**Potential role of Arginine, Glutamine and Taurine in ameliorating osteoporotic biomarkers in ovariectomized rats**

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 **Abstract**

The main purpose of the present study was to evaluate the role of some amino acids namely L-arginine, L-glutamine and taurine in the management of osteoporosis in ovariectomized (OVX) rats. The current study included six groups of female Sprague Dawley rats which were classified as gonad intact control group and five ovariectomized groups: one untreated group served asovariectomized control group another ovariectomized group orally administered with 10% lactose, three ovariectomized groups orally administered with each amino acid dissolved in 10% lactose. The treatment was started after 3 months of ovariectomy and continued for other 3 months. Serum parathyroid hormone (PTH), 1,25 dihydroxyvitamine D3 levels were determined. Insulin like growth factor-1 (IGF-1) and transforming growth factor-β (TGF-β) levels were also estimated. Bone mineral density (BMD) and bone mineral content BMC) of right femur bone of each rat were measured using DEXA technique. Also, histological investigation of the bone sections of left femur of each rat was carried out. The obtained data revealed that ovariectomy decreased serum 1,25 (OH)2 D3, IGF-1 and TGF-β levels whereas, it increased serum PTH level. DEXA results revealed that ovariectomy decreased BMD and BMC of the proximal, distal and mid areas of rat femur bone. These results were well documented by bone histological examination. The selected amino acids could improve all the studied bone biochemical markers significantly. DEXA results also showed that treatment with these amino acids could increase both BMD and BMC of rat femur bone in most areas. The photomicrogrophs of femur bone sectionsof rats treated with the selected amino acids supported the present improvement in bone biomarkers. In conclusion, each of the selected amino acids exhibited antiosteoporotic effects due to the anabolic and/or antiresorptive activity. Taurine produced more pronounced effect on bone biomarkers than the other amino acids. These encouraging results provide new concepts for the development of effective opportunities in the treatment of primary osteoporosis.

**Key words:** Osteoporosis, L-arginine, L-glutamine, taurine, lactose, bone biomarkers, bone mineralization.

**Introduction**

Osteoporosis is a global health problem that will take an increasing significance as people live longer and the world’s population continues to increase in number, thus the manegment of osteoporosis and its complications is an socioeconomic priority **(Kevin, 2007)** Osteoporosis is defined as decreased bone strength and increased susceptibility to fractures (**Preisinger , 2009)**. It is also defined as progressive systemic skeletal disease characterized by low bone mass with a consequent increase in bone fragility and susceptibility to fracture.**(Katherine et al., 2007).** So, there is an urgent need to develop and implement alternative nutritional approaches and policies for treatment of osteoporosis **(Kevin,2007).** This idea comes fromthe fact thatprotein under nutrition is known to play an important role in the pathogenesis of osteoporotic fracture. The mechanisms underlying the bone loss in protein under nutrition appeared to be related to an uncoupling between increased bone resorption and bone formation. This was associated with decreased plasma insulin-like growth factor-1 (IGF-1) level, with anoestrus and decreased muscle mass. Nutritional intervention with amino acid supplements can increase bone mineral mass, bone strength and muscle mass in osteoporotic subjects **(Ammann et al., 2000).**

Amino acids are the building blocks of protein. Essential amino acids (EAA) can

modulate the growth and the differentiation of osteoblasts cultured in vivo, confirming the relationship between osteoporotic hip fracture and inadequate protein intake. Amino acids have mainly enhanced cell growth and alkaline phosphatase activity, and, to a lower degree, collagen synthesis **(Conconi et al.,. 2001).**

Amino acids supplement increased bone mineral mass and strength in ovariectomized protein-deprived rats. This was associated with stimulated bone formation and reduced bone resorption, with an increment of plasma isulin-like growth factor (IGF-1) and limb muscle mass weight **(Ammann et al., 2000).**

 L-arginine represents a key building block to repair damage tissue and bone. Athletes have also found L-arginine to be beneficial for muscle recovery and growth hormone (GH) release from pituitary gland. Oral administration of L-arginine in pharmacological doses induces growth hormone and insulin –like growth factor-1 responses and stimulates nitric oxide synthesis **(Baecker et al., 2005)**. Growth hormone and insulin –like growth factor-1 are important mediators of bone turnover and osteoblastic bone formation, while nitric oxide is a potent inhibitor of osteoclastic bone resorption. Because of this dual effect on physiological regulators of bone remodeling, L-arginine could potentially increase bone formation over bone resorption, and consequently, increase bone mass **(Clementi et al., 2001)**.

Glutamine**,** has a number of unique properties suggesting that this amino acid plays an important role in health and disease.This amino acid makes more than 60% of the skeletal muscle tissue, and it is a fuel for both the digestive tract and the immune system. Also, it is playing a pivotal role in conducting nitrogen to muscle around the body **(Tapiero et al., 2002).** Glutamine may at least in part play a role in mechanisms associated with cellular proliferation and/or differentiation through particular glutamine receptors (GluR) and glutamine transporters functionally expressed in rat calvarial osteoblasts **(Yoneda and Hinoi, 2003).** The cyclization of glutamate produces proline, an amino acid important for synthesis of collagen and connective tissue **(Tapiero, et al. 2002).** Therefore, it has been suggested that glutamine may have a role in the process of bone formation.

Significant amount of taurine is transported to bone tissue, it is reasonable to propose that taurine may play an important role in bone metabolism. Interestingly, taurine has been found to inhibit experimental bone resorption and osteoclast formation and survival **(Koide et al., 1999).** It has inhibitory effects on bacteria-stimulated osteoclast formation *in vitro*. Moreover, this amino acid has stimulatory actions on alkaline phosphatase activity and collagen synthesis **(Park et al., 2001).** It may play a role in osteoblastic differentiation as well as bone matrix formation. Taurine has anti-osteopenic effect in low Ca diet-induced osteopenia in rats, theraby promoting mineralization and finally leading to its bone anabolic action **(Yasutomi et al., 2002).**

 Pharmacological mixture containing amino acids and lactose accelerates and ameliorates bone fracture healing processes. This finding is linked not only to calcium metabolism but also to different biological properties which positively contribute to good healing of bone fractures **(Fini et al ., 1996) .**

 The principal goal of the current study was to develop alternative nutritional therapeutic modalities for the treatment of primary osteoporosis in order to avoid the serious side effects of the traditional hormone replacement therapy for osteoporosis in postmenopausal women. The suggested therapeutic opportunity included the supplementation of some promising amino acids The selected amino acids include L-arginine, L-glutamine, or taurine as effective dietary supplements for management of primary osteoporosis.

**Materials and Methods:**

**Amino acids:** L-arginine, L- glutamine, taurine and lactose were purchased from Sigma Company (U.S.A). **Experimental animals:** Adult female Sprague Dawley rats (120-150g) were obtained from Animal House Colony of the National Research Centre , Cairo, Egypt. The animals were kept in wire bottomed cage at room temperature (25± 2 ºC) under a 12h dark- light cycle and acclimated to the laboratory environment for seven days before use. Animals were fed with standard laboratory diet and water *ad libitum.* The rats were ovariectomized surgically in Hormone Department, Medical Research Division at the National Research Centre. Then, after three months following surgery, the animals were divided into 6 groups as follows: The first group; was untreated (OVX) rats and served as (OVX) control. The second group; (OVX) rats which were orally administered with (1ml/rat/day) lactose 10%. The third group; (OVX) rats which were orally administered with L-arginine dissolved in 10%lactose in a dose of 500mg/kg/day **(Gupta et al., 2005).** The fourth group; (OVX) rats which were orally administered with L-glutamine dissolved in 10% lactose in a dose of 3.2g/kg/day **(Ann et al., 2004).** The fifth group; (OVX) rats which were orally administered with taurine dissolved in 10% lactose in a dose of 50mg/kg/day **(Cetiner et al., 2005)**. Additional untreated gonad intact control group was involved in the present study. The experiment lasted for 3 months.

At the end of the experimental period, the animals were kept fasting for 12 hours and the blood samples were collected from the retro-orbital venous plexus under diethyl ether anesthesia **(Schermer, 1967)**. The blood samples were left to clot and the serum were separated by cooling centrifugation (4º C) at 3000 rpm for 10 min. Serum parathyroid hormone (PTH) was estimated by ELISA procedure according to the method described by **Blum et al. (1993)**. Serum 1, 25-dihydroxyvitamin D3 (Vitamin D3) was determined by Radio immuno assay (125I RIA) according to the method of **Hollis (1986).** Serum insulin-like growth factor-1 (IGF-1), transforming growth factor-β (TGF-β) were determined using ELISA procedure according to the method of **Blum et al. (1993) and Kim et al. (1994)** respectively. The right femur bone of each animal was dissected, cleaned and stored in formalin buffer 10% for measuring bone mineral density (BMD) and bone mineral content (BMC) using dual energy X-ray absorptiometry (DEXA). The left femur bone was also carefully removed cleaned and stored in 10% formic acid solution as a decalcifying agent for 10 days for histological investigation.

**Histological Examination:**

The left femur was embedded in paraffin wax and the microscopic sections of 5μm intervals were taken and stained with hematosylin and eosin (H & E) for histological examinations (**Drury and Wallington, 1980**).

***Statistical Analysis***

 In the present study, all results were expressed as mean **+** S.E of the mean. Data were analyzed by one way analysis of variance (ANOVA) using the Statistical Package for the Social Sciences (SPSS) program, version 11 followed by least significant difference (LSD) to compare significance between groups (**Armitage and Berry, 1987**). Difference was considered significant when *P* value ≤ 0.05.

**Results:**

1. **Effect of amino acids supplementation on PTH and vitamin D:**

Our data indicated that ovariectomy induced significant increase in serum PTH level associated with significant decrease in serum 1,25 (OH)2D3 level in comparison with gonad intact control group. Treatment of ovariectomized rats with arginine, glutamine or taurine caused significant decrease in PTH serum level and significant increase in 1,25 (OH)2D3 serum level comparing with ovariectomized rats that administered lactose only (Table 1).

**Table (1): Effect different amino acids supplementation on serum Parathyroid hormone (PTH) and 1,25 (OH)2D3levels in ovariectomized rats.**

|  |  |  |
| --- | --- | --- |
| **Parameters****Groups** | PTHPg/ml |  1,25 (OH)2D3 (Pg/ml)  |
| Gonad intact control | 36.6±2.8 | 15.7±0.3 |
| OVX Control | 62.5±2.4 a  | 12.8±0.2 a |
| OVX + Lactose | 60.4±2.1 | 13.2±0.25 |
| OVX + Arg.  | 50.2±2.4 b | 15.3±0.34 b |
| OVX + Glut.  | 49.4±1.7 b | 15.2±0.18 b |
| OVX + Tau.  | 45.9±2.8 b | 15.7±0.14 b |

a : Significant change at *P*˂ 0.05 in comparison with gonad intact control.

b : Significant change at *P*˂ 0.05 in comparison with ovariectomized received lactose group.

OVX.: Ovariectomized rats, Arg.: Arginine, Glu.: Glutamine, Tau.: Taurine

1. **Effect of amino acids supplementation on IGF-1 and TGF-β:**

The present data revealed that ovariectomy resulted in significant decrease in each of IGF-1 and TGF-β in serum as compared to gonad intact control group. Treatment with the tested amino acids produced significant increase in IGF-1 and TGF-β serum levels in comparison with ovariectomized rats received lactose only as represented in Table (2).

**Table(2): Effect of different amino acids supplementation on serum insulin-like growth factor-1 (IGF-1), and transforming growth factor-β (TGF-β) levels in ovariectomized rats.**

|  |  |  |
| --- | --- | --- |
| **Parameters****Groups** | IGF-1Ng/ml | TGF-βPg/ml |
| Gonad intact control | 9.7±0.7 | 193.9±9.1 |
| OVX Control | 6.7±0.32 a | 125.8±2.7 a |
| OVX + Lactose | 7.0±0.19 | 131.1±2.3 |
| OVX + Arg. | 8.2±0.21 b | 160±4.0 b |
| OVX + Glut. | 8.5±0.16 b | 171.9±2.7 b |
| OVX + Tau. | 8.8±0.22 b | 179.8±2.1 b |

a : Significant change at *P*˂ 0.05 in comparison with gonad intact control.

b : Significant change at *P*˂ 0.05 in comparison with ovariectomized received lactose group.

OVX.: Ovariectomized rats. Arg.: Arginine, Glu.: Glutamine, Tau.: Taurine

1. **Effect of amino acids supplementation on BMD and BMC:**

The results in table (3) showed that ovariectomy decreased BMD of proximal, mid and distal areas significantly in comparison with gonad intact control group. While ovariectomized rats treated with arginine showed significant increase in BMD of proximal and mid areas and insignificant increase in BMD of distal area in comparison with ovariectomized rats received lactose only. Treatment with glutamine or taurine induced significant increase in BMD of proximal, mid and distal areas in comparison with ovariectomized rats received lactose only. Noteworthy, ovariectomized rats treated with taurine produced significant increase in BMD of proximal, mid and distal areas in comparison with untreated ovariectomized group.

**Table (3): Effect of different amino acids supplementation on bone mineral density (BMD) in ovariectomized rats.**

|  |
| --- |
| **BMD** (mg/cm2) |
| **Parameters Groups** | Proximalmg/cm2 | Midmg/cm2 | Distalmg/cm2 |
| Gonad intact control | 126.4±2.5 | 126.2±1.9 | 131.0± 1.5 |
| OVX Control | 109.4±1.4 a | 112.8±2.2 a | 115.2± 2.9 a |
| OVX + Lactose | 110.7±1.5 | 114.3±2.2a | 117.0± 2.1 |
| OVX+ Arg. | 117.2±1.9 b | 117.6±0.7 b | 122.6± 1.0 |
| OVX+ Glut. | 119.3±1.8 b | 119.8±1.0 b | 125.7± 1.0 b |
| OVX +Tau. | 120.7±1.4 b | 120.6±1.4 b | 127.2± 1.9 b |

a : Significant change at *P*˂ 0.05 in comparison with gonad intact control.

b : Significant change at *P*˂ 0.05 in comparison with ovariectomized received lactose group.

OVX.: Ovariectomized rats. Arg.: Arginine, Glu.: Glutamine, Tau.: Taurine

Our data in table (4) indicated that ovariectomy induced significant decrease in BMC of proximal and distal areas and insignificant decrease in BMC of mid area in comparison with gonad intact control group. Treatment with arginine, glutamine or taurine caused significant increase in BMC of proximal area comparing with ovariectomized received lactose only. While amino acids supplementation induced insignificant increase in BMC of mid area as compared to ovariectomized rats received lactose only with respect to the value of BMC of distal area only, glutamine and taurine supplementation showed significant increase, while supplementation with arginine caused insignificant increase as compared to ovariectomized rats received lactose only. It could be also seen that taurine supplementation increased BMC significantly in comparison with supplementation with arginine.

**Table (4): Effect of different amino acids supplementation on bone mineral content (BMC) in ovariectomized rats.**

|  |
| --- |
| **BMC** (mg/cm2) |
| **Parameters****Groups** | ProximalMg | MidMg | Distalmg |
| Gonad intact control | 72.9±1.0 | 208.3±7.0 | 77.6±1.7 |
| OVX. control | 44.9±1.5 a | 182±4.0 | 50.6±2.1a |
| OVX + Lactose | 45.4±1.0 | 186.8±3.2 | 55.8±3.3 |
| OVX + Arg. | 56.7±3.5 b | 191.3±3.7 | 64.1±0.9 |
| OVX + Glut. | 59.2±3.0 b | 195.5±2.1 | 67.0±0.8 b |
| OVX + Tau. | 65.2±1.2 b | 199.2±4.3 | 70.3±2.0 b |

a : Significant change at *P*˂ 0.05 in comparison with gonad intact control.

b : Significant change at *P*˂ 0.05 in comparison with ovariectomized received lactose group.

OVX.: Ovariectomized rats. Arg.: Arginine, Glu.: Glutamine, Tau.: Taurine

**Histlogical Results:**

 Microscopic examination of left femur bone section of gonad intact control rat represented in **Fig.** (**1**) showed a network of bony trabiculae separated by a labyrinth of interconnecting spaces containing bone marrow. The trabiculae composed of irregular lamellae of bone with Haversian systems and lacunae containing osteocytes.

 Thephotomicrograph of left femur bone sectionof untreated ovariectomized control rat in **Fig.** (**2**) showed the reduction of the cortical and trabicular bone thickness. Many of necrotic areas of bone and resorped cavities on the inner surfacehave been also seen.

Micrograph of a longitudinal section of left femur bone of ovariecomized rat received lactose **(Fig. 3)** showed some necrotic areas of bone and the presence of small cavities.

The photomicrograph of left femur sectionof rats treated with arginine showed the new formed bone and increase in the thickness of bone **(Fig. 4)**.

 It is clear from the photomicrographs of left femur bone sectionsof ovariectomized rats treated with glutamine, the presence of calcified cartilage and the increased thickness of the bony trabiculae (**Fig. 5**).

 A section of left femur bone of ovariectomized rat treated with taurine showed the trabiculae appeared as normal form **(Fig. 6)**.

**Figure (1):** Photomicrograph of femur bone section of gonad intact control rat showing a network of bony trabiculae separated by a labyrinth of interconnecting spaces containing bone marrow. The trabiculae composed of irregular lamellae of bone with Haversian systems and lacunae containing osteocytes (H & E X 400).

**Figure (2):** Micrograph of a longitudinal section of left femur bone of ovariecomized rat showing the reduction of the cortical and trabicular bone thickness. Many of necrotic areas of bone and resorped cavities on the inner surface are also seen. Cartilage layer in the trabecular bone is found (H&E x 150).

**Figure (3):** Micrograph of a longitudinal section of left femur bone of ovariecomized rat treated with lactose showing some necrotic areas bone and the presence of small cavities. The erosion of the outer surface of bone is also appeare (H & E X 150).

**Figure (4):** Micrograph of a longitudinal section of left femur bone of ovariecomized rat treated with arginine in lactose showing an increase in the thickness of bone (H & E X 150).



**Figure (5):** Micrograph of a longitudinal section of left femur bone of ovariecomized rat treated with glutamine in lactose showing the increase in the number of trabiculae(H & E X 150).



**Figure (6):** Micrograph of a longitudinal section of left femur bone of ovariecomized rat treated with tuarine showing normal appearance of bone (H & E X 150).

**Discussion**

Osteoporosis is a chronic condition chiefly affecting postmenopausal women, in whom the skeleton loses a significant percentage of its mineralized mass and mechanical resiliency, thereby becoming prone to fracture (**Fan et al., 2005).** Amino acids have been shown to stimulate bone formation and thus, they might be represented useful agents for the prevention and treatment of osteoporosis **(Conconi et al., 2001).**

The present study showed that ovarietomy induced significant decrease in all the tested parameters except PTH which increased significantly comparing with gonad intact control group.These findings are in agreement with **Segal et al, (2003) and Chen et al, (2007).** It could be explained that ovariectomy induced increase in PTH gene expression and parathyroid cell proliferation( **Silver et al., 1999).**The decrease in 1,25 (OH)2D3 serum level in ovariectomized animals may be due to that estrogen loss is the reason of reducing 1,25(OH)2D3 production and calcium absorption in this rat model **(Ash and Goldin,1988),** While, the reduction of serum IGF-1 level may be explained as estrogen stimulates the autocrine secretions of IGF-1 and TGF-β by osteoblasts which are involved in the stimulation of osteoblasts maturation and growth as well as collagen synthesis and alkaline phosphatase secretion (**Kajdaniuk et al., 1999).** Many reports indicated that reduced plasma levels of IGF-1 are associated with estrogen defieiency and in turn osteoporosis in females. **Calo et al, (2000)** reported that ovariectomy resulted in significant reduction in the number of receptors for both epidermal growth factor (EGF) and IGF-1 in female rats. On the other hand, the reduction in TGF-β resulted from a direct effect of estrogen deficiency on bone cells to decrease the secretion of TGF-β with a concomitant decrease in the deposition of newly formed bone (**Finkelman et al., 1992)**.. This possibility was further supported by the findings that 17β-estradiol administration directly stimulated the production of TGF-β by mouse bone cells and also estrogen replacement therapy *in vivo* corrected the TGF-β deficit (**Finkelman et al., 1992)**. Moreover, it has been reported that estrogen deficiency led to a decrease in TGF-β mRNA which may simply be due to the reduction in cancellous bone volume that occurs following ovariectomy **(Westerlind et al., 1994).**

 The current study also showed a reduction in BMC and BMD in ovariectomized animals as compared to the gonad intact control animals. These results may attributed to that estrogen deficiency resulted in rapid bone loss phase which is associated with loss of BMD **(Shen et al., 2000).** It has been reported that the reduced BMD in ovariectomized rats is also associated with a reduction in dry and ash femur weights as well as a decreased femoral breaking force and energy suggesting the increased risk of fracture **(Park et al., 2008).** The mechanism by which estrogen deficiency could induce bone resorption is that loss of estrogen induced enhanced expression of bone resorbing cytokines (interleukin-1 (IL-1), tumor necrosis factor-alpha (TNF-α) **(Kitazawa et al., 1994)**, interleukin-6 (IL-6) **(Passeri et al., 1993)** and macrophage colony stimulating factor (M-CSF) **(Kimble et al., 1996)** by immune cells and osteoblasts in the bone marrow microenvironment **(Matsushita et al., 2008).** These cytokines are crucial for the pathogenetic mechanisms by which estrogen deficiency leads to increase the expression of functional receptor activator nuclear factor kappa B ligand (RANKL) and to enhance bone resorption and bone loss **(Kwan Tat et al., 2004),** via increasing osteoclast number and activity **(Kassam, 2003)** and promotion nuclear factor kappa B, the key transcription factor in osteoclastogenesis **(Ross, 2003).** Ovariectomy-induced bone resorption could be responsible for the decreasing in calcium and mineral content of the whole femur **(Gaumet et al., 1996).**

Our study revealed that lactose administration in ovariectomized rats led to slight inhibition in serum PTH and slight increase in all of the other tested parameters. These results could be explained as lactose slightly increase serum level of Ca and ionized Ca which inhibit the secreation of PTH, leading to increasing the circulating level of 1,25 (OH)2 D3 by stimulating calcitriol synthesis **(Mastaglia et al., 2006).** Moreover the increase in serum IGF-1 and TGF-B levels with lactose administration may be explained as lactose has a stimulatory effect of lactose on osteoblasts growth and the production of several growth factors **(Kirk et al., 1994).**

It has been found that lactose supplementation to ovariectomised rats induced increase in BMD and BMC, which could be explained as dietary lactose increased bone calcification rate and inhibited bone resorption that lead to the improvement in skeletal growth and minerlization in animals fed lactose **(Shortt and Flynn,1991).**

Our data revealed that arginine supplementation to ovariectomized rats induced significant decrease in PTH level while, it showed significant increase in1,25(OH)2D3,IGF-1,TGF-β and both of BMD and BMC when compared with the ovariectomized rats administered lactose only. Arginine via growth hormone has been found to produce marked increase in

(1, 25(OH)2 D3) due to increasing nephrogonous cyclic AMP(NcAMP) **(Ahmed et al., 2003)**. The increase in IGF-1 level may be due to the role of arginine in stimulating IGF-1 production and collagen synthesis in osteoblasts-like cells. It has been suggested that arginine could increase IGF-1 mRNA transcription and alpha (1) collagen mRNA transcripts and thus, arginine may influence bone formation by enhancing IGF-1 production **(Chevalley et al.,1998**). Lactose appeared to have an intangable role in enhancing serum level of IGF-1 in ovariectomized rats via stimulation of osteoblasts cells growth, but the major role in this respect could be attributed to arginine supplementation. Also arginine directly increased the expression of TGF-β mRNA and TGF-β protein levels as it could increase the production and deposition of matrix components **(Narita et al., 1995).** Furthermore the increase in BMD due to arginine supplementation could be explained as arginine has a dual effect on physiological regulators of bone remodeling. Arginine could potentially increase bone formation over bone resorption, and consequently, increase bone mass **(Van’t Hof and Ralston, 2001).** Additionally, there is growing evidence demonstrated that moderate concentrations of nitric oxide (NO) play an essential physiological role in promoting maintenance of bone density-stimulating new bone formation while suppressing bone catabolism **(Armour et al., 2001).**

Our data indicated that glutamine administration to ovariectomized rats tends to significantly increase 1,25(OH)2D3,IGF-1,TGF-β,BMC and BMD. While it could significantly decrease serum PTH level when compared to ovareiectomized rats administered lactose only.

It has been reported that glutamine supplementation produces glutathione which acts as a potent enhancer of calcium through activation of calcium sensing receptor (CaSR). Glutathione acts as an endogenous modulator of this receptor particularly in the parathyroid gland where this receptor is known to control parathyroid hormone release **(Wang et al., 2006).** Also glutathione could increase circulating 1,25 (OH)2D3 via stimulating enzymatic activity of the renal 1 alpha-hydroxylase of 25-hydroxycholecalciferol **(Schedl et al.,1992).** Furthermore glutamine supplementation increase IGF-1 level due to its conversion to alpha-ketoglutarate (αKG) in the body. Alpha -ketoglutarate has been shown to increase circulating plasma levels of insulin, growth hormone with consequent increase in IGF-1 **(Jeevanandam and Petersen, 1999)**. Growth hormone could stimulate osteoblastic proliferation and differentiation and increase the production of IGF-1 **(Corpas et al., 1993).**

The role of glutamine in inducing the detectable increase in BMD of right femur areas of ovariectomized rats mainly depends on its anabolic effect on bone, since glutamine played specific role in mechanisms associated with cellular proliferation and/or differentiation through particular receptors and transporters functionally expressed in rat calverial osteoblasts **(Yoneda and Hinoi, 2003).** Moreover, Alpha ketoglutrate has been found to increase mineralization, higher volumetric cortical bone density and increase trabecular bone density in animals **(Tatara et al., 2003).** Alsoalpha ketoglutrate is a component of antioxidant glutathione and polyglutamated folic acid. These antioxidants play a role in inhibiting osteoclastogensis via inhibition of reactive oxygen species (ROS) which are necessary for osteoclast activity and bone resorption **(Key et al., 1994)**. The positive effect of α-KG on bone was previously reported in birds since it could increase bone weight, mean relative wall thickness, maximum elastic strength, ultimate strength and volumetric bone mineral density in birds **(Tatara et al., 2005)** Similar findings have been also reported in ovariectomized rats **(Radzki et al., 2002).** Finally, the cyclization of glutamine produces proline, an amino acid important for synthesis of collagen and connective tissue **(Tapiero et al., 2002)** which contributes to the positive influence of glutamine on bone tissue.

The suggested mode of action of α-KG on bone mineralization could be attributed to the efficacy of α-KG to maintain a delicate balance between bone resorption and bone formation that plays an important role in determining bone strength and integrity **(Rodan and Mertin, 2000).** Glutamine may play a role as a signal mediator in mechanisms associated with chondral mineralization through the group III m glutamine receptor (mGluR) subtype functionally expressed by chondrocytes in cartilage **(Wang et al., 2005).** Thus, glutamine could produce the increase in BMC of each of proximal, mid and distal areas of ovariectomized rat right femur bone through its indirect effect on mechanical properties of bone.Lactose has a significant role in promoting the effect of glutamine on BMC of the three regions of right femur bone of OVX rats in the present study.

 The current results showed that taurine administration induced significant decrease in serum PTH level while it produced significant increase in each of 1,25(OH)2D3, IGF-1,TGF-β,BMC and BMD in ovariecomized rats. The effect of taurine in decreasing PTH level could be attributed to its role in increasing magnesium concentration through the activation of extracellular signal regulated protein kinase (ERK) pathway **(Jeon et al., 2007).** The resulting elevation in magnesium contcentration could suppress PTH secretion as magnesium positively affected intestinal calcium absorption and bone metabolism in ovariectomized rats **(Toba et al., 2000).** Taurine has been shown to have direct effect on accelerating vitamin D absorption and in turn increasing serum 1, 25(OH) 2 D3 level in ovariectomized rats treated with taurine. This suggestion is greatly supported by **Petrosian and Haroutounian, (2000). Gaylord et al. (2007)** stated that taurine stimulated pituitary growth hormone with subsequent stimulation of growth hormone –dependent IGF-1 in animals. Thus, growth hormone responsive and IGF-1 secreting cells might require sufficient taurine to secrete IGF-1 at normal levels **(Hu et al., 2000).** Taurine could produce the detectable increase in serum TGF-β level by two mechanisms, stimulatory action of taurine on osteoblastic differentiatation as well as bone matrix formation **(Park et al., 2001),** and indirect effect of taurine in increasing circulating level of 1,25 (OH)2 D3 which in turn led to increasing TGF-β release from bone cells. Lactose may have a role in enhancing TGF-β level in contribution with taurine through stimulation of osteoblast growth and production of growth factors mainly TGF-β **(Petrosian and Haroutounian, 2000).** Moreover taurine has been found to promote osteoblasts mineralization and it could regulate osteoblasts metabolism via stimulation of extracellular signal regulated protein kinase phosphorylation **(Park et al., 2001).** Taurine might also have antiresorptive action through its antioxidant effect **(Lourenco and Camilo, 2002).** Therefore, we could suggest that taurine via scavenging reactive oxygen species, necessary for osteoclast function, could inhibit bone resorption. Therefore, through these common pathways, taurine has a preventive effect on bone loss. The unique role of taurine in modulating mitochondrial Ca2+ homeostasis might be of particular importance under pathological conditions **(Palmi et al., 1999).** Lactose may have a synergistic effect with taurine on increasing BMC of proximal, mid and distal regions of right femur bone.

Histological investigation of bone tissue sections showed that estrogen deficiency is associated with elevated bone resorption caused by a rise of osteoclast number **(Marino, 2003).** Recent study of **Park et al. (2008)** observed that the ovariectomized rats that exhibited osteoporosis within 7 weeks after surgery showed large decreases in the bone volume ratio and trabecular bone thickness.

 Treatment with lactose showed some necrotic areas and appearance of small cavities in the bone. The ability of lactose to facilitate the passage of calcium across the intestine, resulting in improved calcium availability to the skeleton **(Miller et al., 1988).** **Marie and Travers (1983)** observed a slight decrease in osteocytes thickness in rats fed diet containing lactose.

Supplemntation of ovariectomized rats with arginine revealed the formation of new bone. This finding could be explained in the view of the effect of arginine via growth hormone stimulation as well as IGF-1 production. Growth hormone has been found to have a positive effect on chondrocytes and osteoblasts **(Saggese et al., 1995)** as well as it couldincrease the number and function of osteoblasts **(Bouillon, 1991).** This is because of osteoblasts express functional growth hormone receptors (GHR) **(Nilsson, et al., 1995)** indicating that growth hormone (GH) also exerts a direct effect on osteoblasts. A direct effect of GH on osteoblasts is supported by earlier results for the epiphysial growth plate, where it has been demonstrated that GH interacts directly with epiphysial chondrocytes for the regulation of longitudinal bone growth **(Ohlsson et al., 1992 a,b )**.IGF-1 has been found to play a role in trabecular and cortical bone formation **(Conconi et al., 2001)** IGF-1 showed a positive effect on bone formation in vitro as it could stimulate the formation of osteocalcin, collagen and non-collagenous matrix proteins by differentiated osteoblasts and increased the number of functional osteoblasts by promoting osteoprogenitor cell replication **(Visser and Hoekman 1994)**. Lactose assisted arginine in mainting plenty of calcium for strengthening the fragile bone, and increasing bone thickness as appeared in the current study.

 Supplementation of ovariectomized rats with glutamine showed the calcification of cartilage, increased trabecular bone thickness and formation of new bone. This result could be attributed to the functional role of glutamine in chondral mineralization through the group III mGluR subtype functionally expressed by chondrocytes in cartilage **(Wang et al., 2005).** Moreover, glutamine via producing α-KG in the body could promoting bone weight, wall thickness and bone strength **(Tatara et al., 2005).** Recent study of **Polat et al. (2007)** demonstrated that glutamine had positive effects on healing of traumatically fractured bone through attainment of positive nitrogen balance. The role of lactose here is to improve calcium availability to facilitate the formation of new bone.

 Taurine could restore the normal appearance of trabiculae as shown in the micrograph of bone tissue section of left femur of ovariectomized rats treated with taurine in the current work. Considering that a significant amount of taurine is transported to bone tissues, the transcription and translation of taurine occurs in bone forming cells **(Yuan et al.,** **2006)**. Therefore, it is reasonable to propose that taurine may play an important role in bone metabolism. Taurine in the osteoblasts activates extracellular signal regulated protein kinase-2 (ERK2) and phosphorylates transcription factors thus activating collagen gene transcription and protein synthesis. These actions of taurine may be beneficial for osteoblastic differentiation and bone matrix formation (**Park et al., 2001**). Additionally, taurine could stabilize cell membranes, eliminate oxide free radicals, regulate intracellular osmosis and maintain intracellular calcium concentration (**Pasantes-Morales et al., 1998**). It is clear that lactose could enhance the effect of taurine on bone formation through improving the organization of the trabecular bone in ovariectomized rats as shown in the present study from the normal appearance of bone following administration of taurine in lactose.

In conclusion, the present study provided clear evidence that ovariectomy produced marked abnormalities in bone biomarkers and in increasing risk of fracture. Also ovariectomy reduced plasma IGF-1 level and decreased TGF-β, as well as accelerated bone loss phase. The selected amino acids showed positive effect on bone via inhibiting the secretion of parathyroid hormone in concomitant with increasing serum 1,25 dihydroxyvitamin D3 Also, the studied amino acids stimulated the production of IGF-1 and TGF-β and increased bone calcification rate as well as improved calcium availability to the skeleton. Lactose participated in enhancing the positive effect of the selected amino acids on bone. Arginine provided promising effect on bone through stimulation of insulin- like growth factor. Also, arginine via nitric oxide which is involved in increasing basal calcium absorption in small intestine thus stimulating the replication of primary osteoblasts and as well as inhibiting osteoclasting bone resorption. Glutamine through the production of alpha-ketoglutarate and glutathione showed potent effect on bone. These metabolic products of glutamine have a critical role in increasing bone density and strength in addition to bone mineralization. Taurine revealed the most effective action on bone remodeling via stimulating osteoblastic differentiation as well as bone matrix formation. In addition to taurine anabolic effect, it has an antiresorptive action through its antioxidant activity which participates in inhibiting osteoclast function and consequently bone resorption. Our finding might be useful for the future strategies against menopausal bone turnover and implicitly osteoporosis progression.

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**References:**

1. **Ahmad, A.M.; Thomas, J.; Clewes, A.; Hopkins, M.T.; Guzder, R.; Ibrahim, H.; Durham, P.H.; Vora, J.P. and Fraser, W.D. (2003):** Effects of Growth Hormone Replacement on Parathyroid Hormone Sensitivity and Bone Mineral Metabolism. J. Clin. Endocrinol. Metab., 88: 2860– 2868.
2. **Ammann, P.; Bonjour, J.P. and Rizzoli, R. (2000)**: Essential amino acid supplements increase muscle weight, bone mass and bone strength in adult osteoporotic rats J. Musculoskeletal Neuronal Interact., 1: 43 -44.
3. **Ann, J.Y.; Kim, S.J.; Han, S.P.; Kim, J.W.; Kim, H.J.; Do, J.H.; Kim, J.G.; Chang, S.K. and Jeon, W.K. (2004):** Effect of glutamine on the non-steroidal anti-inflammatory drug –induced bacterial translocation. Korean J Gastroenterol. 44: 252-258.
4. **Armitage, P. and Berry, G. (1987):** Comparison of several groups. In: statistical method in medical research 2nd Ed. Blockwell significant publication, Oxford. pp.186-213.
5. **Armour, K.E.; Armour, K.J.; Gallagher, M.E.; Gödecke, A.; Helfrich, M.H.; Reid, D.M.; and Ralston, S.H. (2001)**: Defective bone formation and anabolic response to exogenous nitric oxide synthase. Endocrinology, 142: 760-766.
6. **Ash, S.L. and Goldin, B.R. (1988):** Effects of age and estrogen on renal vitamin D metabolism in the female rat. American Journal of Clinical Nutrition, 47: 694-699.
7. **Baecker, N.; Boese, A.; Schoenau, E.; Gerzer, R. and Heer, M. (2005):**L-Arginine, the Natural Precursor of NO, Is Not Effective for Preventing Bone Loss in Postmenopausal Women. Journal of Bone and Mineral Research, 20:471-479.
8. **Blum, WF.; Albersson-Wikland, K.; Rosberg, S. and Ranke MB. (1993):** Serum levels of insulin-like growth factor-1 (IGF-1) and IGF binding protein 3 reflect spontaneous growth hormone secretion. J. Clinical Endocrinol. Metab., 76:610-616.
9. **Bouillon, R. (1991):** Growth hormone and bone. Horm. Res., 36: 49-55.
10. **Calo L.; Castringnano R.; Davis P.A.; Carraro G.; Pagnnini S.; Semplicini A. and D'angelo A. (2000):** Role of insulin-like growth factor –I in primary osteoporosis: a correlative study. J Endocrinol. Invest., 23: 223-227.
11. [**Chen, C.C**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Chen%20CC%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**.;** [**Liu, M.H**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Liu%20MH%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**.;** [**Wang, M.F**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Wang%20MF%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**. and** [**Chen, C.C**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Chen%20CC%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**. (2007):** Effects of aging and dietary antler supplementation on the calcium-regulating hormones and bone status in ovariectomized SAMP8 mice. Chin J Physiol., 50: 308-314
12. **Centiner, M.; Sener, G.;, Sehirli, A.O.; Eksioglu –Demiralp, E.; Ercan, F.; Sirvanci, S.; Gedik, N.; Akpulat, S.; Tecimer, T. and Yegen, B.C. (2005):** Taurine protects against methotrexate- induced toxicity and inhibits leukocyte death . Toxicol Appl&Pharmacol., 209: 39-50.
13. [**Chevalley, T**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Chevalley%20T%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**.;** [**Rizzoli, R**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Rizzoli%20R%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**.;** [**Manen, D**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Manen%20D%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**.;** [**Caverzasio, J**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Caverzasio%20J%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**. and** [**Bonjour, J.P**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Bonjour%20JP%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**. (1998):** Arginine increases insulin-like growth factor-I production and collagen synthesis in osteoblast-like cells**.** Bone, 23: 103-109.
14. **Clementi, G.; Fiore, C.E.; Mangano, N.G.; Cutuli, V.M.; Pennisi, P.; Caruso, A.; Prato, A.; Matera, M. and Amico-Roxas, M. (2001):** Role of soy diet and L-arginine in cyclosporin-A-induced osteopenia in rats. Pharmacol Toxicol., 88: 16-19.
15. **Conconi, MT.; Tommasini, S.; Muratori, E. and Parnigotto, PP. (2001):** Essential amino acids increase the growth and alkaline phosphatase activity in osteoblasts cultured in vitro. Farmaco., 56:755-761.
16. [**Corpas, E**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Corpas%20E%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**.;** [**Harman, S.M**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Harman%20SM%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**. and** [**Blackman, M.R**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Blackman%20MR%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**.(1993):** Human growth hormone and human aging. Endocr. Rev., 14: 20-39.
17. **Drury, R.A.B. and Wallington, E.A. (1980):** Preparation and fixation of tissues in Carlton’s histological techniques 4th edition, Oxford University Press-London. 36-56
18. **Fan, U.; Liu, J.; Wang, S.; Wang, H.; Shi, F.; Xiong, L.; He, W. and Peng, X. (2005):** Functional proteome of bones in rats with osteoporosis following ovariectomy. Life Sciences, 76: 2893-2901.
19. **Fini, M.; Giardino, R.; NicoliAldini, N.; Martini, L.; Rocca, M. Bertoni, F.; Capelli, S.; Cantelli forti, G.; Sapone, A.; Rossetti, A.; Morrone, G. and Giavaresi, G. (1996):** Role of Lactose, arginine and lysine combination in fracture heling (an experimental study). Ann. Ital Chir., 67: 77-82
20. **Finkelman, R.; Bell, N.H.; Strongt, D.D.; Demers, L.M. and Baylink, D.J. (1992):** Ovariectomy selectively reduces the concentration of transforming growth factor p8 in rat bone: Implications for estrogen deficiency-associated bone loss (postmenopaUsal osteoporosis/insulin-lke growth factor I/insulin-like growth factor H). Proc. Nati. Acad. Sci. USA., 89: 12190-12193
21. **Gaumet, N**.; **Seibel, M.J.; Braillon, P.; Giry, J.; Lebecque, P.; Davicco, M.J.; Coxam, V.; Rouffet, J.; Delmas, P.D. and Barlet, J.P. (1996):** Influence of ovariectomy on bone metabolism in very old rats. Calcif. Tissue Int., 58: 256-262.
22. **Gaylord T.G.; Barrows F.T.; Teague A.M.; Johansen KA.; Overtrf K.E. and Shepherd B. (2007):** Supplementation of taurine and methionine to all-plant protein diets for rainbow trout (Oncorhynchus mykiss). Aquaculture, 269: 514-524.
23. **Gupta, V.; Gupta, A.; Saggu, S.; Divekar, H.M.; Grover, S.K. and Kumar, R. (2005):** Anti–stress and Adaptogenic Activity of L-Arginine supplementation . Evid Based Complement Alternat Med. 2: 93-97.
24. **Hollis, B.W. (1986):** “Assay of Circulating 1,25-dihydroxyvitamin D Involving a Novel Single-Cartridge Extraction and Purification Procedure,” Clinical Chemistry,32 (11): 2060.
25. [**Hu, J.M**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Hu%20JM%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**.;** [**Rho, J.Y**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Rho%20JY%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**.;** [**Suzuki, M**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Suzuki%20M%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**.;** [**Nishihara, M**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Nishihara%20M%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**. and** [**Takahashi, M**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Takahashi%20M%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**. (2000):** Effect of taurine in rat milk on the growth of offspring. J. Vet. Med. Sci., 62: 693-698.
26. [**Jeevanandam, M**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Jeevanandam%20M%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**. and** [**Petersen, S.R**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Petersen%20SR%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**. (1999):** Substrate fuel kinetics in enterally fed trauma patients supplemented with ornithine alpha ketoglutarate. Clin Nutr. 18: 209-217.
27. **Jeon, S.H.; Lee, M.Y.; Kim, S.J.; Joe, S.G.; Kim, G.B.; Kim, I.S.; Kim, N.S.; Hong, C.U.; Kim, S.Z.; Kim, J.S. and Kang, H.S. (2007):** Taurine increases cell proliferation and generates an increase in [Mg2+]i accompanied by ERK 1/2 activation in human osteoblast cells. FEBS Letters, 581: 5929-5934.
28. **Kajdaniuk D.; Marek B.; Swietochowska E.; Buntner B. and Matuszawska G. (1999):** Insulin-like growth factor –I: pathological and clinical aspects. Pol Merk Lek., 41: 205-207.
29. **Kassem, M.; Brixen, K. and Mosekilde, L. (2003):** Ageing of the human skeleton and its contribution to osteoporotic fractures. In: Aspinall R, editor. Ageing of organs and systems. Dordreeht: Kluwer Academic.
30. **Katherine M. McLeod,3 Susan E. McCann,4 Peter J. Horvath,3 and Jean Wactawski-Wend** **(2007)** Predictors of Change in Calcium Intake in Postmenopausal Women after Osteoporosis Screening1,2 J. Nutrition. 137: 1968–1973.
31. **Key, L.L.; Wolf, Jr.; Gundberg, W.C. and Ries, W.L. (1994):** Superoxide and bone resorption. Bone, 15: 431–436.
32. **Kevin D. Cashman** **(2007)**: Diet, nutrition and bone health. J.Nutrition,137:2507s-2512s
33. **Kim, SJ.; Romeo, D.; Yoo, YD. and Park, k. (1994):** Transforming growth factor-beta: expression in normal and pathological conditions. Horm. Res. 42:5-8.
34. **Kimble, R.B.; Srivastava, S.; Ross, F.P.; Matayoshi, A. and Pacifici, R. (1996):** Estrogen deficiency increases the ability of stromal cells to support murine osteoclastogenesis via an interleukin-1 and tumor necrosis factor-mediated stimulation of macrophage colony-stimulating factor production. J Biol Chem., 271: 28890-28897.
35. **Kirk, S.J.; Hurson, M. and Regan, M.C. (1994):** Arginine stimulates wound healing and immune function in elderly human beings JPEN, 18: 559-560.
36. **Kitazawa, R.; Kimble, R.B.; Vannice, J.L.; Kung, V.T. and Pacifici, R. (1994):** Interleukin-1 receptor antagonist and tumor necrosis factor binding protein decrease osteoclast formation and bone resorption in ovariectomized mice. J Clin Invest., 94: 2397-2406.
37. **Koide, M.; Okahashi, N.; Tanaka, R.; Kazuno, K.; Shibasaki, K.; Yamazaki, Y.; Kaneko, K.; Ueda,N.; Ohguchi, M.; Ishihara, Y.; Noguchi, T and Nishihara, T. (1999):** Inhibition of experimental bone resorption and osteoclast formation and survival by 2-aminoethanesulphonic acid. Arch. Oral Biol., 44: 711-719.
38. **Kwan Tat, S.; Padrines, M.; Theoleyre, S.; Heymann, D. and Fortun, Y. (2004):** IL-6, RANKL, TNF- alpha/IL-1: interrelations in bone resorption pathophysiology. Cytokine Growth Factor Rev., 15: 49-60.
39. **Lourenco, R. and Camilo, M.E. (2002):** Taurine: a conditionally essential amino acid in humans? An overview in health and disease. Nutr. Hosp. 17: 262-270.
40. **Matsushita, H.;** **Barrios, J.A.; Shea, J.E. and Miller, S.C. (2008):** Dietary fish oil results in a greater bone mass and bone formation indices in aged ovariectomized rats. J. Bone Miner Metab., 26: 241-247.
41. **Marie, P.J. and Travers, R. (1983):** Effects of magnesium and lactose supplementation on bone metabolism in the X-linked hypophosphatemic mouse. Metabolism., 32: 165-171.
42. [**Mastaglia, S.R**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Mastaglia%20SR%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**.;** [**Pellegrini, G.G**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Pellegrini%20GG%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**.;** [**Mandalunis, P.M**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Mandalunis%20PM%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**.;** [**Gonzales Chaves, M.M**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Gonzales%20Chaves%20MM%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**.;** [**Friedman, S.M**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Friedman%20SM%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**. and** [**Zeni, S.N**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Zeni%20SN%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**. (2006):** Vitamin D insufficiency reduces the protective effect of bisphosphonate on ovariectomy-induced bone loss in rats. Bone, 39: 837-44.
43. **Miller, S.C.; Miller, M.A. and Omura, T.H. (1988):** Dietary lactose improves endochondral growth and bone development and mineralization in rats fed a vitamin D-deficient diet. Journal of Nutrition, 118: 72-77.
44. **Narita, I.; Border, W.A.; Ketteler, M.; Ruoslathi, E. and Noble, N.A. (1995):** L-Arginine may mediate the therapeutic effects of low protein diets. Proc. Natl. Acad. Sci. USA 92: 4552– 4556.
45. **Nilsson, A.; Swolin, D.; EnerbEck, S. and Ohlsson, C. (1995)**: Expression of functional growth hormone receptors in cultured human osteoblast-like cells. J. Clin. Endocrinol. Metab., 80: 3483-3488.
46. **Ohlsson, C.; Nilsson, A.; Isaksson, O.G.P. and Lindahl, A. (1992)**a : Effect of growth hormone and insulin-like growth factor-l on DNA synthesis and matrix production in rat epiphyseal chondrocytes in monolayer culture. J. Endocrinol.133: 291-300.
47. [**Palmi, M**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Palmi%20M%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**.;** [**Youmbi, G.T**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Youmbi%20GT%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**.;** [**Fusi, F**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Fusi%20F%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**.;** [**Sgaragli, G.P**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Sgaragli%20GP%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**.;** [**Dixon, H.B**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Dixon%20HB%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**.;** [**Frosini, M**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Frosini%20M%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**. and** [**Tipton, K.F**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Tipton%20KF%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**. (1999):** Potentiation of mitochondrial Ca2+ sequestration by taurine. Biochem. Pharmacol., 58: 1123-1131.
48. **Park, J.A.; Ha, S.K.; Kang, T.H.; Oh, M.S.; Cho, M.H.; Lee, S.Y.; Park, J.H. and Kim, S.Y. (2008):** Protective effect of apigenin on ovariectomy-induced bone loss in rats. Life Sciences, 82: 1217-1223.
49. [**Park, S**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Park%20S%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**.;** [**Kim, H**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Kim%20H%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**. and** [**Kim, S.J**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Kim%20SJ%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**. (2001):** Stimulation of ERK2 by taurine with enhanced alkaline phosphatase activity and collagen synthesis in osteoblast-like UMR-106 cells. Biochem Pharmacol. 62: 1107-1111.
50. **Pasantes-Morales, H.; Quesada, O. and Moran, J. (1998):** Taurine: An osmolyte in mammalian tissues. Adv. Exp. Med. Biol., 442: 209-217.
51. **Passeri, G.; Girasole, G.; Jilka, R.L. and Manolagas, S.C. (1993):** Increased interleukin-6 production by murine bone marrow and bone cells after estrogen withdrawal. Endocrinology, 133: 822-828.
52. **Petrosian, A.M. And Haroutounian, J.E. (2000):** Taurine as a universal carrier of lipid soluble vitamins: a hypothesis. Amino Acids, 19: 409-421.
53. **Polat, O.; Kilicoglu, S.S. and Erdemli, E. (2007):** A controlled trial of glutamine effects on bone healing. Adv. Ther., 24: 154-160.
54. **Preisinger E (2009)** Physiotherapy and exercise in osteoporosis and its complications. Z Rheumatol. Sep;68(7):534-6,538.
55. **Radzki, R.P.; Bienko, M.; Puzio, I.; Filip, R.; Pierzynowsk, S.G. and Studzinsk, T.** **(2002)**: The influence of alpha-ketoglutarate (AKG) on mineralization of femur in rats with established osteopenia. Acta. Orthop. Scand., 73(Suppl. 304): 52.
56. **Rodan, G.A. and Martin, T.J.** **(2000)**. Therapeutic approaches to bone diseases. Science, 289: 1508–1514.
57. **Ross, F.P. (2003):** Interleukin 7 and estrogen-induced bone loss. Trends Endocrinol Metab., 14: 147-149.
58. **Saggese, G.; Baroncelli, G.I.; Federico, G. and Bertelloni, S. (1995):** Effects of growth hormone on phosphocalcium homeostasis and bone metabolism. Horm Res., 44: 55-63.
59. **Schedl, H.P.; Wilson, H.D.; Horst, R.L.; Christensen, K and Brown, K. (1992):** Glutathione effects on vitamin D metabolism in control and streptozotocin diabetic rat. Nutr-Res. Tarrytown, N.Y. : Pergamon press., 12: 541-1547.
60. **Schermer, S. (1967):** The Blood Morphology of Laboratory Animals, 3rd ed., F.A. Davi, Philadelphia p.42.
61. **Segal, E.; Dvorkin, L.; Lavy, A.; Geila S.; Rozen, R.D.; Yaniv, I.; Raz, B.; Tamir, A. and Ish-Shalom, S. (2003):** Bone Density in Axial and Appendicular Skeleton in Patients with Lactose Intolerance: Influence of Calcium Intake and Vitamin D Status. Journal of the American College of Nutrition, 22: 201-207.
62. **Shen, V.; Birchman, R.; Wu, D.D. and Lindsay, R. (2000):** Skeletal effects of parathyroid hormone infusion in ovariectomized rats with or without estrogen repletion. J. Bone Miner. Res., 15: 740-746.
63. **Shortt, C. and Flynn, A. (1991):** Effect of dietary lactose on salt-mediated changes in mineral metabolism and bone composition in the rat British Journal of Nutrition, 66: 73-81.
64. [**Silver, J**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Silver%20J%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_RVAbstractPlus)**.;** [**Yalcindag, C**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Yalcindag%20C%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_RVAbstractPlus)**.;** [**Sela-Brown, A**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Sela-Brown%20A%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_RVAbstractPlus)**.;** [**Kilav, R**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Kilav%20R%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_RVAbstractPlus)**. and** [**Naveh-Many T**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Naveh-Many%20T%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_RVAbstractPlus)**. (1999):** Regulation of the parathyroid hormone gene by vitamin D, calcium and phosphate. Kidney Int Suppl., 73: 2-7.
65. **Tapiero, H.; Mathe, G.; Couvreur, P. and Tew, K.D. (2002):** Glutamine and glutamate. Biomed Pharmacother., 56: 446-457.
66. [**Tapiero, H**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Tapiero%20H%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**.;** [**Mathé, G**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Math%C3%A9%20G%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**.;** [**Couvreur, P**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Couvreur%20P%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**. and** [**Tew, K.D**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Tew%20KD%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**. (2002):** II. Glutamine and glutamate. Biomed Pharmacother., 56(9): 446-457.
67. **Tatara, M.**R.; **Brodzki, A; Krupski, W.; Sliwa, E.; Silmanowicz, P.; Majcher, P.; Pierzynowski, S.G. and Studzinski, T. (2005):** Effects of α-ketoglutarate on bone homeosatasis and plasma amino acids in turkeys. Poultry Science, 84: 1604-1609.
68. **Toba, Y.; Kajita, Y.; Masuyama, R.; Takada, Y.; Suzuki, K. and Aoe, S.** **(2000):** Dietary Magnesium Supplementation Affects Bone Metabolism and Dynamic Strength of Bone in Ovariectomized Rats**.** Journal of Nutrition., 130: 216-220.
69. **Van't Hoff, R.J. and Ralsion, S.H. (2001):** Nitric oxide and bone. Immunology, 103: 255-261.
70. **Visser, J.J. and Hockman, K. (1994):** [Arginine supplementation in the prevention and treatment of osteoporosis.](http://www.ncbi.nlm.nih.gov/pubmed/7877530?ordinalpos=49&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DefaultReportPanel.Pubmed_RVDocSum) Med Hypotheses.;43:339-42.
71. [**Wang, M.; Yao, Y.; Kuang, D. and Hampson, D.R.**](http://www.ncbi.nlm.nih.gov/pubmed/16455645?ordinalpos=2&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_RVDocSum) **(2006):** Activation of family C G-protein-coupled receptors by the tripeptide glutathione. J. Biol. Chem. 281: 8864-8870.
72. **Westerlind, K.C.; Wronski, T.J.; Evans, G.L. and Turner, R.T. (1994):** The Effect of Long-Term Ovarian Hormone Deficiency on Transforming Growth Factor β and Bone Matrix Protein mRNA Expression in Rat Femora.[Biochemical and Biophysical Research Communications](http://www.sciencedirect.com/science/journal/0006291X), 200: 283-289.
73. [**Yasutomi, C**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Yasutomi%20C%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**.;.;** [**Nakamuta, H**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Nakamuta%20H%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**.;** [**Fujita, T**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Fujita%20T%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**.;** [**Takenaga, T**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Takenaga%20T%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**. and** [**Koida, M**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Koida%20M%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**. (2002):** Anti-osteopenic effect of taurine: possible involvement of activated MEK-ERK-Cbfa1 signaling Nippon Yakurigaku Zasshi., 120: 114-115.
74. [**Yoneda, Y**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Yoneda%20Y%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**. and** [**Hinoi, E**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Hinoi%20E%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**. (2003):** Functional expression of machineries for glutamate signaling in bone. Nippon Yakurigaku Zasshi., 122: 14-17
75. [**Yoneda, Y**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Yoneda%20Y%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**. and** [**Hinoi, E**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Hinoi%20E%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**. (2003):** Functional expression of machineries for glutamate signaling in bone. Nippon Yakurigaku Zasshi., 122: 14-17.
76. [**Yuan, L.Q**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Yuan%20LQ%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**.;** [**Xie, H**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Xie%20H%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**.;** [**Luo, X.H**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Luo%20XH%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**.;** [**Wu, X.P**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Wu%20XP%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**.;** [**Zhou, H.D**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Zhou%20HD%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**.;** [**Lu ,Y**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Lu%20Y%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**. and** [**Liao, E.Y**](http://www.ncbi.nlm.nih.gov/sites/entrez?Db=pubmed&Cmd=Search&Term=%22Liao%20EY%22%5BAuthor%5D&itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_DiscoveryPanel.Pubmed_RVAbstractPlus)**. (2006):** Taurine transporter is expressed in osteoblasts. Amino Acids, 31: 157-163.