

Antifungal Activity And Flavonoid Composition Of *Wiesnerella denudata* steph

¹Rachana Mishra* and D. L. Verma²,
^{1&2}Department of Chemistry, Kumaun University,
SSJ Campus, Almora-263601, (Uttarakhand) India.

*Email- 09411102476m@gmail.com

Phone numbers: +91-9411102476¹, +91-5962-233849(R)²

Abstract: *Wiesnerella denudata* Steph, a liverwort of family Marchantiaceae and is an endemic species of bryoflora to Indian Himalayan Region. Aqueous extract of the liverwort has been used as a medicine to cure cough, bronchitis and asthma by the local inhabitants of Kumaun Hills. The Botanically identified liverwort, specimen No. 3 extracted with 50% H₂O-MeOH by cold percolation method. The concentrated H₂O layer of the extract was partitioned with CH₂Cl₂. 50% HOAc fractionation of CH₂Cl₂ soluble on cellulose CC gave a dark fluorescent band and it was eluted and collected separately. An eluate of the fraction was evaporated to dryness and it gave antifungal test against the conidial suspension of *Aspergillus flavus* and *Aspergillus niger* by the standard method of thin layer autobiography. An antifungal active fraction afforded four flavone glycosides, apigenin-7-O-β-D-glucoside, acacetin-7-O-α-L-rhamnoside, luteolin-3-O-glucuronide and luteolin-5-O-β-D-glucopyranoside. [Academia Arena, 2009;1(6):42-45]. (ISSN 1553-992X)

Keywords: *Wiesnerella denudata* Steph, medicine, cough, bronchitis, asthma

Introduction

Wiesnerella denudata Steph, a liverwort of family Marchantiaceae and an endemic to Indian Himalayan Region (IHR), has widely been used as a medicine to cure cough, bronchitis and asthma (Tewari, 1984). *Marchantia*, a prominent flavonoid producing genus of family Marchantiaceae, is characterized by the presence of glucosides and glucuronides of apigenin, luteolin, acacetin, chrysoeriol and genkwanin. The dominant flavonoids have been reported from the genus *Marchantia* are apigenin-4'-7-di-O-glucuronide, apigenin-7-O-glucoside, apigenin-7-O-β-D-glucuronide, apigenin-4'-O-glucuronide, chrysoeriol-7-O-β-D-glucuronide and luteolin-6-8-di-C-glucoside (Markham and Porter, 1974; 1975 and 1978). The flavonoids isolated from various members of genus *Marchantia* have widely been employed to chemotaxonomic and phylogenetic studies (Markham *et al.*, 1977, Campbell *et al.*, 1979, Schier, 1974). There are few studies reported by Verma and his co-workers, which are for medicinal plants and have widely been referred by various workers of medicinal chemistry (Khetwal and Verma, 1983, 1984, 1986, 1990; Khetwal *et al.*, 1985, 1986, Mishra, 2008, Mishra and Verma, 2009a, 2009b and 2009c).

Documenting the flavonoid composition from various species of Marchantiaceae to chemotaxonomic and phylogenetic studies, the genus *Wiesnerella* has been separated from the family Marchantiaceae and new family Wiesnerellaceae has been created for the members of genus *Wiesnerella* (Campbell *et al.*,

1979, Schier, 1974). The flavonoids, luteolin-5-O-glucuronide, apigenin-7-O-rhamnoside and acacetin-7-O-rhamnoside have previously been isolated from *Wiesnerella denudata* (Campbell *et al.*, 1979). The terpenoids, germacranolides and guainolides have also been isolated from *Wiesnerella denudata* (Asakawal, 1982). Present communication reveals the presence of flavone glycosides and antifungal activity from CH₂Cl₂ soluble of *Wiesnerella denudata*. This is first report on chemical constituents of *Wiesnerella denudata* native to India Himalayan region.

Material and method

1. Plant material and authentication:

Wiesnerella denudata Steph was collected from Dhobi ghat of Naini Tal at the altitude 2100m. It was identified by Prof. K. R. Verma, Department of Botany, Kumaun University, S. S. J. Campus, Almora (Uttarakhand). Its vouch. Specimen no. 3 has been deposited in the plant taxonomy laboratory of Botany Department of Kumaun University at Almora Campus, Uttarakhand, India.

2. Extraction and isolation of flavonoid positive fraction:

About 400gm air dried and powdered sample of *Wiesnerella denudata* was extracted with aqueous methanol (50:50) by cold percolation method for six days. The extract was filtered and concentrated under reduced pressure in Rota evaporator at 60^o C. The residue was partitioned between CH₂Cl₂:H₂O. The CH₂Cl₂ soluble was evaporated to dryness. It was chromatographed on cellulose (Merck CC) and eluted with 50% HOAc. A single broad dark purple fluorescent band was observed on CC and it was eluted and collected separately by

monitoring with UV light. An elute of the fraction was evaporated to dryness and residue was dissolved in MeOH. It was examined on 2DPC using t-BAW (3:1:1) and 30% HOAc as a developing solvent. A total of seven spots were discernible on 2DPC after spraying with aqueous solution of FeCl_3 and $\text{K}_4\text{Fe}(\text{CN})_6$ (1:1). Out of seven spots four were identified as flavonoids after spraying PC with NH_3 , ZrOCl_2 and NA (Naturstoffreagenz, A) reagents (Mabry *et al.*, 1970).

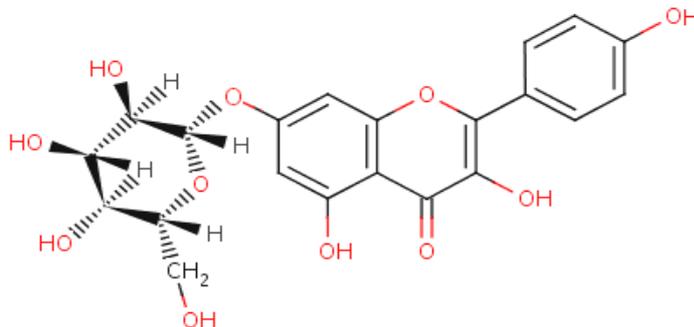
3. Screening of antifungal activity: The flavonoid positive fraction, an eluate of dark purple fluorescent band observed on CC after fractionation of CH_2Cl_2 soluble with 50% HOAc, was screened for antifungal activity by the standard method of thin layer autography using SiO_2 an absorbent and conidial suspension of *Aspergillus flavus* and *Aspergillus niger* in sugar salt medium as a spraying reagent (Homans and Fush, 1970; Pero and Owens, 1971). The TLC plate was developed with CH_2Cl_2 : MeOH (90:10). The dried and developed plate was sprayed with conidial suspension of *Aspergillus flavus* in sugar salt medium. The plate was incubated at 27°C for three days. Two zones of inhibition were observed on TLC.

4. Isolation of flavonoids from antifungal active fraction:

The flavonoid positive fraction, a bioassay guided antifungal active, was dissolved in MeOH. It was banded on Whatmann No. 3 PC (8 sheets) and repeatedly developed in BAW (BuOH : AcOH : H_2O , 4:1:5, upper layer). After three times repeated development, four fluorescent bands were observed on PC at Rf 65, 60, 58 and 55 which representing Frac-I, Frac-II, Frac-III and Frac-IV respectively. Each band was cut and eluted separately by monitoring with UV light. Four compounds A, B, C and D were isolated from Frac-I, Frac-II, Frac-III and Frac-IV respectively. An eluate of each fraction was finally purified on Sephadex LH-20 using 50% aqueous MeOH as an eluent.

Result and discussion

Compound [A], a dark purple fluorescent on paper chromatogram under UV light, gave positive tests for HCl , FeCl_3 and α -naphthol. Complete acid hydrolysis of compound [A] gave apigenin (CoPC) and glucose. UV spectrum gave MeOH (270, 341); AlCl_3 (277, 300, 386); NaOMe (267, 389); $\text{NaOAc}+\text{H}_3\text{BO}_3$ (268, 339) and NaOAc (272, 350). It was identified as apigenin-7-O-glucoside.



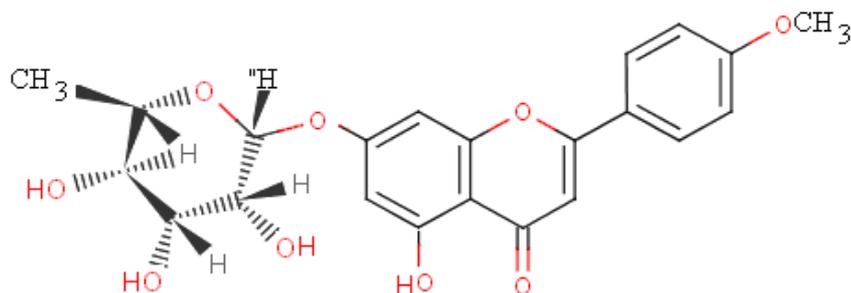
Compound [A]

Compound [B], a dark purple fluorescent on paper chromatogram under UV light, gave positive tests for FeCl_3 , $\text{Mg}+\text{HCl}$ and α -naphthol. Complete acid hydrolysis of the compound gave acacetin (CoPC) and rhamnose. FABMS (-ve) of the compound B gave a molecular ion at m/z 429

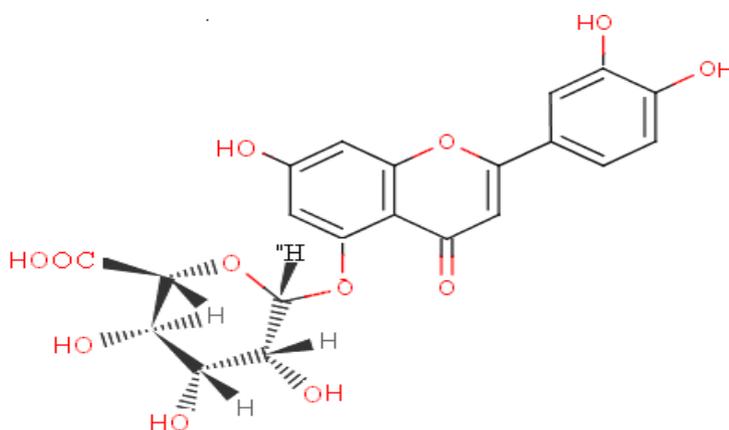
and other prominent ions observed at m/z 283 [m/z 429-rham] $^-$ indicating release of one mole of rhamnose from apigenin-4'- OCH_3 . It was identified as acacetin-7-O-rhamnoside on the basis of ^1H NMR studies (see, table no. 1).

Table [1]: ^1H NMR of compound [B] in DMSO-d_6 (400MHz)

Shift	Multiplicity	H-attributed
6.87	1H(s)	H-3
6.35	1H, d, J=2.0Hz	H-6
6.75	1H, d, J=2.0Hz	H-8
7.12	2H, d, J= 8.8Hz	H-3'/5'
8.05	2H, d, J= 8.8Hz	H-2'/6'
5.70	1H, d, J=1.2Hz	H-1''
3.10-4.10	(m)	Remaining protons of rhamnose
1.20	3H,d,J=6.0Hz	$-\text{OCH}_3$

**Compound [B]**

Compound [C] is a dark purple fluorescent on paper chromatogram under UV light, gave positive test for FeCl_3 , $\text{Mg}+\text{HCl}$ and α -naphthol. Normal acid hydrolysis with 1.5N HCl of the compound afforded luteolin (CoPC) and glucuronic acid (CoPC). The position of attachment of glucuronic acid to luteolin was assigned on the basis of ^1H NMR studies (see, table no. 2). The compound [C] was identified as luteolin-3-O-glucuronide.

**Compound [C]****Table [2]: ^1H NMR of compound [C] in DMSO-d_6 (400MHz)**

Shift	Multiplicity	H-attributed
6.20	1H, d, $J=2.0\text{Hz}$	H-6
6.53	1H, d, $J=2.0\text{Hz}$	H-8
6.81	1H(s)	H-3
6.98	1H, d, $J=8.0\text{Hz}$	H-5'
7.65	1H, dd, $J=2.0, 8.0\text{Hz}$	H-6'
7.80	1H, d, $J=2.0\text{Hz}$	H-2'
5.10	1H, d, $J=7.5\text{Hz}$	H-1''
3.10-4.10	(m)	Remaining proton of glucuronic acid

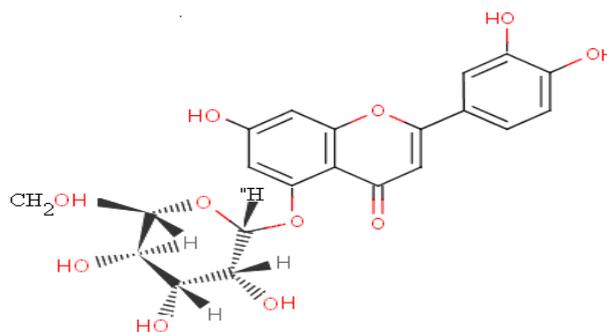
Compound [D] is a blue fluorescent on paper chromatogram under UV light and a slower moving component on PC in BAW. FABMS(-ve) of the compound gave a molecular ion at m/z 447 $[\text{M}-\text{H}]^-$ and prominent ion at m/z 285 [m/z 447-glucose] indicating the release of glucose moiety from luteolin. Complete acid hydrolysis of compound [D] with 2NHCl supported the presence luteolin (CoPC) and glucose (CoPC).

The compound [D] appeared as a blue fluorescent on PC under UV light with and without the presence of NH_3 vapours indicating the release of glucose moiety from 5 position.

It has further been supported by ^1H NMR studies (in DMSO-d_6 and 400MHz), see table no. 3. The compound [D] was identified as luteolin-5-O- β -D-glucopyranoside.

Table [3]: ¹HNMR of compound [D] in DMSO-d₆ (400MHz)

Shift	Multiplicity	H-attributed
6.55	1H(s)	H-3
6.69	1H, d, J=2.0Hz	H-6
6.79	1H, d, J=2.0Hz	H-8
6.87	1H, d, J= 8.8Hz	H-5
7.35	1H, d, J= 2.0Hz	H-2'
7.37	1H, dd, J=2.0, 8.8Hz	H-6'
5.10	1H,d,J=7.5Hz	H-1''
3.0-4.0	(m)	Remaining protons of glucose

**Compound [D]**

Acknowledgements: We thank to the authority of Central Drug Research Institute (CDRI), Lucknow (U. P.), India for their kind co-operation in the structural analysis of flavonoids by ¹HNMR, UV and MS spectral studies.

Address for correspondence: ¹Mrs. (Dr.) Rachana Mishra; Department of Chemistry, SSJ Campus, Almora-263 601, Uttarakhand (India).

²Dr. D. L. Verma, Associate Professor of Chemistry, Department of Chemistry, SSJ Campus, Almora-263 601, Uttarakhand (India).

References

- Asakawa, Y. (1982) Chemical constituents of Hepaticae in W. Hertz H., Greisebaets and G. W. Kirby (eds.) Progress in Chemistry of Natural Products, **42**, 1-285, Springer.
- Campbell, E. O., Markham, K. R., Moore, N. A. Porter, L. J. and Wallace, J. W. (1979) Taxonomic and phylogenetic implications of the comparative flavonoid chemistry of species in the family Marchantiaceae, J. Hattori Bot Lab, **45**, 185-199.
- Homans, A. L. and Fuschs, A., (1970) J. Chromatography, **51**, 327.
- Khetwal KS, and Verma DL. Natural and Applied Science Bulletin. 1983; **34(4)**: 337-338.
- Khetwal KS and Verma DL. Indian J. of Pharmaceutical Sciences. 1984; **46(1)**: 25-26.
- Khetwal KS, Verma DL and Tandon AK. Indian Drugs. 1986; **24**: 116-117.
- Khetwal KS, Verma DL, Pathak RP, Manral K, Tandon AK and Manju Joshi. Indian Drugs. 1985; **23(3)**: 126-128.
- Khetwal KS and Verma DL. Fitoterapia. 1986; **LVII(2)**: 128.
- Khetwal KS and Verma DL. Indian Drugs. 1990; **28(2)**: 99-100.
- Pero, R. W. and Owens, R. G., (1971) App. Microbiology, **21**, 546.
- Mabry, T. J, Markham, K. R. and Thomas, M. B., (1970) The systematic identification of the flavonoids, Springer, New-York.
- Markham, K. R. and Porter, L. J., (1978) Chemical Constituents of Bryophytes, Prog. in Phytochemistry, **5**, 181-272
- Markham, K. R., and Porter, L. J. (1974) Flavonoids of the liverwort Marchantia polymorph, Phytochemistry, **13**, 1937-1942.
- Markham, K. R. and Porter, L. J. (1975) Flavone glucuronoids of New Zealand liverwort Marchantia Marcopora, Phytochemistry, **14**, 1093-1097.
- Markham, K. R. Porter, L. J. and Campbell, E. O. (1977) The usefulness of flavonoid characters in studies of the taxonomical phylogeny of liverworts, Bryophytorum, Bibliotheca, **13**, 378-398.
- Mishra R. Chemical investigation of some ferns of Kumaun Hills, Ph. D. Thesis, Kumaun University, Naini Tal, 2008.
- Mishra R. and Verma DL. Nature and Science. 2009a; **7(6)**: 82-85.
- Mishra R. and Verma DL. New York Science Journal. 2009b; **2(5)**: 93-95.
- Mishra R. and Verma DL. J. American Science 2009c; **5(4)**: 183-188.
- Mishra R. and Verma DL. (to be published).
- Mishra R. and Verma DL. (to be published).
- Schier, W. (1974) Untersuchungen zur chemotaxonomic der Marchantiales, Nova Hedwigia, **25**, 249-266.
- Tewari, S. D., Ph. D. thesis, Kumaun University, Naini Tal (India), 1984.

5/11/2009