995; Wani and Lee, 1995; Strick *et al*, 1997; Adam, 1999; Saffan-Samia, 2001; Tung-Yunn *et al*, 2003; and Ordog *et al.*, 2004).

Previous studies showed that Algae extract (Ebeid- Sanaa, 2007; El-Sawy,2008b, Gamal,2013, Mohamed, and El-Sehrawy,2013, Oraby,2013; Ahmed *et el* 2013; Ahmed *et el*, 2014; Eshmawy, 2015, Abd El-aaty,2015 and Ahmed *et el* 2015; Ca sources (joutamance *et el*, 2002, El-Shafey *et el* 2002, El-Tanany *et el*, 2004, Young Ho *et el*, 2004, Sarawy *et el*, 2012; Chakerolhasseini *et el*, 2016; Habasy- Randa *et el*,2016; Mohamrd *et el*,2017; Meena *et el*, 2017 and Hikal *et el*,2017 and amino acids (Ahmed *et el* 2014a; Ahmed *et el* 2014b, El-Khawaga, 2014, Hassan-Huda,2014, Sayed-Ola, 2014b and Mohamed,2016) were very effective in improving, growth, plant nutritional, status, yield and fruit quality of different fruit crops.

The target of this study was examining the effect of spraying Algae extract, Carboxylic -Ca and glycine on fruiting of Williams banana plants.

2. Material and Methods

This study was carried out during two successive seasons of 2016/ 2017 and 2017/ 2018 on third and fourth ratoons of Williams banana plants grown in a private fruit orchard situated at El-Gendia village at eastern bank of Bni Mazar city, Minia Governorate where the soil is silty clay and well drained and with a water table not less than two meters deep. The stools ate planted at 3.5 x 3.5 m apart. Surface irrigation system was followed.

The target of this study was examining the effect of spraying Algae extract, Carboxylic -Ca and glycine on fruiting of Williams banana plants.

Analysis of the tested soil at 0.0 to 90.0 cm depth was carried out according to the procedures that outlined by **Black (1965)** and the obtained data are shown in Table (1).

Table (1): Analysis of the teste	d soil
----------------------------------	--------

Constituents	Values
Particle size distributions	
Sand %	4.7
Silt %	60.0
Clay %	35.3
Texture	Silty clay
pH (1: 2.5 extract)	7.92
E.C. (1: 2.5 extract) (mmhos/1cm/25 $^{\circ}$ C)	1.72
O.M. %	1.42
CaCO ₃ %	2.22
Total N %	0.09
Available P (ppm, Olsen)	5.2
Available K (ppm, ammonium acetate)	402.2

Horticultural practices such as fertilization with 25 tons farmyard manure ($0.25 \ \%N$, $1.0\% \ K2O$ and $0.4\% \ P2O5$); 2.25 tons ammonium nitrate ($33.5 \ \%N$), one ton triple calcium superphosphate ($37.5 \ \%P_2O_5$) and 1.5 tons potassium sulphate ($48 \ \% \ K_2O$) per fed., irrigation with Nile water, hoeing as well as training, pest and fungi control were carried out as usual. Farmyard manure was added once at the mid of Jan. Ammonium nitrate was splitted into equal batches and added as two batches for each month started from fourteen April to October for each

season. Potassium sulphate was divided into two equal batches applied before the emergence of the inflorescences and after fruit setting. Triple calcium superhosphate was added twice, the first with farmyard manure and the second before the emergence of the inflorescence.

This study included the following sixteen treatments from two factors (A & B). The first factor (A) comprised from the following four carboxylic – Ca and glycine treatments:

a₁) Non material

a2) Carboxylic –Ca at 0.1%

a₃) Glycine at 0.1%

a₄) Both at 0.1%

The second factor (B) contained the following four Algae extract concentrations

 $\begin{array}{c} b_1) \ 0.0 \ \% \left(\ 0.0 \ g/ \ L^{-1} \right) \\ b_2) \ 0.05 \ \% \left(\ 0.5 \ g/ \ L^{-1} \right) \\ b_3 \ 0.1 \ \% \left(\ 1 \ g/ \ L^{-1} \right) \end{array}$

 b_4) 0.2 % (2 g/L⁻¹)

Each treatment was replicated three times, one stool per each. Each stool contained three plants for fruiting in the current season plus three suckers for fruiting in the following season. Carboxylic -Ca, glycine and Algae extract were sprayed three times at the first week of May and at one month intervals. Triton B as a wetting agent was added to all solutions of Carboxylic -Ca, glycine and Algae extract. Chemical analysis of Algae extract are shown in table (2) (according to **James, 1994)**.

Table (2): Analysis of Algae extract

Constituents	Values
Moisture %	6.0
O.M. %	45.6
Inorganic matter %	45.6
Proteins %	6.8
Carbohydrates %	5.5
Aliginic acid %	10.2
Mannitol %	4-7
Total N %	1.0 - 1.0
Total P%	0.02-0.09
Total K %	1.0-1.2
Total Ca %	0.2-1.5
Total S %	3-9
Total Mg %	0.5-0.9
Cu (ppm)	1.0-6.0
Fe (ppm)	50-200
Mn (ppm)	5-12
Zn (ppm)	10-100
B (ppm)	20-100
Mo (ppm)	1-5
Cytokinins %	0.02
IAA %	0.03
ABA %	0.03

This experiment was set up in a randomized complete Block design (RCBD) in split plot arrangement which the four Carboxylic -Ca concentrations and the four Algae extract concentrations occupied the main and sub- plots, respectively.

After the emergence of the inflorescence at the beginning of July for both the third and fourth ratoons, the following characteristic were determined.

1- Pseudostem height in (cm) was measured from the soil surface up to the petiole of the last emerged leaf.

2- Girth of pseudostem (cm) in the base, middle and top of pseudostem was recorded then the average was calculated.

3- Number of green leaves per plant was recorded.

4- Leaf area was recorded according to the following equation:-

 $LA = 0.67 (L \times W) + 107.15$ (Ahmed and Morsy, 1999) which L= leaf length (cm) and W = leaf width (cm)

5- Assimilation area $(m)^2$ was calculated by multiplying average leaf area by number of green leaves/ plant.

Leaf samples were taken from the third upper leaf in the descending leaves from the top of the plant after bunch shooting in September in the two seasons. A simple of 10x 10(cm) area from the middle part of the leaf blades as recommended by Summer (1985) was taken for mineral analysis. Leaf samples were washed several times with tap water then with distilled water after they were dipped in 0.1 NHCl Samples were oven dried at 60- 70 °C until a constant weight and then ground by using an electric mill. Weight of 0.5 gram of the ground material was digested using a mixture of perchloric acid: sulphoric acid (1: 10) (v/v) according to the (Chapman and Pratt (1965). The clear digestion was quantitavely transferred to 100 ml volumetric flask. In this solution, the following nutrients were determined:

a) Total nitrogen was determined by using the micro- Kjeldahl method as described by (Cottenie *et al.*, 1982).

b) Phosphorus was determined by using the method of Olsen as report by (Chapman and Pratt 1965).

c) Potassium was determined by using Flame photometric, according to the method of (wild *et el* (1985).

d) Magnesium was determined by using versene method (Cottenie *et al.*, 1982).

The bunches of both the third and fourth ratoons were picked at the middle of November in

2016/ 2017 and 2017/ 2018 seasons when the fingers reached three quarter stage. Bunch weight in kg (before artificial ripening),and average hand weight (g.) were recoded. Three hands were taken from the base middle and distal end of the bunch as a composite sample for the physical and chemical determination. Then, the chosen hands were wrapped with newspaper and arrested in closed wooden boxes with a glass surface to achieve artificial ripening and after the fingers ripened, the following physical determinations were carried out:

1- The weight of the finger (g) by using an analytical balance.

2- Fingers dimensions (diameter and length).

3- The weight of pulp and peel (g) by weighing the pulp and peel and expressed as a percentage from the finger weight.

4- The ratio between pulp and peel.

A composition fresh sample was taken from the pulp and mixed using an electric blender, then the following chemical constituents were determined:

1-Percentage of total soluble solids by using a handy refractometer.

2- Percentage of reducing, non reducing and total sugars as well as starch by using methods of (Lane and Eynon 1965) that outlined in (A.O.A.C. 2000).

3- Percentage of total acidity (expressed as g malic acid per 100 g pulp) by using titration against 0.1 N sodium hydroxide and using phenolephthalein as an indicator (A.O.A.C., 2000).

The obtained data were analyzed and the differences between different treatment means were compared using new L.S.D. test according to (Mead *et al.*, 1993).

3. Results and Discussion

1-Effect of single and combine applications of carboxylic-Ca, glycine and Algae extract on some vegetative growth aspects:

Data in Tables (3 & 4) show the effect of single and combine applications of Ca, glycine and seaweed extract on pesudostem height and girth, number of green leaves /plant and leaf area of Williams banana plants during 2016/2017 and 2017/2018 seasons.

It is clear from the obtained data that subjecting the plants to calcium Ca and /or glycine each at 0.1% had signified promotion on the aforementioned growth traits rather the control. Using the amino acid namely glycine at 0.1% significantly surpassed. The application of Ca at 0.1% in enhancing the investigated characteristics. Using Ca in combined with glycine each at 0.1% significantly enhanced there growth attributes compared with using each singly. The maximum values were recorded due to using both materials together. Similar trend was noticed during both seasons.

It is clear from the obtained data that treating Williams banana plants with Algae extract at 0.05% to 0.2% significantly was accompanied with Algae extract at 0.05% to 0.2% stimulating the four growth aspects namely of leaves /plant and leaf area relative to the control (0.1% Algae extract). The promotion on three growth aspects was related to increase in Algae extract concentrations from 0.0 to 0.2 % significant differences on their growth aspects were recorded among all concentrations of Algae extract except among of the height two concentrations namely 0.1 and 0.2%.

The height values were recorded on the plants that received three sprays of Algae extract at 0.2%.

The lowest values were recorded on untreated plants. These resulted were true during both seasons.

Data in Tables (5 & 6) show that the interaction between Algae extract, Ca and glycine had significant effects on all vegetative growth aspects. The maximum values were recorded on the plants that received three sprays of a mixture of Algae extract at 0.2% plus Ca and glycine each at 01%.

Under each treatment the leaf area reached 1.41 and 1.46 m² during both seasons, respectively. Control treatments gave the lowest values of leaf area ($0.85 \& 0.88 m^2$) during both seasons, respectively the same trend was noticed during 2016/2017 and 2017/2018 seasons.

2-Effect of single and combine applications of carboxylic-Ca, glycine and Algae extract on N, P, K and Mg

Tables (5 & 6) show the effect of single and combine applications of Ca, glycine and Algae extract on the percentage of N, P, K and Mg in the leaves of Williams banana plants.

Concerting the effect of using Ca and/or glycine. The obtained data show that single and combined application of Ca- nitrate and glycine each at 0.1% significantly enhancing the percentage of N, P, K and Mg in the leaves relative to the control. Using glycine at 0.1 was significantly superior than using Ca at the same concentrations in enhancing these nutrients. Using both materials together significantly surpassed the application of each in this respect. Treating the plants with Ca and glycine each at 0.1% gave the maximum values. The untreated plants received the lowest values. Similar trend was noticed during both seasons.

Subjecting Williams banana plants to Algae extract at 0.05 to 0.2 % significantly was followed by enhancing the percentage of N, P, K and Mg in the leaves of Williams banana plants over the use of Algae extract at 0.0% there was gradual promotion on these nutrients with increasing concentrations of

Algae extract from 0.0 to 0.2 %. Increasing concentrations of Algae extract from 0.1 to 0.2 % had no significant stimulations on these nutrients. The maximum values were recorded on the plants that received three sprays of Algae extract at 0.2%. The untreated plants produced the minimum values. These results were true during seasons.

All nutrients in the leaves (N, P, K, Mg) were significantly enhancing in response to using all materials (Algae extract, Ca and glycine) at the higher concentrations namely 0.2%,0.1 and 0.1 % respectively relative to the control The untreated plants gave the lowest values. These results were true during both seasons.

3-Effect of single and combine applications of carboxylic Ca, glycine and Algae extract on weights of bunch and hand;-

Data in Table (7) show the effect of single and combine applications of Ca, glycine and Algae extract on weights of bunch and finger of Williams banana plants.

Treating Williams banana plants with Ca and/or glycine each at 0.1 significantly improved weights of bunch and hand over the control. Using glycine was significantly favourable than using Ca in this respect. Using Ca in combined with glycine significantly improved weights of bunch and hand relative to the use of any material alone. The heaviest bunches and hands were receded on the plants that received Ca and glycine each at 0.1% together. The untreated plants received the highest bunch and hands. Similar trend was noticed during both seasons.

Spraying Algae extract at 0.05 to 0.2% significantly improved weights of bunches and hands over the check treatment. The promotion was in proportional to the increase in Algae extract concentrations. Meaningless promotion on weight of bunch and hand was observed with increasing concentrations of Algae extract concentrations from 0/1 to 0.2 %. Therefore from economical point of view it is suggested to use Algae extract at 0.1 %.

The untreated plants gave the lowest values. These results were true during both seasons.

The interactions all factors had significant promoitive effect on the weight of bunch and hand. From economical point of view it is suggested to use Algae extract besides Ca and glycine each at 0.1%. three times to produce the highest values of brunch and hand were 29.6 and 2.15 kg in the first season and 30.6 and 2.09 kg in the second season, respectively. The untreated plants produced 21.0 and 1.29 kg during 2016/2017 season and 20.0 and 1.31 kg during 2017/2018 season, respectively. The percentage of increment on the average bunch weight and hand weight over the control.

In the first season reached 42.4 and 66.7% and in the second one reached 53.0 and 54.5% respectively. These results were true during both.

4-Effect of single and combine applications of carboxylic-Ca glycine and Algae extract on some physical and chemical characteristics of the fruit of Williams banana plants:-

Data in tables (98to13) Show the effect of single and combine applications of Ca, glycine and Algae extract on some physical and chemical characteristics of the finger weight, length and width, fruit pulp and peel %, pulp/peel, starch %,T.S.S %, reducing, total and non-reducing sugars % and titrilable acidity of the fruit of Williams banana plants.

Treating Williams banana plants with Ca and /or glycine each at 0.1 significalty was very effective in improving fruit quality in terms of increasing fingers weight and dimensions (length & width), fruit pulp%, T.S.S%, total, reducing and nonreducing sugars and decreasing fruit peel weight%; starch % and titratible acidity % relative to the control.

Using glycine was significantly favourable than using Ca in enhancing fruit quality. Combined applications of Ca and glycine each at 0.15 gave the best results with regard. To fruit quality than using each alone. Unfavourable effect on fruit quality was

attributed to unsparing these materials. Similar trend was noticed during 2016/2017 & 2017/2018 seasons.

It is evident from the obtained data that fruit was significantly improved in response to treating the tress three times with Algae extract at 0.05 to 0.2% over the control. This promotion was significantly appeared in terms of increasing weight, length and width of fruit pulp %, T.S.S %, total, reducing and non- reducing sugars and decreasing fruit peel weight%; starch % and titratible acidity.

The promotion on fruit quality was associated concentration of Algae extract. increasing Significantly different parameters were observed among all concentrations except among all height two concentrations namely 0.1% and 0.2%. The untreated produced undesirable effects on fruit quality. Similar results were announced during 2016/2017 and 2017/2018 seasons.

It is obvious from the obtained data that fruit quality parameters was significantly varied among all the combinations of the investing treatment. The best results with regarded to fruit quality were obtained due to treating the plants three times with a mixture of Algae extract at 0.2 % as well as glycine and Ca due at 0.1%. The plants treated with Algae extract, Ca and glycine each at 0.0%. These results were true during both seasons.

Table (3):- Effect of single and combined applications of Ca, glycine and Algae extract extracts on The number of green leaves /plant and leaf area (m ²) of Williams banana plants during 2016/2017 and 2017/2018
seasons.
The number of green leaves /plant

The number of green	1 leave	es /plant											
	Ca a	nd glycine trea	tments (A	N)									
	2016	/2017				2017/2018							
Algae extract	a_1	a ₂	a 3	a ₄		a 1	a ₂	a 3	a ₄	_			
Treatments (B)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)			
b _{1Algae} at 0.0%	12.0	14.0	16.0	18.0	15.0	13.0	15.0	17.0	20.0	16.3			
b ₂ Algaeat 0.05%	13.0	16.0	18.0	20.0	16.8	15.0	18.0	21.0	23.0	19.3			
b ₃ Algaeat 0.1%	16.0	18.0	20.0	24.0	19.5	17.0	20.0	24.0	25.0	21.5			
b ₄ Algaeat 0.2%	16.0	18.0	20.0	24.0	19.5	17.0	20.0	24.0	25.0	21.5			
Mean (A)	14.3	16.5	18.5	21.5		15.5	18.3	21.5	23.3				
		Α	B	AB			Α	В	AB				
New L.S.D at 5 %		1.0	1.0	2.0			1.0	1.0	2.0				
	leaf a	urea (m ²)											
	Ca a	Ca and glycine treatments (A)											
	2016	/2017				2017/2018							
Algae extract	\mathbf{a}_1	a ₂	a ₃	a ₄		a_1	a ₂	a ₃	a ₄				
Treatment (B)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)			
b _{1Algae} at 0.0%	0.85	0.96	1.06	1.18	1.01	0.88	0.99	1.10	1.23	1.05			
b _{2Algae} at 0.05%	0.98	1.10	1.20	1.31	1.15	1.01	1.14	1.25	1.35	1.19			
b ₃ Algae at 0.1%	1.08	1.21	1.33	1.40	1.26	1.11	1.25	1.40	1.45	1.30			
b ₄ Algae at 0.2%	76.1	1.09	1.22	1.34	1.41	1.12	1.26	1.40	1.46	1.31			
Mean (A)	1.00	1.12	1.23	1.33		1.03	1.16	1.29	1.37				
		Α	В	AB			Α	В	AB				
New L.S.D at 5 %		1.0	0.04	0.08			0.07	0.06	0.12				

pseudo stem height (o	cm)													
	Ca a	Ca and glycine treatments (A)												
Algae extract	2016	/2017			2017	/2018								
Treatments (B)	a_1	a ₂	a 3	84		a_1	a ₂	a 3	a 4	ĺ				
	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)				
b1 Algaeat 0.0%	1.58	1.79	1.99	2.20	1.89	1.69	1.84	2.05	2.25	1.95				
b Algaeat 0.05%	1.76	1.99	2.22	2.51	2.13	1.85	2.04	2.25	2.45	2.15				
b ₃ Algaeat 0.1%	2.00	2.21	2.50	2.72	2.36	2.07	2.25	2.45	2.65	2.36				
b ₄ Algaeat 0.2%	2.05	2.22	2.51	2.74	2.38	2.10	2.26	2.46	2.66	2.37				
Mean (A)	1.83	2.03	2.31	2.54		1.42	2.09	2.30	2.50					
		Α	В	AB			Α	В	AB					
New L.S.D at 5 %		0.16	0.14	0.28			0.18	0.15	0.30					
	Pseu	do stem girth (e	cm)											
		nd glycine trea /2017	tments (A)	2017/2018									
Algae extract	a_1	82	a ₃	a ₄		\mathbf{a}_1	a2	a ₃	a ₄					
Treatments (B)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)				
b ₁ Algae at 0.0%	73.9	75.0	76.1	77.4	75.6	74.0	75.9	77.3	79.0	76.6				
b ₂ Algae at 0.05%	74.9	76.1	77.5	79.0	76.9	76.1	77.9	79.9	82.0	79.0				
b ₃ Algae at 0.1%	76.0	78.0	79.1	81.0	78.5	77.4	79.4	81.4	84.0	80.6				
b ₄ Algae at 0.2%	76.1	78.3	79.3	81.9	78.9	77.5	79.5	81.7	84.5	81.0				
Mean (A)	75.2	76.9	78.0	79.8		76.3	78.2	80.1	82.4					
		Α	В	AB			Α	В	AB					
New L.S.D at 5 %		1.0	0.8	1.6			1.1	0.9	1.8					

Table (4):- Effect of single and combined applications of Ca, glycine and Algae extract extracts on pseudo stem height (cm) and Pseudo stem girth (cm) of Williams banana plants during 2016/2017 and 2017/2018 seasons

Table (5):- Effect of single and combined applications of Ca, glycine and Algae extract extracts on The percentage of N and P in The leaves of Williams banana plants during 2016/2017 and 2017/2018 seasons Leaf N %

Leaf N %														
	Ca a	Ca and glycine treatments (A)												
Algae extract	2016	/2017			2017/2018									
Algae extract Treatments	\mathbf{a}_1	a ₂	a ₃	a 4		\mathbf{a}_1	a ₂	a ₃	a 4					
(B)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)				
b ₁ Algae at 0.0%	2.41	2.53	2.66	2.80	2.60	2.40	2.55	2.68	2.82	2.61				
b ₂ Algae at 0.05%	2.55	2.71	2.84	2.99	2.77	2.58	2.74	2.87	3.01	2.80				
b ₃ Algae at 0.1%	2.71	2.86	2.99	3.14	2.93	2.74	2.89	3.01	3.18	2,96				
b ₄ Algae at 0. 2%	2.72	2.87	2.99	3.15	2.93	2.75	2.90	3.02	3.19	2.97				
Mean (A)	2.60	2.74	2.87	3.02		2.62	2.77	2.90	3.05					
		Α	B	AB			Α	В	AB					
New L.S.D at 5 %		0.07	0.06	0.12			0.06	0.07	0.14					
	Leaf	Р %												
	Ca a	Ca and glycine treatments (A)												
A1	2016	/2017			2017/2018									
Algae extract	a 1	a ₂	a 3	a 4		a_1	a ₂	a 3	a 4					
Treatments (B)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)				
b ₁ Algaeat 0.0%	0.19	0.28	0.36	0.44	0.32	0.17	0.29	0.37	0.43	0.32				
b ₂ Algae at 0.05%	0.28	0.36	0.45	0.56	0.41	0.29	0.37	0.46	0.57	0.42				
b ₃ Algaeat 0.1%	0.36	0.44	0.50	0.64	0.44	0.36	0.43	0.50	0.64	0.48				
b Algae at 0.2%	0.37	0.45	0.50	0.64	0.49	0.38	0.44	0.51	0.65	0.50				
Mean (A)	0.30	0.38	0.45	0.57		0.30	0.38	0.46	0.57					
		Α	B	AB			Α	В	AB					
New L.S.D at 5 %		0.05	0.04	0.08			0.03	0.03	0.06					

Bunch weight (kg) /	plant												
	Ca a	nd glycine trea	tments (A										
Algae extract	2016	/2017			2017	/2018							
Algae extract Treatments	\mathbf{a}_1	a ₂	a ₃	a ₄		\mathbf{a}_1	a ₂	a ₃	a ₄				
(B)	Non	0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine				
b _{1Algae} at 0.0%	21.0	22.5	24.0	23.5	23.3	20.0	23.0	24.6	26.0	23.4			
b _{2Algae} at 0.05%	22.6	24.0	25.9	28.0	25.1	23.1	24.6	26.5	28.7	25.7			
b ₃ Algae at 0.1%	24.0	25.7	27.1	29.6	26.6	24.6	26.2	27.9	30.6	27.3			
b _{Algae} at 0.2%	24.2	26.0	27.5	29.9	26.9	24.7	26.4	28.0	30.7	28.2			
Mean (A)	23.0	24.6	26.1	28.3		23.1	25.1	26.8	29.0				
		Α	В	AB			Α	В	AB				
New L.S.D at 5 %		0.6	0.7	1.4			0.6	0.6	1.2				
	Hano	l weight (kg)											
	Ca a	Ca and glycine treatments (A)											
Algae extract	2016	/2017			2017/2018								
Treatments	a_1	a ₂	a 3	84		a ₁	a ₂	a 3	a 4				
(B)	Non	0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)			
b _{1Algae} at 0.0%	1.29	1.42	1.55	1.71	1.49	1.31	1.46	1.59	1.75	1.53			
b Algae at 0.05%	1.42	1.55	1.71	1.90	1.65	1,47	1.60	1.75	1.95	1.69			
b _{3Algae} at 0.1%	1.55	1.72	1.86	2.15	1.82	1.60	1.76	1.90	2.09	1.84			
b ₄ Algae at 0.2%	1.56	1.73	1.90	2.16	1.84	1.61	1.77	1.91	2.11	1.85			
Mean (A)	1.46	1.61	1.76	1.98		1.50	1.65	1.79	1.98				
		Α	В	AB			Α	В	AB				
New L.S.D at 5 %		0.07	0.06	0.12			0.05	0.05	0.10				

Table (6):- Effect of single and combined applications of Ca, glycine and Algae extract extracts onbunch weight (kg) / plant and hand weight (kg) of Williams banana plants during 2016/2017 and 2017/2018 seasons

Table (7):- Effect of single and combined applications of Ca, glycine and Algae extract on Fruit weight (g) and fruit length (cm) of Williams banana plants during 2016/2017 and 2017/2018 seasons

Fruit weight (g)													
	Ca an	d glycine treat	ments (A)										
Algae extract	2016/2	2017				2017/2018							
Treatments	a 1	a ₂	a ₃	a ₄		a 1	a ₂	a 3	a ₄				
(B)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine				
b ₁ Algaeat 0.0%	88.0	90.0	92.1	95.0	91.3	90.0	91.2	93.2	96.2	92.7			
b Algae at 0.05%	90.5	92.9	95.0	97.5	94.0	91.5	94.0	96.2	99.0	95.2			
b _{3Algae} at 0.1%	93.0	95.1	98.0	100.0	96.5	94.1	96.2	99.2	99.3	97.2			
b4Algaeat 0.2%	93.3	95.2	98.3	100.6	96.9	94.1	96.3	99.3	99.5	97.3			
Mean (A)	91.2	93.3	95.9	98.3		92.4	94.4	97.0	98.5				
		Α	В	AB			Α	В	AB				
New L.S.D at 5 %		1.1	0.9	1.8			1.0	1.0	2.0				
	Fruit	length (cm)											
	Ca an	Ca and glycine treatments (A)											
Algae extract	2016/2	2017			2017/2018								
Treatments	\mathbf{a}_1	a ₂	a 3	a 4		\mathbf{a}_1	a ₂	a ₃	a 4				
(B)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)			
b ₁ Algae at 0.0%	14.9	15.01	15.13	12.26	15.08	15.00	15.12	15.31	15.44	15.22			
B2 Algae at 0.0%	15.30	15.70	16.00	16.12	15.61	15.60	15.75	16.10	16.25	15.93			
B3 Algae at 0.0%	15.60	16.00	16.13	16.26	16.00	15.80	16.05	16.22	16.33	16.10			
b _{4Algae} at 0.0%	15.62	16.03	16.14	16.27	16,02	15.81	16.06	16.24	16.34	16.11			
Mean (A)	15.36	15.69	15,85	15.98		15.55	15.75	15.97	16.09				
		Α	В	AB			Α	В	AB				
New L.S.D at 5 %		0.06	0.05	0.10			0.06	0.04	0.08				

Finger width (cm)													
	Ca an	d glycine treat	ments (A)										
Algon ovtwort	2016/2	2017				2017/2018							
Algae extract Treatments	a ₁	a ₂	a 3	a ₄		\mathbf{a}_1	a ₂	a 3	a ₄				
(B)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)			
b _{1Algae} at 0.0%	7.81	7.91	8.2	8.12	7.97	7.76	7.90	8.05	8.20	7.98			
b _{2s} Algae at 0.05%	7.92	8.04	8.24	8.50	8.18	7.91	8.09	8.30	8.50	8.20			
b ₃ Algaeat 0.1%	8.03	8.26	8.70	8.71	8.35	8.09	8.33	8.50	8.71	8.41			
b _{4Algae} at 0.2%	8.04	8.28	8.41	8.72	8.36	8.10	8.34	8.51	8.72	8.42			
Mean (A)	7.95	8.12	8.27	8.51		7.97	8.17	8.34	8.53				
		Α	В	AB			Α	В	AB				
New L.S.D at 5 %		0.05	0.05	0.10			0.06	0.05	0.10				
	Perce	ntage of fruit p	ulp %										
	Ca an	Ca and glycine treatments (A)											
Algae extract	2016/2	2017			2017/2018								
Algae extract Treatments	\mathbf{a}_1	a ₂	a ₃	a 4		\mathbf{a}_1	a ₂	a ₃	a 4				
(B)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)			
b _{1Algae} at 0.0%	73.11	73.66	74.15	74.60	73.88	73.31	74.00	74.50	75.80	74.40			
b _{2Algae} at 0.0%	73.70	74.5	75.0	76.0	74.80	73.90	74.91	75.81	76.81	75.38			
b _{3Alga} at 0.0%	74.11	75.91	76.90	77.77	76.17	74.31	76.81	77.82	79.82	77.19			
b _{4Algae} at 0.0%	74.12	75.92	76.91	77.75	76.18	74.32	76.83	77.83	79.85	77.21			
Mean (A)	73.76	75.00	75.74	76.54		73.96	75.64	76.49	78.06				
		Α	В	AB			Α	В	AB				
New L.S.D at 5 %		0.21	0.19	0.38			0.23	0.20	0.40				

Table (9):- Effect of single and combined applications of Ca, glycine and Algae extract Finger width (cm) and
The percentage of fruit pulp %) of Williams banana plants during 2016/2017 and 2017/2018 seasons

 Table (8):- Effect of single and combined applications of Ca, glycine and Algae extract on The percentage of fruit peels and The pulp /peel in finger of Williams banana plants during 2016/2017 and 2017/2018 seasons

 Fruit peal %

Fruit peal %													
	Ca and	l glycine treatr	nents (A)										
Algae extract	2016/2	017				2017/2018							
Treatments	\mathbf{a}_1	a ₂	a ₃	a ₄		a ₁	a ₂	a 3	a ₄				
(B)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)			
b _{1Alga} at 0.0%	26.89	26.34	25.83	25.40	26.12	26.69	26.00	25.50	24.20	23.60			
b _{2Algae} at 0.05%	26.30	25.50	25.00	24.00	25.20	26.00	25.09	24.19	23.19	24.62			
b _{3Algae} at 0.1%	25.89	24.09	23.10	22.23	23.0	25.69	23.19	22.18	20.18	22.81			
b4Algaeat 0.2%	25>85	24>08	23.09	22.22		25.68	23.18	22.17	20.15	22.80			
Mean (A)	26.24	25.00	24.26	23.46		26.02	24.37	23.50	21.93				
		Α	В	AB			Α	В	AB				
New L.S.D at 5 %		0.22	0.20	0.40			0.20	0.18	0.36				
	Pulp/P	eel of finger											
	Ca and	Ca and glycine treatments (A)											
Algae extract	2016/2	017	-		2017/2018								
Treatments	a 1	a ₂	a 3	a 4		a 1	a ₂	a 3	a 4				
(B)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)			
b _{1Algae} at 0.0%	2.72	2.80	2.87	2.94	2.83	2.75	2.85	2.92	3.13	2.91			
B2Algae at 0.05%	2.80	2.92	3.00	3.17	2.97	2.83	2.99	3.13	3.31	3.07			
B _{3Algae} at 0.1%	2.86	3.15	3.33	3.50	3.21	2.89	3.31	3.51	3.96	3.42			
B4Algae at 0.2%	2.86	3.15	3.33	3.50	3.21	2.89	3.31	3.51	3.96	3.42			
Mean (A)	2.81	3.01	3.13	3.28		2.84	3.12	3.27	3.59				
		Α	В	AB			Α	В	AB				
New L.S.D at 5 %		0.05	0.04	0.08			0.05	0.05	0.10				

Starch %							2					
	Ca an	d glycine treat	ments (A)									
Al	2016/2017						2017/2018					
Algae extract Treatments	\mathbf{a}_1	a ₂	a 3	a ₄		\mathbf{a}_1	a ₂	a 3	a ₄			
(B)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)		
b _{1Alga} at 0.0%	1.62	1.50	1.38	1.25	1.44	1.66	1.50	1.35	1.20	1.43		
b _{2sAlgae} at 0.0%	1.49	1.25	1.12	0.99	1.21	1.49	1.35	1.20	1.03	1.27		
b _{3Algae} at 0.0%	1.36	1.12	0.99	0.85	1.08	1.29	1.03	0.90	0.77	1.01		
b _{4Algae} at 0.0%	1.35	1.11	0.98	0.84	1.07	1.29	1.02	0.88	0,76	0.99		
Mean (A)	1.46	1.25	1.12	0.98		1.43	1.23	1.10	0.94			
		Α	В	AB			Α	В	AB			
New L.S.D at 5 %		0,07	0.06	0.12			0.05	0.06	0.12			
	T.S.S	%										
	Ca an	Ca and glycine treatments (A)										
Algae extract	2016/2017						2017/2018					
Treatments	\mathbf{a}_1	a ₂	a 3	a 4		\mathbf{a}_1	a ₂	a 3	a 4			
(B)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine			
b _{1Algae} at 0.0%	18.11	18.41	18.85	19.20	18.54	18.20	18.50	18.94	19.30	18.74		
b2seaweed at 0.05%	18.50	19.00	19.50	20.00	19.25	18.61	19.10	19.60	20.11	19.36		
b _{3Algae} at 0.1%	19.00	19.49	20.00	20.25	19.69	19.10	19.60	20.20	20.50	19.85		
b _{4Algae Algae} at 0.2%	19.04	19.50	20.01	20.27	19.71	19.11	19.61	20.22	20.52	19.87		
Mean (A)	18.72	19.24	19.85	21.57		18.76	19.13	19.74	20.11			
		Α	В	AB			Α	В	AB			
New L.S.D at 5 %		0.10	0.09	0.18			0.11	0.10	0.20			

Table (9):- Effect of single and combined applications of Ca, glycine and Algae extract on The percentage of	f
starch and The T.S.S % in The pulp of Williams banana plants during 2016/2017 and 2017/2018 seasons	

Table (10):- Effect of single and combined applications of Ca, glycine and Algae extract on The percentage of total and reducing sugars in % in The pulp of Williams banana plants during 2016/2017 and 2017/2018 seasons

Total sugars %												
	Ca an	d glycine treat	ments (A)									
Al	2016/2017						2017/2018					
Algae extract Treatments	\mathbf{a}_1	a ₂	a 3	a 4		a_1	a ₂	a 3	a ₄			
(B)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)		
b _{1Algae} at 0.0%	15.11	15.35	15.60	15.85		15.20	15.44	15.70	15.95			
b _{2sAlgae} at 0.05%	15.36	15.60	15.86	16.20		15.46	15.87	15.87	16.30			
b _{3Algae} at 0.1%	15.60	15.86	16.30	16.60		15.71	15.96	16.39	16.69			
b _{4Algae} at 0.2%	15.61	15.87	16.31	16.61		15.72	15.97	16.40	16.70			
Mean (A)												
		Α	В	AB			Α	В	AB			
New L.S.D at 5 %		0.14	0.12	0.24			0.12	0.11	0.22			
	Reduc	ing Sugars %										
	Ca an	Ca and glycine treatments (A)										
Algae extract	2016/2017						2017/2018					
Algae extract Treatments	\mathbf{a}_1	a ₂	a 3	a 4		a_1	a ₂	a 3	a ₄			
(B)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)		
b _{1Algae} at 0.0%	5.01	5.16	5.32	5.50		4.99	5.20	5.36	5.55			
b _{2Algae} at 0.05%	5.17	5.33	5.50	5.66		5.21	5.37	5.55	5.71			
b _{3Algae} extract extracts at 0.1%	5.35	5.55	5.71	5.91		5.40	5.60	5.76	5.96			
b _{4AlgaeAlgae} at 0.2%	5.36	5.56	5.72	5.92		5.41	5.61	5.77	5.97			
Mean (A)												
		Α	В	AB			Α	В	AB			
New L.S.D at 5 %		0.09	0.07	0.14			0.08	0.06	0.12			

Non-reducing sugars	s %											
	Ca an	d glycine treat	ments (A)									
Al	2016/2017						2017/2018					
Algae extract Treatments	\mathbf{a}_1	a ₂	a ₃	a 4		a 1	a ₂	a 3	a 4			
(B)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)	Non	Carboxylic Ca 0.1%	Glycine 0.1%		Mean (b)		
b _{1Algae} at 0.0%	10.10	10.19	10.28	10.35	10.23	10.21	10.24	10.34	10.40	10.25		
b _{2Algae} at 0.0%	10.19	10.27	10.36	10.54	10.34	10.25	10.34	10.32	10.59	10.38		
b _{3Algae} at 0.0%	10.25	10.31	10.59	10.69	10.46	10.31	10.36	10.63	10.73	10.51		
b _{4Algae} at 0.0%	10.25	10.31	10.59	10.69	10.46	10.31	10.36	10.63	10.73	10.51		
Mean (A)	10.20	10.27	10.46	10.57		10.27	10.33	10.48	10.61			
		Α	В	AB			Α	В	AB			
New L.S.D at 5 %		0.06	0.04	0.08			0.03	0.04	0.08			
	Titrat	able acidity %										
	Ca an	Ca and glycine treatments (A)										
Algae extract	2016/2017						2017/2018					
Treatments	a 1	a ₂	a ₃	a ₄		a ₁	a ₂	a ₃	a ₄			
(B)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)		
b _{1Algae} at 0.0%	0.399	0360	0.330	0.300	0.347	0.401	0.359	0.339	0.299	0.347		
b _{2Algae} at 0.05%	0.360	0.330	0.300	0.200	0.314	0.358	0.329	0.298	0.265	0.313		
b _{3Algae} at 0.1%	0.330	0.300	0.260	0.230	0.280	0.329	0.299	0.257	0.228	0.278		
b _{4Algae} at 0.2%	0.329	0.299	0.259	0.228	0.278	0.328	0.298	0.256	0.227	0.277		
Mean (A)	0.355	0.322	0.287	0.255		0.353	0.321	0.285	0.254			
		Α	В	AB			Α	В	AB			
New L.S.D at 5 %		0.014	0.015	0.030			0.016	0.014	0.028			

Table (11):- Effect of single and combined applications of Ca, glycine and Algae extract on The percentage of non-reducing sugars and titratable acidity in The pulp of Williams banana plants during 2016/2017 and 2017/2018 seasons

Table (12):- Effect of single and combined applications of Ca, glycine and Algae extract extracts on The percentage of K and Mg of Williams banana plants during 2016/2017 and 2017/2018 seasons

Leaf K %													
	Ca and glycine treatments (A)												
Algae extract	2016/2017						2017/2018						
Algae extract Treatments	\mathbf{a}_1	a ₂	a 3	a 4		\mathbf{a}_1	a ₂	a 3	a 4				
(B)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)			
b _{1Algae} at 0.0%	2.79	2.90	3.01	3.12	2.96	2.74	2.91	3.02	3.13				
b _{2sAlgae} at 0.0%	2.91	3.05	3.16	3.30	3.11	2.91	3.06	3.16	3.29				
b _{3Algae extract extracts} at 0.0%	3.03	3.20	3.30	3.42	3.24	3.06	3.25	3.35	3.46				
b _{4Algae} at 0.0%	3.04	3.20	3.31	3.43	3.25	3.07	3.26	3.35	3.46				
Mean (A)	2.94	3.09	3.12	3.32		2.95	3.12	3.22	3.24				
		Α	В	AB			Α	В	AB				
New L.S.D at 5 %		0.06	0.05	0.10			0.05	0.06	0.12				
	Leaf	Mg %											
	Ca a	Ca and glycine treatments (A)											
Algae extract	2016	/2017			2017/2018								
Treatments	a_1	a ₂	a 3	a 4		a_1	a ₂	a 3	a 4				
(B)	Non	0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)	Non	Carboxylic Ca 0.1%	Glycine 0.1%	Both Carboxylic Ca & glycine	Mean (b)			
b _{1Algae} Algaeat 0.0%	0.41	0.48	0.56	0.70	0.54	0.37	0.51	0.59	0.71	0.55			
b _{2sAlgae} at 0.05%	0.50	0.61	0.66	0.76	0.63	0.52	0.64	0.70	0.81	0.67			
b _{3Algae} at 0.1%	0.57	0.71	0.76	0.83	0.72	0.61	0.75	0.85	0.96	0.79			
b _{4Algae} at 0.2%		0.72	0.77	0.84	0.73	0.62	0.76	0.86	0.98	0.81			
Mean (A)	0.53	0.63	0.69	0.78		0.53	0.67	0.75	0.87				
		Α	В	AB			Α	В	AB				
New L.S.D at 5 %		0.04	0.03	0.06			0.06	0.05	0.70				

4. Discussion

1- Effect of amino acids:-

Amino acids as organic nitrogenous compounds are the building blocks in the synthesis of proteins, which are formed by as process in which ribosomes catalyze the polymerization of amino acids (Davies, 1982). Several hypothesis have been proposed the explain for the role of amino acids in plant. Available evidence suggests several alternative routes of IAA and ethylene synthesis in plants, starting from amino acids In this respect, (Taiz and Zeiger 2002) suggested that the regulatory effect of certain amino acids like phenylalanine and ornithine in plant development appeared through their influence on the biosynthesis of gibberellins, tryptophan and methionine in building of I.A.A and ethylene respectively. These results are in agreement with those obtained by Ahmed et el (2014); El-Khawaga, (2014), Hassan-Huda, (2014); Sayed-**Ola,2014 and Mohamed (2016).**

2- Effect of Calcium:_

Calcium has been known as an essential element for trees for a long time and it has received much attention from the fertility standpoint and also very important for facilitating marketing of the fruits. Calcium is required for cell division and chromosome stability. As Ca-pectate, it is a constituent of the middle lamella of cell walls (**Dodd** *et el*,2010). Calcium indirectly influences many enzyme systems such as amylase and ATPase. It also seems to have a regulating role in respiration and prolonging shelf life of fruits (**Marschner**, 2010).

The results of Joutamance et el (2002); El-Shafey et el (2002); El-Tanany et el (2004) Young Ho et el (2004); Sarawy et el (2012); Chakerolhasseini et el,2016; Habasy- Randa et el (2016); Mohamrd et el (2017); Meena et el (2017) and Hikal et el (2017) supported the present results.

3- Effect of Algae extract

The application of Algae extract which contains most nutrients, organic compound, enzyme, vitamins, antioxidants, amino acids and natural hormones is fast becoming an accepted practice. It increases yield quantitatively and qualitatively in various fruit crops (Soliman *et al.*, 2008 and Spinelli *et al.*, 2010).

These results are in agreement with those obtained by Ebeid- Sanaa,2007); El-Sawy,2008), (Gamal,2013) (Mohamed and, El-Sehrawy, (2013b); Oraby, (2013a); Ahmed *et el*, (2014); Eshmawy, (2015); Abd El-aaty,2015) and Ahmed *et el* (2015).

Conclusion

For promoting yield and fruit characteristics of Williams banana grown under Minia region

conditions, it is suggested to use a mixture of carboxylic -Ca and glycine plus Algae extract each at 0.1% three times a year at the first week of May and at one month intervals.

References

- 1. Abd El-aaty, M.S.H. (2015): Relation of fruiting in Sakkoti and Bartemoda date palms with spraying seaweed extract. M.Sc. Thesis Fac. of Agric. Minia Univ. Egypt.
- Adam, M. S. (1999): The promotive effect of cyanobacteria *Nostoc muscorum* on the growth of some crop plants- Acta Microbiologioolinica, 48 (2): 163-171.
- Ahmed, F. F. and Morsy, M. H. (1999): A new method for measuring leaf area in different fruit species. Minia J. of Agric. Res. & Develop., Vol. (19) pp 97-105.
- Ahmed, F.F., Akl, M.M.A. and Oraby A.A.F. (2013a): Partial replacement of inorganic nitrogen fertilizer by spraying some vitamin, yeast and seaweed extract in Ewaise mango orchard under Upper Egypt conditions. Stem Cell (3): 110-120.
- Ahmed, F.F.; Mohamed, H.A.A and Gad El-Kareem, M.R. (2014a): The promotive effect of seaweed extract on fruiting of Zaghloul date palms grown under Minia region. 5th International Date Palm Conf, 16-18 Mar. Emirates Palace Abu Dhabi United Arab Emirates.
- Ahmed, F.F.; Ali, A.H.S.; Sayed, E.S. and Sayed- Ola, M.O. (2014b): Using some amino acids enriched with certain nutrients for improving productivity of El-Saidy date palms. World Rural Observations. 6 (2)20-27.
- 7. Ahmed, F.F.; Gobara, A.A.; Abo El- Komsan, E.E. and Gamal, A.F. (2008): Growth and fruiting of Washington Navel orange trees as affected by some antioxidant and Algae extract treatments. Inter. Conf. for Environ.
- 8. Ahmed, F.F.; Mohamed, M.A.; Mohamed A.Y. and Abd El-aaty, M.S. (2015): Response of Sakkoti and Bartemoda date plms to spraying seaweed extract. World Rural Observations.
- Ahmed, F.F.; Mansour, A.E.M.; Abd El-Rahman, M.A. Merwad, M.A. and Mostafa, K.A. (2013): Response of Valencia orange trees to foliar application of roselle, turmeric and seaweed extracts. J. of Applied Sci. Res. 9 (I): 960-964.
- Association of Official Agricultural Chemists (2000): Official Methods of Analysis (A.O.A.C), 12th Ed., Benjamin Franklin Station, Washington D.Q, U.S.A. pp. 490-510. 6 (3): 72-78.

- Black, C.A. (1965): Methods of Soil Analysis. Amer. Soc. of Agron., Madison, Wisconsin, U.S.A. pp 1 - 20.
- 12. Chakerolhosseini, M. R., Khorassani, R., Fotovat, A., and Basirat, M. (2016): Effect of foliar spray of calcium and zinc on yield, nutrients concentration and fruit quality of orange. IIOAB Journal, 7 (8): 124-129.
- Chapman, H. D. and Pratt, P. P. (1965): Methods of Analysis for Soils, Plants and Water. Univ. of California. Division of Agric., Sci. 172-173.
- Cottenie, A.; Verloo, M.; Kiekens, L.; Velgle, G. and Amerlynuck, R. (1982): Chemical Analysis of Plant and Soil. 34-51. Laboratory of Analytical and Agroch. State Univ. Belgium, Gent.
- Davies, D.D. (1982): Physiological aspects of protein tumour Encycl. Plant physiol. New series (nucleic acids and proteins, structure, biochemistry and physiology of proteins). Springer Verla, Berlin, New York, pp. 190-228.
- 16. Dodd, A. N., Kudla, J., and Sanders, D. (2010): The language of calcium signaling. Annual review of plant biology, 61, 593-620.
- Ebeid Sanaa (2007): The promotive effect of seaweed extract and boron on growth and fruiting of Hindy Bisinnara mango trees. Minia J. of Agric. Res. & Develop. (27): 3 pp 579-594.
- El-Khawaga, A.S. (2014): Impact of vitamins B and C, glutamic acid and silicon on fruiting of Superior grapevines. World Rural Observations. 6 (4): 57-62.
- El-Shafey, Y. H., El-Rahman, A. A., and El-Azaze, A. M. (2002): Effect of foliar application with calcium on yield and fruit quality of Valencia orange trees. Bulletin-Faculty of Agriculture, Cairo University, 53(2), 275-288.
- El- Tanany, M. M.; Abdel Messih, M. N. and Shama, M. A. (2011): Effect of foliar application with potassium, calcium and megnesium on yield, fruit quality and mineral composition of Washington Navel orange trees. Alex Sci. Exchange 32 (1): 65-72.
- 21. Eshmawy, E.M.Sh. (2015): relation of fruiting in Saeidy date Palm with spraying salicylic acid and seaweed extract. Ph. D. Thesis Fac. of Agric. Minia Univ.
- 22. Gamal, A. P.O. (2013): Fruiting of Washington Navel orange trees in relation to application of Seaweed extract Boron and citric acid Ph.D. Thesis Fac. of Agric. Minia Univ., Egypt.

- Habasy, R. E. Y.; Helal, M. E.; El-Rahman, A. M. A.; Ahmed, F. F. (2016): Effect of calcium and boron sources and methods of application on growth yield and fruit quality of Washington Navel orange trees. Arab Universities Journal of Agricultural Sciences 2016 Vol. 24 (1): 185-193.
- 24. Hassan- Huda, M.1. (2014): Impact of effective microorganisms and amino acids enriched with some nutrients on growth and fruiting of Valencia orange trees. Ph. D. Thesis Fac. of Agric. Minia Univ, Egypt.
- Hikal, A. R. F.; Ibrahim, M. A. and Abdelaziz, R. A. (2017): Effect of Different Treatments of Calcium and Boron on Productivity and Fruit Quality of Navel Orange Fruits. Egypt. J. Hort. Vol. 44 (1): 119- 126.
- 26. James, B. (1994): Chapters from life Ann. Rev. Physico. Plant Mol. Biology 45: 1-23.
- Jutamance, K. Eoomkham, S. Puichakum, A. Krisanapook and Phavahutanon, K. (2002): Effect of calcium. boron and sorbital on pollution and fruit set in mango cv. Nandokami. Acta. Hort. 575 (2): 829 - 934.
- Kullk, M. M. (1995): The potential cyanobacteria (blue green algae) and algae for using in the biological control of plant. Pathogenic Bacteria and Fungi European J. of plant pathology, 101(6): 585-599.
- 29. Lane, J. H. and Eynon, L. (1965): Determination of reducing sugars by means of Fehlings solution with methylene blue as indicator A.O.AC. Washington D.C.U.S.A. pp.490-510.
- Mead, R.; Currnow, R. N. and Harted, A. M. (1993): Statistical Methods in Agricultural and Experimental Biology. 2nd Ed. Chapman and Hall, London pp. 10- 44.
- 31. Meena, M. K.; Jain, M. C.; Singh, J and Sharma, M. (2017): Effect and Economic Feasibility of Preharvest Spray of Calcium Nitrate, Boric Acid and Zinc Sulphate on Yield Attributing Characters of Nagpur Mandarin (*Citrus Reticulata Blanco*). Horticulture International Journal. 1 (1).
- Mohamed, M.A.A. (2016): Physiological studies on the effect of some silicon, boron and amino acid treatments on some olive acid. Ph. D. Thesis Fac. of Agric. Al- Azhar Univ. Assiut branch Egypt.
- Mohamed, A. Y. and El- Sehrawy, O. A. M. (2013): Effect of seaweed extract on fruiting of Hindy Bisinnara mango trees. J. of Arner. Sci. 9 (6): 537 - 544.
- 34. Mohamed, M. A. A., El-Khalek, A. A., and Elmehrat, H. G. (2017): Pre-harvest Potassium

Silicate, Chitosan and Calcium Chloride Application Improves Mango Fruits (cv. Zebda) Quality and Storability. Egypt J. Horti. 44 (1): 17-32.

- 35. Oraby, A.A.F. (2013): Partial replacement of inorganic nitrogen fertilization by spraying some vitamins, yeast and seaweed extract in Ewaise mango orchards under Upper Egypt conditions. M. Sc. Thesis Fac. of Agric. Minia Univ. Egypt.
- Ordog, V. Stirck, W.A.; Lenobel, R.; Bancirova, M.; Van Staden, J.; Szigeti, J. and Nemeth, L. (2004): Screening microalgae for some potentially useful agricultural and pharmaceutical secondary metabolites. J. Appl. Phycol. 16:309-314.
- Saffan- Samia, E. (2001): Allelopathic effects of cyanobacterial exudates on some metabolic activities of Cyanara cardunculus seeds during germination, Egypt J. of Botechnol Vol. 10, July, pp. 157 - 178.
- Sarawy, S.M.A.; Gadalla, E.G. and Mostafa, E.A. (2012): Effect of calcium nitrate and boric acid on fruit set and fruit quality of cv. Amhat date palm. World. Agric. Sci. 8 (5): 506-515.
- 39. Sayed- Ola, M.O. (2014): Effect of certain amino acids enriched with some nutrients on growth and fruiting of El- Saidy date palms growing under new valley governorate climatic conditions. M.Sc. Thesis Fac. of Agric. Minia Univ. Egypt.
- Seleem Basma, M and Abd El- Hameed, H. M. (2008): Effect of the stimulant aminoquelant- Ca on yield and berries quality of Thompson seedless grapevines. Minia. J. of Agric. Res. & Develop. Vol. (28) No. 1 pp 13 -21.

12/16/2018

- Spinelli, F.; Fieri, G.; Noferini, M.; Sprocatti. M.; and Costa, G. (2010): Perspectives on the use of a seaweed extract to moderate the negative effects of alternate bearing in apple trees. Journal of Hort. Sci. & Biot. (2009) p. 131-137.
- 42. Strick, W. A.; Staden, J.; Vani and Staden J. (1997): Screening of some South African seaweeds for cytokinin- like activity. South African J. of Botany, 40(6): 161-164.
- 43. Summer, M.E. (1985): Diagnosis and Recommendation Integrated System (DRIS) as a guide to orchard fertilization. Hort. Abst. 55(8): 7502.
- 44. Tung Yunn, H. O.; Quigg, A.; Finkel, Z. V.; Milligan, A. J.; Wgman, K.; Falkowski, P. G. and Morel, F. M. M. (2003): The elemental composition of some marine phytoplankton. J. of Phycology (39),1: 10-20.
- 45. Wani, S. P. and Lee, K. K. (1995): Microorganisms as biological for sustainable agriculture In organic agriculture, theory and practices (ed.) P.K. Thampan, Peekay Tree Crops Development Foundation, Gandhi Nagar Cochin 682 - 220, p. 36-67.
- 46. Wilde, S. A.; Corey, R. B.; Layer, J. G. and Voigt, G. K. (1985): Soils and Plant Analysis for Tree Culture. Oxford and IBH publishing Co., New Delhi, India.
- 47. Young Ho, K.; M. Young Eel and H. Seung Gab (2004): Effect of calcium formulae foliar application on the water spot out break and fruit quality of Satsuma mandarin in the plastic house. Korean Journal of Hort. Sci. & Technology. 22 (1): 50-54.